## सी डी एफ डी CDFD

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अधिदेश Mandate

## अधिदेश

सीडीएफडी सोसाइटी के संगम ज्ञापन तथा नियम एवं विनियमों में बताए गए के अनुसार डीएनए फिंगरप्रिंटिंग एवं निदान केन्द्र की स्थापना जिन उद्देश्यों के लिए हुई वे निम्न प्रकार हैं :
i. पितृत्व विवाद, आप्रवास और अस्पतालों में नवजात शिशुओं की अदला-बदली जैसे मामलों में निजी पक्षों सहित विविध अभिकरणों के लिए पर्याप्त अदायगी पर डीएनए प्रोफाइलिंग और उससे संबंधित विश्लेषण का वैज़ानिक अनुसंधान करना।
ii. अपराध अन्वेषण अभिकरणों को डीएनए फिंगरप्रिंटिंग और उससे संबंधित विश्लेषण तथा सुविधाएँ प्रदान करना।
iii. अपराध अन्वेषण और परिवार मामलों में डीएनए प्रोफाइल विश्लेषण और उससे संबंधित तकनीकों के साक्ष्य संबंधी मूल्य को समझने में पुलिस कर्मियों, न्यायिक वैज्ञानिकों, वकीलों तथा न्यायपालिका की सहायता करना।
iv. आनुवंशिक अव्यवस्थाओं को संसूचित करने हेतु डीएनए नैदानिक विधियाँ सिद्ध करना और इस प्रकार के संसूचन के लिए संपरीक्षाएँ विकसित करना।
v. पादप और जंतु कोशिका माल, कोशिका लाइनों के प्रामाणीकरण के लिए डीएनए फिंगरप्रिंटिंग तकनीकों का उपयोग करना और ऐसे प्रयोजनों के लिए आवश्यकतानुसार नईे संपरीक्षाएँ विकसित करना।
vi. डीएनए फिंगरप्रिंटिंग तकनीकों पर प्रशिक्षण प्रदान करना।
vii. मूलभूत, अनुप्रयुक्त अनुसंधान एवं विकास कार्य करना।
viii. देश में चिकित्सा संस्थाओं, जन-स्वास्थ्य अभिकरणों और उद्योग को परामर्शी सेवाएँ प्रदान करना।
ix. केंद्र के उद्देश्यों से संगत क्षेत्रों में विदेशी अनुसंधान संस्थाओं एवं प्रयोगशालाओं और अन्य अंतर्राष्ट्रीय संगठनों के साथ सहयोग करना।
x. अनुसंधान छात्रों को स्नातकोत्तर उपाधियों के लिए पंजीकृत कर सकने के प्रयोजन हेतु उच्चतर अधिगम के मान्यता प्राप्त विश्वविद्यालयों एवं संस्थाओं के साथ संबंधन स्थापित करना।
xi. भारत सरकार, राज्य सरकारों, देश में स्थित पूर्त संस्थाओं/न्यासों, व्यक्तियों और उद्योग से नकद के रूप में या अन्य रूपों में अनुदान, दान एवं अंशदान प्राप्त करना।
xii. केंद्र सरकार के पूर्व अनुमोदन से प्रशिक्षण कार्यक्रमों, वैज्ञानिक अनुसंधान और अन्य गतिविधियों के लिए अंतर्राष्ट्रीय संगठनों सहित विदेशी स्रोतों से आर्थिक सहायता प्राप्त करना।
xii. केंद्र की गतिविधियों को चलाने के लिए जैसा आवश्यक या सुविधाजनक हो, कोई भी संपत्ति चल या अचल या भवनों एवं निर्माणों को निर्मित करने, सुधार करने, परिवर्तित करने, गिरा देने या मरम्मत करने हेतु उपहार, क्रय, विनिमय, पट्टा, भाड़े पर लेने द्वारा या अन्यथा किसी भी तरह अर्जित करना।
xiv. केंद्र के प्रयोजन हेतु, भारत सरकार और अन्य प्रोनोटों, विनिमय पत्रों, चैक या अन्य परक्राम्य लिखतों को आहरित करना और स्वीकार करना, तैयार करना और पृष्ठांकित करना, बट्टा करना और परक्रामण करना।
xv. केंद्र को सौंपी गई निधियों या धन को निवेश करने के लिए, ऐसी प्रतिभूतियों को खोलने या ऐसे तरीके से, जो कि समय-समय पर शासी परिषद् द्वारा निर्धारित किए जाते हैं, इस प्रकार के निवेश को विक्रय या पक्षांतरण करना।
xvi. उक्त सभी उद्देश्यों या उनमें से किसी उद्देश्य की प्राप्ति के लिए सभी ऐसे अन्य विधिसम्मत कार्य, जैसा आवश्यक, प्रासंगिक या सहायक हो, करना।
xvii. केंद्र के उद्देश्यों को वास्तविक बनाने के लिए प्रोफेसरों अन्य संकाय पदों, अभ्यागत अध्येतावृत्तियों सहित अध्येतावृत्तियों, अनुसंधान एवं संवर्ग पदों, छात्रवृत्तियों आदि को संस्थापित करना।
xviii. केंद्र के वैज़ानिक एवं प्रौद्योगिकीय कार्य के लिए प्रयोगशालाओं, कार्यशालाओं, भंडार, पुस्तकालय, कार्यालय और अन्य सुविधाओं को स्थापित करना।
xix. तकनीकी जानकारी को उद्यमकर्ताओं और उद्योगों से प्राप्त या उनको अंतरण करना; और
xx पेटेंटों, डिज़ाइनों एवं तकनीकी जानकारी जो कि केंद्र द्वारा विकसित की गई हो, को पंजीकृत करना और केंद्र के हित में ऐसे पेटेंटो/डिज़ाइनों/तकनीकी जानकारी के किसी भाग को अंतरण करना।

## MANDATE

The objectives for which the Centre for DNA Fingerprinting and Diagnostics (CDFD) was established, as enumerated in Memorandum of Association and Rules and Regulations of CDFD Society, are as follows:
i. To carry out scientific research pertaining to DNA profiling and related analysis in civil cases like paternity disputes, immigration, and exchange of newborns in hospitals, for various agencies including private parties, on appropriate payment;
ii. To provide DNA fingerprinting and related analysis and facilities to crime investigation agencies;
iii. To assist police personnel, forensic scientists, lawyers and the judiciary in understanding the evidential value of the DNA profile analysis and related techniques in crime investigation and family matters;
iv. To establish DNA diagnostic methods for detecting genetic disorders and to develop probes for such detection;
v. To use DNA fingerprinting techniques for the authentication of plant and animal cell material, cell lines and to develop new probes where necessary for such purposes;
vi. To provide training in DNA fingerprinting techniques;
vii. To undertake basic, applied and developmental R \& D work;
viii. To provide consultancy services to medical institutions, public health agencies and industry in the country;
ix. To collaborate with foreign research institutions and laboratories and other international organizations in fields relevant to the objectives of the Centre;
x. To establish affiliation with recognized universities and institutions of higher learning for the purpose of enabling research scholars to register for post-graduate degrees;
xi. To receive grants, donations and contributions in cash or in other forms from the Government of India, State Governments, Charitable Institutions/Trusts, individuals and industry within the country;
xii. To receive, with the prior approval of the Central Government, monetary assistance from foreign sources including international organizations for training programmes, scientific research and other activities;
xiii. To acquire by gift, purchase, exchange, lease, hire or otherwise howsoever, any property movable or immovable or to construct, improve, alter, demolish or repair buildings and structures as may be necessary or convenient for carrying on the activities of the Centre;
xiv. For the purpose of the Centre, to draw and accept, make and endorse, discount and negotiate Government of India and other Promissory Notes, Bills of Exchange, Cheques or other Negotiable Instruments;
$x v$. For investing the funds of or money entrusted to the Centre, to open such securities or in such manner as may from time to time be determined by the Governing Council and to sell or transpose such investment;
xvi. To do all such other lawful acts as may be necessary, incidental or conducive to the attainment of all or any of the above objectives;
xvii. To institute Professorships, other faculty positions, fellowships including visiting fellowships, research and cadre positions, scholarships, etc. for realizing the objectives of the Centre;
xviii. To establish, maintain and manage laboratories, workshops, stores, library, office and other facilities for scientific and technological work of the Centre;
xix. To acquire or transfer technical know-how from/to entrepreneurs and industries; and
xx. To register patents, designs \& technical know-how that may be developed by the Centre and transfer any portion of such patents/designs/technical know-how in the interest of the Centre.

निदेशक का संदेश
From the Director's Desk

## निदेश्रक का संदेश

डीएनए फिंगरप्रिंटिंग एवं निदान केंद्र (सीडीएफडी) की स्थापना 1996 में भारत सरकार के जैव प्रौद्योगिकी विभाग (डीबीटी) के स्वायत्त संस्थान के रूप में की गई। यह देश में आण्विक जीवविज्ञान के विविध क्षेत्रों में आधुनिकतम अंतरविषयक अनुसंधान और सेवा प्रदान करने के अधिदेश के साथ बनाए गए अग्रणी अनुसंधान संस्थानों में से एक है। पिछले वर्षों में केंद्र में सभी क्षेत्रों की गतिविधियों में उल्लेखनीय वृद्धि देखी गई है और अन्वेषण के अनेक नए क्षेत्र आरंभ किए गए हैं। वर्ष 2012-13 के दौरान सीडीएफडी की उपलब्धियों और प्रगति के विवरण इस प्रगति रिपोर्ट में आगे बताए गए हैं और मैं नीचे उनमें से कुछ पर प्रकाश डालना चाहता हूं।

इस प्रतिवेदनाधीन अवधि के दौरान, डीएनए फिंगरप्रिंटिंग सेवा प्रयोगशाला में केंद्र और विभिन्न राज्य सरकारों की न्याय पालिका तथा कानून प्रवर्तन एजेंसियों द्वारा लगभग 175 मामले अग्रेषित किए गए थे। देश में डीएनए प्रोफाइलिंग सेवाओं के विस्तार के प्रति अपने प्रयासों में सीडीएफडी ने उड़ीसा और आंध्र प्रदेश की राज्य सरकारों के साथ किसी लागत के बिना डीएनए प्रोफाइलिंग सेवाएं प्रदान करने के लिए एक समझौता ज्ञापन किया है, जिसके लिए हमारा विश्वास है कि यह न्याय प्रदायगी प्रणाली में एक प्रौद्योगिकी प्रदर्शक के रूप में कार्य करेगा। उड़ीसा में समझौता ज्ञापन व्यवस्था के भाग के रूप में सीडीएफडी ने जीव विज्ञान संस्थान (आईएलएस), भुवनेश्वर के साथ मिलकर आईएलएस में डीएनए प्रोफाइलिंग की एक प्रयोगशाला स्थापित की है। नैदानिक सेवाओं के क्षेत्र में निजाम इंस्टीटचूट अॅफ मे डिकल साइं सिस (एनआईएमएस), हैदराबाद में चिकित्सा आनुवंशिकी विभाग की स्थापना की गई जो सफलता पूर्वक कार्यरत है और यहां लगभग 3500 रोगियों का आनुवंशिक मूल्यांकन तथा परामर्श देने का कार्य किया गया है। इस वर्ष नैदानिक प्रभाग में कार्यरत मेरे सहयोगियों ने एक्सोम सीक्वेंसिंग के माध्यम से जीन जीजेसीट में एक सजाति


परिवार का नवीन उत्परिवर्तन अभिज्ञात किया है और बौद्धिक विकलांगता तथा दौरे के लक्षणों वाली एक महिला रोगी में -ARHGEF9 के मानचित्रण का कार्य भी सफलता पूर्वक पूरा किया है। इस वर्ष, एपीडा-सीडीएफडी बासमती डीएनए विश्लेषण केंद्र में 200 बासमती चावल के नमूनों का परीक्षण उनकी शुद्धता के लिए किया गया। आण्विक आनुवंशिकी की प्रयोगशाला में रेशम कीट एमएमएनपीवी अंत:क्रियाओं के अध्ययन में बीएमएनपीवी वायरस से उत्पन्न माइक्रो आरएनए को पहचानने में सफलता मिली है जो डीएनए बंधन प्रोटीन सहित वायरल लेट जीन (सिस लक्ष्य) की अभिव्यक्ति का नियमन करता है और बॉम्बिक्स मोरी में वायरस की लंबित अभिव्यक्ति के लिए महत्वपूर्ण है। एंथ्रिया आसमा की आनुवंशिक विविधता और आबादी की संरचना में इस समूह का अध्ययन टसर रेशम कीट के विशिष्ट इकोटाइप के परिरक्षण और असम क्षेत्र में सीमित मूंगा रेशम कीट की घटती आबादी के संरक्षण में उपयोगी रहा है। इसके अलावा, इस प्रयोगशाला के अनुसंधान कर्ताओं ने वाणिज्यिक रेशम कोकून गुणों वाले परजीनी रेशम कीट सफलता पूर्वक तैयार किए हैं जो बैकुलोवायरस प्रतिरोधक हैं और ये परजीनी लाइनें वर्तमान में बहु स्थल क्षेत्र परीक्षणों के अनुमोदन की प्रतीक्षा में हैं।

कवकी रोगजनन प्रयोगशाला के अनुसंधान कर्ताओं ने टीएचपी- 1 मानव मोनोसायटिक कोशिकाओं से उत्पन्न मैक्रोफेज के साथ कैंडिडा ग्लाब्राटा के बीच अंत: क्रिया को समझा है, जिससे प्रकट होता है कि सी. ग्लाब्राटा कोशिकाओं में फेगोलाइसोसोमल परिपक्वता की रोकथाम की क्षमता होती है जो अभिक्रियात्मक ऑक्सीजन प्रजाति (आरओएस) के उच्च स्तर के साथ जीवित रह सके और पोषी कोशिकाओं की प्रतिक्रिया में आईएल - 4 स्राव आरंभ कर सकें। जीवाण्विक कोशिका जीव विज्ञान प्रयोगशाला द्वारा निर्धारण किया गया है कि माइको बैक्टीरियम ट्यूबरकुलोसिस (एमटीबी) प्रोटीन जैसे एमटीबी एचएसपी 60 के कोशिकीय स्थानीकरण के साथ मैक्रोफेज में टोल के समान ग्राहियों से अंत: क्रिया पर मैक्रोफेज की कार्यात्मकता पर प्रभाव हो सकता है और पोषी में टी - कोशिका प्रतिरक्षी प्रत्युत्तर के ध्रुवी करण से एमटीबी रोग जनकता प्रभावित हो सकती है। इस समूह ने चूहा मॉडल पर कार्य करते हुए एमटीबी की उत्तरजीविता और प्रोटीन पीपीई 18 की महत्वपूर्ण भूमिका इनवीवो दर्शाई है।

पादप सूक्ष्मजीवों की अंतः क्रिया पर हमारे अध्ययनों से XadM के लाक्षणीकरण में सहायता मिली है, जो चावल के जीवाणु रोगाणु जैंथोमोनाज़ ओरीजी पीवी ओरीजी में जैंयोमोनाज़ का एक नया आसंजन प्रोटीन है और शीष्र संलग्नता, कॉलोनी निर्माण और बायोफिल्म निर्माण में महत्वपूर्ण भूमिका निभाता है।

आण्विक अर्बुद शास्त्र प्रयोगशाला के अध्ययनों में प्रकट हुआ है कि एआरआईडी।बी, जो एसडब्ल्यूआई/एसएनएफ क्रोमेटिन रिमॉडलिंग कॉम्प्लैक्स के एक घटक की इनकोडिंग करता है, पेनक्रियाज़ के कैंसर के लिए एक नया ट्यूमर संदमक जीन है और विशिष्ट पीएएच उत्परिवर्तन से भारत में फेनिल कीटोनूरिया पैदा करता है। स्तनी आनुवंशिकी प्रयोगशाला में किए गए अनुसंधान से कैंसर जनन और विकास में डीएनए मेथिल ट्रांसफरेज़ डीएनएमटी3l और डीएनएमटी2 की भूमिका का पता चला है। इस समूह ने एमटीबी से चुनौती देने पर पोषी

कोशिका में होने वाली एपिजेनेटिक बदलावों को भी अभिज्ञात किया है।

कोशिका संकेतन प्रयोगशाला के कार्य में प्रदश्शित किया गया है कि फॉस्फा इनोसिटॉल यौगिक आईपी7 द्वारा आरएनए पॉलीमरेज 1 के पाइरोफॉस्फोराइलेटिंग घटकों द्वारा ईस्ट में राइबोसोम जैव जनन का नियमन किया जाता है, और इस प्रकार आरआरएनए संश्लेषण का नियंत्रण किया जाता है। यह भी दर्शाया गया है कि इनोसिटोल पायरोफॉस्फेट का आईपी6के 1 द्वारा संश्लेषण स्तनधारी कोशिकाओं में समजात पुन: संयोजन माध्यित डीएनए मरम्मत के पूरे होने में एक भूमिका निभाता है। अनुलेखन प्रयोगशाला एसेरिशिया कोलाई में कारक आश्रित अनुलेखन समापन और समापन रोधी गतिविधि के आण्विक आधार को समझने में सक्रिय रूप से संलग्न है। जीवाण्विक आनुवंशिकी प्रयोगशाला में कारक आश्रित अनुलेखन समापन और डीएनए-आरएनए हाइब्रिड (आर-लूप्स) के निर्माण में इससे बचने में की भूमिका का अध्ययन भी किया गया है, जो अनुलेखन-द्विगुणन विवादों को बढ़ावा देता है। इस समूह ने विलेय परिवहन और नियमन तथा अलारमोन पीपी जीपीपी के अनेक कार्यों की खोज ई. कोलाई में भी की है।

अभिकलनात्मक जीव विज्ञान प्रयोगशाला में मानव वायरल सेतु प्रोटीन-प्रोटीन अंत: क्रिया नेटवर्कों के अध्ययन में दर्शाया गया है कि कुछ वायरल प्रोटीन आरटीकुलेशन बिन्दुओं के रूप में इनके बीच अन्यथा बाइकनेक्टिड मानव प्रोटीन अंतः क्रिया नेटवर्क सेतु का कार्य करते हैं। ये संयोजन मानव प्रोटीन-प्रोटीन अंत: क्रियाओं पर प्रभाव डालने के लिए वायरसों द्वारा प्रयुक्त प्रक्रियाओं को समझने में महत्वपूर्ण हैं।

तुलनात्मक रूप से केंद्र के नए संकाय सदस्यों ने विचारोत्तजक कार्य किए हैं, जो चुनौती पूर्ण है इनमें कोशिका चक्र नियमन, कोशिका मृत्यु और कोशिका उत्तर जीविता मार्ग तथा ड्रोसोफिला तंत्रिका विकास अध्ययन के क्षेत्र रहे। इसके अलावा, डॉ. देवयानी हल्दर ने हाल ही में केंद्र का

कार्य भार संभाला है और क्रोमेटिन जीव विज्ञान तथा एपिजेनेटिक्स के क्षेत्र में कार्य आरंभ किया है। मुझे विश्वास है कि हमारे युवा सहकर्मियों द्वारा आरंभ की गई इन वैज्ञानिक यात्राओं से आने वाले वर्षों में अपार लाभ मिलेंगे। मुझे यह बताते हुए खुशी है कि हेल्डन पीठ के रूप में डॉ. डी पी कस्बेकर के आने से 'टीम सीडीएफडी' बहुत सशक्त बन गई है। न्यूरोस्पोरा आनुवंशिकी के क्षेत्र में डॉ. कस्बेकर के अग्रणी कार्य के साथ उनका संकाय और अनुसंधान वेत्ताओं के प्रति मार्गदर्शन कौशल से सीडीएफडी को अनेक प्रकार से लाभ मिलेगा। यह भी हर्ष का विषय है कि प्रतिवेदनाधीन अवधि के दौरान, नेशनल इंस्टीट्यूट ऑफ एग्रोबायोलॉजिकल साइंसिज़ (एनआईएएस), सुकुबा, जापान के प्रो. काजुई मिता, जो रेशम कीट आनुवंशिकी में अंतरराष्ट्रीय ख्याति प्राप्त व्यक्ति हैं, ने केंद्र का आमंत्रण स्वीकार किया और असाधारण विदेशी वैज्ञानिकों के लिए जैव प्रौद्योगिकी पीठ हेतु डीबीटी योजना की शर्तों के अनुसार अतिथि प्रोफेसर के रूप में यहां लगभग एक वर्ष का समय बिताया।

स्मरणीय है कि दिसंबर 2009 में सीडीएफडी तथा आईकेपी ज्ञान पार्क, हैदराबाद ने मिलकर कुछ विकसित प्रौद्योगिकियों के वाणिज्यीकरण के लिए एक महत्वकांक्षी कार्यक्रम आरंभ किया और इसे सीडीएफडी द्वारा पेटेंट कराया गया। इस कार्यक्रम के परिणाम मिलने आरंभ हो गए हैं और केंद्र में ई. कोलाई का उपयोग करते हुए एल-आर्जीनीन उत्पादन हेतु सूक्ष्म जीव प्रक्रम से संबंधित प्रौद्योगिकी को बायोनरी बाइप्रोडक्ट्स प्रा. लि. को लाइसेंस दिया गया है, जिसे डीबीटी - एसबीआईआरआई योजना से निधिकरण समर्थन प्राप्त है।

विमटा लैब्स लिमिटेड में सीडीएफडी की प्रयोगशाला जंतु सुविधा, शमीरपेट (नामपल्ली में वर्तमान परिसर से लगभग 45 किलोमीटर की दूर पर) में पूरी तरह कार्यशील है। सीडीएफडी के अनेक अनुसंधान कर्ताओं ने इस सुविधा में परियोजनाएं आरंभ की हैं, जहां वर्तमान में आईपी 6 के, एनएनएटी, सी 57 बीएल $/ 6$, एफओएक्सएन 1 एनयू और बाल बी / सी सहित चूहों के पांच आंतरिक रूप से प्रजनन

करने वाले विभेद उपलब्ध हैं।
इस वर्ष भी सीडीएफडी के अनेक संकाय सदस्यों और अध्येताओं को प्रतिष्ठित पुरस्कार और सम्मान प्राप्त हुए हैं। इनमें शामिल हैं सीनियर इनोवेटिव यंग बायोटेक्नोलॉजिस्ट एवॉर्ड, आईसीएमआर क्षणिक ओरेशन एवॉर्ड, फेलोशिप ऑफ द इण्डियन एकेडमी, बैंगलोर, और पद्म श्री पुरस्कार। इस प्रतिवेदनाधीन अवधि के दौरान पांच अनुसंधानकर्ताओं को पीएचडी डिग्री प्रदान की गई है। सीडीएफडी द्वारा निरंतर प्रतिभाशाली, युवा और सक्रिय पीएचडी अध्येताओं, पोस्टडॉक्टरल अध्येताओं, परियोजना सहयोगियों और ग्रीष्मकालीन प्रशिक्षुओं को आकर्षित किया जाता है।

मैंने पिछले वर्ष बताया था कि सीडीएफडी उप्पल, हैदराबाद के भारतीय सर्वेक्षण विभाग की भूमि पर सरकार से वित्तीय अनुमोदन मिलने के पश्चात् स्थायी परिसर निर्मित करने का इच्छुक है। वर्तमान स्थिति यह है कि वास्तुकारों, परियोजना प्रबंधन परामर्श दाताओं और सिविल कार्य के लिए संविदाकारों का चयन पूरा हो चुका है। पुनः, भारतीय विमान पत्तन प्राधिकरण, आ. प्र. प्रदूषण नियंत्रण बोर्ड, हैदराबाद महानगर जल आपूर्ति और सीवेज बोर्ड, अग्निशमन सेवा और आ. प्र. राज्य आपदा प्रतिक्रिया सहित विभिन्न एजेंसियों से सांविधिक अनुमोदन लिए गए हैं। हमें आशा है कि भूमि कार्य जल्दी ही आरंभ होगा, ताकि केंद्र में संकाय और अन्य कार्यकर्ताओं के सामने आने वाली स्थान और मूल संरचना की कमी को जल्दी से जल्दी दूर किया जा सके।
मैं शासी परिषद, अनुसंधान क्षेत्र पैनल-वैज्ञानिक सलाहकार समिति, शैक्षिक परिषद, वित्त समिति, भवन निर्माण समिति और डीबीटी से सभी गतिविधियों के लिए प्राप्त अपार सहयोग के प्रति अपना आभार व्यक्त करता हूँ। मैं सभी सदस्यों और अधिकारियों को धन्यवाद देता हूँ , जिन्होंने हमारी गतिविधियों तथा उपलब्धियों में अपना समय और प्रयास का योगदान दिया है।

केवल इतना ही नहीं, उपरोक्त बताई गई सफलताओं के साथ मैं सीडीएफडी परिवार के सभी सदस्यों के प्रति

अपना हार्दिक आभार व्यक्त करता हूँ, मैं एक अत्यंत दुखद समाचार के साथ अपना सदेश समाप्त करने के लिए बाध्य हूँ । वर्ष 2012 के अंतिम दिन हमारे महान संकाय सदस्य और वरिष्ठ सहकर्मी, डॉ. जे नागाराजू अल्पकालीन बीमारी के बाद चल बसे। डॉ. नागराजू ने 1998 में सीडीएफडी में कार्यभार संभाला, जो आण्विक आनुवंशिकी प्रयोगशाला के प्रमुख और डीएनए फिंगरप्रिंटिंग सेवा प्रयोगशाला के समन्वयक थे। उन्हें 2011 में केंद्र की खुराना पीठ पर भी नियुक्त किया गया था। वे आनुवंशिकी के क्षेत्र में एक अंतरराष्ट्रीय रूप से जानी मानी हस्ती वाले एक वैज्ञानिक ही नहीं बल्कि रेशम कीट जीव विज्ञान और

बासमती चावल में विशेष रुचि रखने के अलावा इस संस्थान में बौद्धिक और प्रशासनिक क्षमताओं के आधार थे और अनेक लोगों के सलाहकार तथा मार्गदर्शक थे। इन सब से परे वे एक नजदीकी व्यक्तिगत मित्र थे। उनकी स्मृति में सबसे अच्छी श्रृद्धांजली यही होगी कि केंद्र उन ऊंचाइयों को पाने के लिए और भी कठिन प्रयास करे, जिनके लिए डॉ. नागाराजू ने इतने जोश के साथ अपने स्वयं के अनुसंधान प्रयास आगे बढ़ाए।

ज गौरीशंकर
31 मार्च, 2013


## Director's Message

The Centre for DNA Fingerprinting and Diagnostics (CDFD), established in 1996 as an autonomous institute of the Department of Biotechnology (DBT) of the Government of India, is one amongst the pioneering research institutions in the country with the mandate to provide services and to undertake cutting edge interdisciplinary research in diverse areas of molecular biology. Over the years, the Centre has witnessed significant growth in all phases of activities, and several new frontiers of investigation have been initiated. The achievements and progress of CDFD's research during 2012-2013 are described in the chapters that follow in this progress report, and I would like to highlight a few of them below.

During this reporting period, the Laboratory of DNA Fingerprinting Services was forwarded about 175 cases by the judiciary and law enforcing agencies of the Union and different State Governments. In its efforts towards expansion of DNA profiling services in the country, the CDFD has entered into MoU's with the State Governments of Orissa and Andhra Pradesh to provide DNA profiling services at no cost basis, which we believe would serve as a technology demonstrator for the justice delivery system. As part of the MoU arrangement in Orissa, the CDFD has joined hands with the Institute of Life Sciences (ILS), Bhubaneshwar, to establish a DNA Profiling Laboratory at the ILS. In the area of Diagnostic services, the Medical Genetics Department established at the Nizam's Institute of Medical Sciences (NIMS), Hyderabad has been running successfully and around 3500 patients underwent genetic evaluation and counselling. My colleagues working in the Diagnostics division have identified through exome sequencing, a novel mutation in a consanguineous family in a gene GJC2, and have also successfully carried out mapping of ARHGEF9 in a female patient with symptoms of intellectual disability and seizures. The APEDA-CDFD Centre for Basmati DNA Analysis tested close to 200 basmati rice samples for their purity during this year.

The Laboratory of Molecular Genetics, while studying silkworm-MmNPV interactions, has been successful in identifying BmNPV virus-derived microRNA that regulates the expression of viral late genes (cis targets) including DNA a binding protein which is important for the late expression of the virus in Bombyx mori. This group's study of the

genetic diversity and population structure of Antheraea assama has been useful for preservation of unique ecotypes of tasar silkmoths and conservation of declining populations of muga silkmoths confined to the Assam region. In addition, the researchers in this laboratory have successfully generated transgenic silkworms resistant to baculovirus with commercial silk cocoon properties and these transgenic lines are presently awaiting approval to be taken for multi-location field trials.

Researchers in the Laboratory of Fungal Pathogenesis have deciphered the interaction between Candida glabrata with macrophages derived from THP-1 human monocytic cells, revealing that $C$. glabrata cells possess ability to prevent phagolysosomal maturation, to survive high levels of reactive oxygen species (ROS), and to invoke IL-4 secretion response in the host cells. The Laboratory of Molecular Cell Biology has determined that cellular localization of Mycobacterium tuberculosis (Mtb) proteins such as Mtbhsp60, upon interaction with Toll-like receptors in macrophages, can influence macrophage functions; and that the polarization of T-cell immune responses can affect Mtb virulence in the host. This group, while working on a mouse model, has demonstrated an important role of protein PPE18 in replication and survival of Mtb in vivo.
Our studies on plant-microbe interactions have helped in the characterization of XadM, which is a novel adhesion protein that plays an important role in early attachment, colonization and biofilm formation of the rice bacterial pathogen Xanthomonas oryzae pv. oryzae.

Studies in the Laboratory of Molecular Oncology have revealed that ARID1B, which encodes a component of the SWI/SNF chromatin remodelling complex, is a novel tumour suppressor gene for pancreatic cancers and that unique $P A H$ mutations cause phenylketonuria in India. Research in the Laboratory of Mammalian Genetics has dissected out the role of DNA methyltransferases Dnmt3l and Dnmt2 in carcinogenesis and development. This group has also identified epigenetic changes that the host cell undergoes when challenged with Mtb.
Work in the Laboratory of Cell Signalling has demonstrated that the phospho-inositol compound $\mathrm{IP}_{7}$ regulates ribosome biogenesis in yeast by pyrophosphorylating components of RNA polymerase I, thereby controlling rRNA synthesis. It has also been shown that inositol pyrophosphates synthesised by IP6K1 play a role in the completion of homologous recombinationmediated DNA repair in mammalian cells.
The Laboratory of Transcription has been actively engaged in understanding the molecular basis of factor-dependent transcription termination and antitermination in Escherichia coli. The Laboratory of Bacterial Genetics is also studying factordependent transcription termination and its role in avoidance of formation of RNA-DNA hybrids (Rloops) that provoke transcription-replication conflicts. This group has also investigated the mechanisms of solute transport and regulation, as well as the multiple functions of the alarmone ppGpp in E. coli.
The Laboratory of Computational Biology, while studying human viral bridge protein-protein interaction networks, has shown that some of the viral proteins act as articulation points to bridge otherwise unconnected nodes in the biconnected human protein interaction network. These connections may hold keys to understand the mechanisms used by viruses for hijacking human protein-protein interactions.
The Centre's comparatively new faculty members have embarked upon exciting, yet challenging, studies in areas such as cell cycle regulation, cell death and cell survival pathways, and Drosophila neural development. In addition, Dr Devyani Haldar joined the Centre very recently and is to initiate work in the area of Chromatin Biology and Epigenetics. I am confident that these scientific journeys which have been initiated by our young colleagues would yield immense dividends in the years to come.

I am excited to report that 'Team CDFD' has been strengthened by the joining of Dr DP Kasbekar as Haldane Chair. Dr Kasbekar's pioneering research in the area of Neurospora genetics, together with his cosmic skills in mentoring faculty and research scholars, will enrich the CDFD in numerous ways. It is a pleasure also to mention that during the period under report, Prof. Kazuei Mita, an internationally renowned authority in silkmoth genomics from the National Institute of Agrobiological Sciences (NIAS), Tsukuba, Japan, accepted the Centre's invitation and spent almost a year here as Visiting Professor, in line with terms of the DBT scheme for Biotechnology Chair for outstanding overseas scientists.

It may be recalled that in December 2009, the CDFD and IKP-Knowledge Park, Hyderabad had initiated an ambitious programme for development and commercialization of certain technologies developed and patented by CDFD. This programme has started bearing fruit, and the Centre's technology related to microbial process for Larginine production using $E$. coli strains has been licensed to M/s Bionary Bioproducts Pvt. Ltd. with funding support secured from the DBT-SBIRI scheme.

CDFD's Laboratory Animal Facility at M/s Vimta Labs in Shamirpet ( $\sim 45 \mathrm{kms}$ from the current campus at Nampally) is fully operational. Several CDFD researchers have initiated projects in this facility which presently houses five inbred mouse strains including Ip6k1, Nnat, C57BL/6, Foxn1 ${ }^{\text {nu }}$ and Balb/c.

This year too, several of the CDFD faculty and scholars have been recipients of prestigious awards and honours. These include the Senior Innovative Young Biotechnologist Award; ICMR Kshanika Oration Award; Fellowship of the Indian Academy of Sciences, Bangalore; and the Padma Shri award. During this reporting period, five research scholars were conferred with PhD degrees. CDFD continues to attract bright, young and dynamic PhD scholars, postdoctoral fellows, project associates and summer trainees into its midst.

I had reported last year on the intention to construct CDFD's permanent campus on land of the Survey of India at Uppal, Hyderabad, subject to final financial approvals being received from the Government. The current status is that the selection of architects, project management consultants and contractors for civil work has been completed. Further, statutory approvals of various agencies,
including the Airports Authority of India, A.P. Pollution Control Board, Hyderabad Metropolitan Water Supply and Sewerage Board, Fire Services and A.P. State Disaster Response have been taken. We do hope that the ground work would commence shortly so that the space and infrastructure constraints presently being faced by the faculty and other workers in the Centre are alleviated as soon as possible.

I take this opportunity also to acknowledge the unstinted co-operation which the Centre has received all along for its activities from the Governing Council, Research Area PanelsScientific Advisory Committee, Academic / Finance / Building Committees and, of course, the DBT. I wish to thank all the members and officials for their time and effort in supporting our activities and achievements.

Notwithstanding the successes reported above, for which I extend sincere gratitude to all members of the CDFD family, I am constrained to close this
message on an extremely sad note. On last day of the calendar year 2012, our illustrious faculty member and senior colleague Dr J Nagaraju passed away after a brief illness. Dr Nagaraju had joined the CDFD in 1998, was Head of the Laboratory of Molecular Genetics, and Co-ordinator of the Laboratory of DNA Fingerprinting Services; he was also appointed to the Khorana Chair of the Centre in 2011. He was not just a scientist of international renown in the area of genetics with particular reference to silkmoth biology and basmati rice, but also the backbone of this Institute in both intellectual and administrative capacities and a mentor and guiding light for many. Above all, he was a close personal friend. There can be no more fitting tribute to his memory than for the Centre to strive even harder to achieve the heights that Dr Nagaraju had so passionately cared for in his own research endeavours.

J Gowrishankar
March 31, 2013


## सेवाएँ <br> Services

# LABORATORY OF DNA FINGERPRINTING SERVICES 

| Faculty | Madhusudan Reddy Nandi |
| :---: | :---: |
| Other Members | SPR Prasad <br> Ch V Goud <br> Devinder Kumar <br> Sanjukta Mukerjee <br> **DS Negi <br> Girnar Vijay Amrutrao <br> **Chandra Shekhar Singh <br> Shruti Das Gupta |
| Coordinator | *J Nagaraju DP Kasbekar |
| * deceased 31 Dec. 2012 <br> ** presently posted at DNA Profiling Laboratory of CD <br> Bhubaneswar, Odisha |  |
| Objectives |  |
| 1. To provide DNA fingerprinting services in cases forwarded by law-enforcing agencies and judiciary of State and Federal Governments, relating to murder, sexual assault (rape), paternity, maternity, child swapping, body identification, kidney transplantation, etc.; |  |

2. To develop human resources skilled in DNA fingerprinting, to cater to the needs of State and Federal Government agencies;
3. To impart periodical training to manpower involved in DNA fingerprinting sponsored by State and Federal Government agencies;
4. To provide advisory services to State and Federal Government agencies in establishing DNA Fingerprinting facility; and
5. To create DNA marker databases of different caste populations of India.

Summary of services provided until the beginning of this reporting year (April 1, 2011 - March 31, 2012)

A total number of 94 cases were received for DNA fingerprinting examination during this reporting period. Of these, 50 cases related to paternity/ maternity, 33 cases related to identification of deceased, 5 cases were pertaining to sexual assault (rape), 3 cases were related to murder and 3 cases pertaining to biological relationship (kidney transplantation). Seventeen States and Union Territories of India have availed DNA fingerprinting services of CDFD during this period. Andhra Pradesh forwarded the highest number of cases (33) followed
by Chhattisgarh (12), Karnataka (10), Punjab (10), Kerala (6), Maharashtra (3), Uttar Pradesh (4), Bihar (3), Delhi (3), Jammu \& Kashmir (2), Jharkhand (1), Madhya Pradesh (1), Orissa (1), Puducherry (1), Tamil Nadu (1) and Uttarakhand (1).

Details of services provided in the current reporting year (April 1, 2012 to March31, 2013)

Breakup of the cases during this reporting period is given below under following heads:
Biological relationship 05
Murder 13
Sexual assault (Rape) 19
Maternity/paternity 70
Identity of the deceased individuals 79
Total number of cases $\overline{\mathbf{1 8 6}}$
A total number of 186 cases were received for DNA fingerprinting examination during the current reporting period (2012-2013). Of these, 70 cases related to paternity/maternity, 79 cases related to identification of deceased, 19 cases were pertaining to sexual assault (rape), 13 cases were related to murder and 5 cases were pertaining to biological relationship (kidney transplantation). Seventeen states and Union Territories of India have availed DNA fingerprinting services of CDFD during this period. Andhra Pradesh forwarded the highest number of cases (104) followed by Bihar (2), Chandigarh (1), Chhattisgarh (13), Delhi (1), Goa (2), Jammu \& Kashmir (2), Karnataka (6), Kerala (4), Madhya Pradesh (1), Odisha (33), Puducherry (1), Punjab (10), Sikkim (1), Tamil Nadu (2), Uttar Pradesh (2) and West Bengal (1) (Figure 1).

During this reporting period, an amount of Rs. 12,39,594/- (Rupees twelve lakhs thirty nine thousand five hundred and ninety four only) has been received towards DNA fingerprinting analysis charges, which is inclusive of service charge as levied by Govt. of India.
The cases involving identification of the deceased ( $42 \%$ ) and paternity ( $38 \%$ ), constituted the bulk of the cases received during this reporting year (Figure 2).

Some prominent cases reported by CDFD during April 1, 2012 to March 31, 2013

1. Paternity dispute case involving a prominent politician - forwarded by Delhi High Court.
2. Identification of victims of the train accident case from Nellore district of Andhra Pradesh.
3. Identification of victims in a triple murder case by a suspected Army deserter: forwarded by CBI, Chennai branch.

Summary of the state-wise break-up of DNA fingerprinting cases:

| State/Union <br> Territory | Biological <br> Relationship | Identity of <br> Deceased | Maternity/ <br> Paternity | Murder | Sexual <br> Assault (Rape) | No. of <br> Cases |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| Andhra Pradesh | 04 | 58 | 34 | 01 | 07 | 104 |
| Bihar | - | - | 02 | - | - | 02 |
| Chandigarh | - | - | 01 | - | - | 01 |
| Chhattisgarh | - | 06 | 07 | - | - | 13 |
| Delhi | - | - | 01 | - | - | 01 |
| Goa | - | - | 02 | - | - | 02 |
| Jammu \& Kashmir | - | 01 | 01 | - | - | 02 |
| Karnataka | - | - | 05 | 01 | - | 06 |
| Kerala | - | 01 | 03 | - | - | 04 |
| Madhya Pradesh | - | - | 01 | - | - | 01 |
| **Odisha | - | 03 | 08 | 10 | 12 | 33 |
| Puducherry | - | - | 01 | - | - | 01 |
| Punjab | - | 08 | 01 | 01 | - | 10 |
| Sikkim | - | 01 | 01 | - | - | 01 |
| Tamil Nadu | - | 01 | 01 | - | - | 02 |
| Uttar Pradesh | - | - | - | - | 02 |  |
| West Bengal | 01 | 79 | 70 | 13 | 19 | 186 |
| Total number <br> of cases | 05 | - |  |  | - | - |

* Among the 104 cases received from Andhra Pradesh, 56 cases were received from Andhra Pradesh State Forensic Science Laboratory (APFSL), Hyderabad under a Memorandum of Understanding (MoU) between CDFD, APFSL and the Government of Andhra Pradesh.
** Among the 33 cases received from Odisha, 9 cases were received at CDFD from DPL-CDFD and 24 cases were received at DPL-CDFD, ILS, Bhubaneswar under an MoU between CDFD, ILS and the Government of Odisha.

4. Matching of DNA profiles in a suspected homicide case of a Pastor from Kerala: forwarded by CBI, Ernakulum, Kerala branch.

## Deposition of evidence in Courts of Law

During this reporting year, the DNA experts defended their reports in 10 cases in various Hon'ble Courts throughout the country.
Training / Lectures / Workshops on DNA fingerprinting examination

## Training

1. Training on DNA profiling techniques to

personnel from Forensic Science Laboratory, Govt. of NCT of Delhi from 14.03.2012 to 30.04.2012.
2. Training on DNA profiling techniques to scientific officer from Forensic Science Laboratory, Madhuban, Haryana from 14.07.2012 to 30.07.2012.
3. Training on DNA profiling techniques to the scientists from the State Forensic Science Laboratory, Lucknow, Uttar Pradesh during 07.05.2012 to 16.05.2012 and 26.11.2012 to 01.12.2012.
4. Training in collection, storage and transportation of biological samples for DNA profiling to Odisha Police Investigation Officers on 11.03.2013 and 25.03.2013 at DPL-CDFD, ILS Campus, Bhubaneswar.

## Lectures/ Workshops

1. Delivered lecture for senior Police Officers at the Sardar Vallabhbhai Patel National Police Academy, Hyderabad on 11.05.2012.
2. Lecture at CDFD for the benefit of post-graduate biology teachers from Kendriya Vidyalaya, Hyderabad on 23.05.2012.


Figure 2. Types of cases received (\%)
3. National Disaster Management Authority (NDMA)-CDFD Workshop on "Identification of victims of mass disasters" held on 08.06.2012 at Hyderabad.
4. Lecture for senior IPS Officers of different states of India from Administrative Staff College of India, Hyderabad on 01.08.2012.
5. Delivered lecture for students at BITS Pilani, Hyderabad Campus on 12.09.2012.
6. Delivered lecture for senior Police Officers at the Sardar Vallabhbhai Patel National Police Academy, Hyderabad on 30.01.2013.
7. Delivered lecture for Probationary IPS Officers at the Sardar Vallabhbhai Patel National Police Academy, Hyderabad on 08.02.2013.
8. Lecture for Air Force Officers from Air Force Intelligence School, Lohegaon, Pune on 12.02.2013.
9. Lecture for post-graduate students and faculty from Aurora's Degree and PG College, Hyderabad on 25.03.2013.
10. Awareness programmes on best practices for collection, storage and transportation of biological samples for DNA profiling at different District Police Head Quarters of Odisha.

Publications

1. Dalal A, Bhavani GSL, Togarrati PP, Bierhals T, Nandineni MR, Danda S, Danda D, Shah H, Vijayan S, Gowrishankar K, Phadke SR, Bidchol AM, Rao AP, Nampoothiri S, Kutsche K and Girisha KM (2012). Analysis of the WISP3 gene in Indian families with progressive pseudorheumatoid dysplasia. American Journal of Medical Genetics A 158A: 28202828.
2. Ranganath P, Sharma V, Danda S, Nandineni MR and Dalal A (2012). Novel mutations in the neuraminidase-1 (NEU1) gene in two patients of sialidosis in India. Indian Journal of Medical Research 136: 1048-1050.

## DIAGNOSTICS DIVISION

| Faculty | Ashwin B Dalal | Staff Scientist |
| :---: | :---: | :---: |
| Adjunct Faculty | Prajnya Ranganath Shagun Agarwal | Assistant Professor, NIMS Assistant Professor, NIMS |
| PhD Students | Anusha Uttarilli <br> Anjana Kar <br> Ashish Bahal | Senior Research Fellow Junior Research Fellow Junior Research Fellow (Since May 2012) |
| Other Members | Aneek Das Bhowmik <br> TNageswara Rao <br> GR Savithri <br> Angalena R <br> PRajitha <br> Usha Rani Dutta <br> Jamal Md Nurul Jain <br> Bhagwati Sharan Sharma <br> C Krishna Prasad <br> R Sudheer Kumar <br> Savita Wangnekar <br> Matta Divya <br> V Subhash <br> Vijay Kumar Pidugu <br> Seetalakshmi S <br> A Sirisha <br> Rajini Jonna <br> Sri Lakshmi BG <br> Sai Shruthi C <br> CH Deepika | Research Associate <br> Research Associate (Since Jan. 2013) <br> Senior Technical Officer <br> Senior Technical Officer <br> Technical Officer <br> Technical Officer <br> Technical Officer <br> Technical Assistant <br> Technician <br> Technician <br> Project Assistant (Since Jul. 2012) <br> Project Assistant (Since Dec. 2012) <br> Project Assistant (Since Aug. 2012) <br> Project Assistant (Till Jul. 2012) <br> Project Assistant (Till Jul. 2012) <br> Project Assistant (Till Jan. 2013) <br> Project Assistant (Till Feb. 2013) <br> Research Assistant (Till Nov. 2012) <br> Research Assistant (Till Jun. 2012) <br> Research Assistant (Till Mar. 2013) |
| Objectives <br> 1. To conduct genetic evaluation for patients/ families with genetic disorders; |  | (I) Details of services provided in the current reporting year (April 1, 2012 - March 31, 2013) Clinical Genetics |
| 2. To develop n genetic analys chromosomal <br> 3. To act as natio and quality co genetic diseas | ods and assays for ngage in research on gene disorders; <br> ral center for analysis genetic tests for few | tal of 3458 patient samples were analysed for etic testing, during the year 2012-13. These sisted of patients with chromosomal disorders, nogenic disorders, mental retardation, genital malformations, inborn errors of tabolism, and other familial disorders. |
| 4. To impart trai patients with $g$ | netic evaluation of ders. | Department of Medical Genetics established zam's Institute of Medical Sciences, rabad is running successfully. A total of 2310 ts were examined and counseled in the unit 2012-13. |

Genetic investigations done during 2012-13

| Investigation | Total cases | Positives |
| :---: | :---: | :---: |
| Cytogenetics | 1213 | $160(13.2 \%)$ |
| Proband | 1096 | $150(13.6 \%)$ |
| Prenatal | 117 | $10(8.5 \%)$ |
| Molecular Genetics | 1233 | $458(37 \%)$ |
| Proband | 1150 | $433(37.6 \%)$ |
| Prenatal | 83 | $25(30 \%)$ |
| Biochemical Genetics | 1012 | $272(27 \%)$ |
| Proband | 991 | $266(26.8 \%)$ |
| Prenatal | 21 | $6(28.6 \%)$ |

Cytogenetics

| Disease | Abnormality | No. of cases |
| :---: | :---: | :---: |
| Down Syndrome | Trisomy 21 | 58 |
|  | 47,SC+21 | 2 |
|  | $46, \mathrm{XY}$,rob(21;21) +21 | 1 |
|  | 46, XX,rob(21;21) +21 | 2 |
|  | 46,XY,t(14;21)+21 | 3 |
|  | 47,XY, +21, inv(9) | 1 |
|  | $47, \mathrm{Xinv}(\mathrm{Y})+21$ | 1 |
|  | 47,XX+21[42]/46, XX[8] | 1 |
| Edward Syndrome | 47,SC,+18 | 2 |
| Patau Syndrome | 47,SC+13,9qh+,15s+ | 1 |
| Turner Syndrome | Monosomy X (45, X ) | 2 |
|  | iso $\mathrm{X},(46, \mathrm{X}, \mathrm{i}(\mathrm{X})$ ) | 3 |
|  | Mosaic 45, $\mathrm{X} / 46, \mathrm{X}, \mathrm{i}(\mathrm{X})$ | 1 |
|  | Mosaic 45, X/ 46, XX | 1 |
|  | 46,X, del(X)(p22.2) | 1 |
|  | 45Y,del(X)(p22.2) | 2 |
|  | 46,XX/45,X,del(X)(q26 ${ }^{\text {ater }}$ ) | 1 |
| Klinefelter Syndrome | 47,XXY | 11 |
| Sex reversal | Phenotypic female with 46, XY | 6 |
|  | Phenotypic male with $46, \mathrm{XX}$ | 2 |
|  | 47,XX+marker | 2 |
|  | 47,XY+marker | 1 |

Fluorescence in situ Hybridization (FISH)

| Disease/translocation | No. of cases | No. of positives |
| :--- | :--- | :---: |
| Prader-Willi Syndrome | SNRPN(15q11)/PML(15q24) | 8 |
| Di-George Syndrome | TUPLE(22q11.2)/ARSA(22q13) | 6 |
| Williams-Beuren | ELN(7q11)/Control(7q22) | 8 |
| Marker chromosome | WCP-15, WCP-22SE(14)/(22), <br> SE(X)/(Y),Acro-p-arm | 23 |
| Spectral karyotyping |  | 6 |

Quantitative Fluorescent PCR (QF-PCR)

| QF-PCR kit | Patients | Positives |
| :---: | :---: | :---: |
| Prenatal QF-PCR | 44 | 1 |
| MLPAkit(P064) MR-1 | 42 | 5 |
| MLPAkit(P064) MR-2 | 4 | 0 |

Structural chromosomal abnormalities

| Inversions |  |
| :---: | :---: |
| 46,XY, inv(9) | 9 |
| 46,XX, inv(9) | 5 |
| 46,X, inv(Y) | 1 |
| Deletions |  |
| 46,XY,del(5)(p)1 |  |
| 46,XX,del(9)(p22) | 1 |
| Translocations |  |
| 45,XY,rob(13;14) | 1 |
| 45,SC,rob(13;14)pat | 1 |
| 46,XX,t(1;10((q43;q24.3) | 1 |
| 46.XY,t(1;5)(p36.3;q31.1) | 1 |
| 46,XY,(4;10)(q13.3;p15) | 1 |
| 46,XX,t(3;15) | 1 |
| 46,XX,t(4;18)(q12;q11.2) | 1 |


| 46,XX,t(7;14) | 1 |
| :---: | :---: |
| 46,XY,t(3;7)(q27;p24) | 1 |
| 46,XX,t(6;13) | 1 |
| 46,XX,t(1;21)(q42.1;q22.3) | 1 |
| 46,XX,t (2;9)(q31;q22) | 1 |
| 46,XY,t(4;5)(q34;q23.1)mat | 1 |
| 46,XX,t(4;5)(q34;q23.1) | 1 |
| Polymorphic variants |  |
| $\begin{aligned} & 46, X X, 9 q h+, 46, X Y, 9 q h+ \\ & 46, X X, 9 q h-, 46, X X, 1 q h+ \\ & 46, X Y, 1 q h+, 46, X Y, 16 q h_{+} \\ & 46, X X, 22 p+, 46, X Y, 22 p+ \\ & 46, X Y, 21 p+, 46, X X, 15 p+ \\ & 46, X Y, 15 p+ \end{aligned}$ | 15 |
| Total | 46 |

Biochemical Genetics

| Disease/Test | Positives |
| :--- | :---: |
| Urine Metabolic Screening <br> tests (340) | 87 |
| Amino acid disorders (N=246) | 59 |
| Maple syrup urine disease | 2 |
| Non Ketotic Hyperglycinemia | 5 |
| Hyperornithinemia | 7 |
| Tyrosinemia | 1 |
| Phenylketonuria | 3 |
| Other amino acid disorders | 41 |
| Lysosomal storage disorders <br> (n=405) | 120 |
| Hurler syndrome (14) | 6 |
| Hunter syndrome (13) | 9 |
| Sanfilippo B (8) | 1 |
| Morquio A disease (30) | 14 |
| Arylsulphatase B (14) | 5 |
| Sly disease (10) | 0 |
| GM1-Gangliosidosis (42) | 10 |
| Fucosidosis (1) | 0 |
| Gaucher disease (37) | 12 |


| Disease/Test | Positives |
| :--- | :---: |
| Krabbe disease (32) | 6 |
| Pompe disease (11) | 4 |
| Nieman Pick disease (34) | 14 |
| Mucolipidosis (12) | 11 |
| Metachromatic Leukodystrophy (82) | 20 |
| Fabry's disease (9) | 4 |
| Mannosidase (2) | 0 |
| Hexosaminidase A/B (54) |  |
| Tay Sachs disease | 1 |
| Sandhoff disease | 3 |
| Prenatal diagnosis (21) | 6 |
| Sandhoff disease (3) | 1 |
| Metachromatic Leukodystrophy (7) | 1 |
| Gaucher's disease (1) | 1 |
| Hunter syndrome (1) | 0 |
| Hurler syndrome (1) | 0 |
| MPS VI (3) | 1 |
| Morquio A disease (3) | 2 |
| GM1- Gangliosidosis (1) | 0 |
| Niemann Pick disease (1) | 0 |

## Molecular Genetics

| Disorders | Cases | Positive | Negative |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: |
| DMD/BMD | 225 | 155 | 70 |  |  |
| DMD Carrier Analysis | 24 | 15 | 09 |  |  |
| Spinal Muscular Atrophy | 101 | 51 | 50 |  |  |
| SMA Carrier Analysis | 43 | 20 | 23 |  |  |
|  |  | Normal | Homozygous | Heterozygous | Compound <br> Heterozygous |
| $\beta$-thalassemia/Sickle cell | 173 | 14 | 55 | 79 | 25 |
| Factor V Leiden | 129 | 126 | - | 03 | - |
| Factor II mutation | 88 | 88 | - | - | - |
| Cystic Fibrosis | 77 | 69 | 08 |  |  |
| Triplet Repeat Disorders |  | Positive | Negative |  |  |
| Friedreichs Ataxia | 66 | 22 | 44 |  |  |
| Myotonic Dystrophy | 38 | 25 | 13 |  |  |
| Huntington Disease | 45 | 23 | 22 |  |  |
| SCA Panel (1,2,3,6 \& 7) | 86 | 29 | 57 |  |  |
| DRPLA | 03 | - | 03 |  |  |
| Fragile X Syndrome | 52 | 05 | 47 |  |  |


| Prenatal Diagnosis |  |  |  |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: |
| DMD | 7 | 1 | 6 |  |  |
| Spinal Muscular Atrophy | 20 | 5 | 15 |  |  |
|  |  | Normal | Homozygous | Heterozygous | Compound <br> Heterozygous |
| $\beta$-thalassemia | 56 | 10 | 13 | 27 | 6 |

Cpd Heterozygous= Compound Heterozygous

## (II) Diagnostics Research

Project 1: Cloning, characterization and analysis of chromosomal rearrangements in human genetic disorders.

Summary of work done until beginning of this reporting year (Upto March 31, 2012)
Structural chromosomal rearrangements that alter the genome architecture can result in human disease phenotypes. Cloning the breakpoint can provide the quickest route to identifying the disease gene in patients with such rearrangements. This project deals with the molecular characterization of chromosomal breakpoint 46,XX,t(X;20)(q13;p13) (Figures $1 \mathrm{~B} \& 1 \mathrm{C}$ ) in a patient with delayed milestones and seizures (Figure 1A).

Lymphoblastoid cell lines of the patient were established and HUMARA assay was performed, which showed a skewed $X$ inactivation of the normal X chromosome. Array CGH studies confirmed that this translocation is not associated with any gains or losses at the breakpoints and elsewhere in the genome. We followed a positional cloning approach for mapping the chromosomal breakpoints X;20 and identified the breakpoint spanning BAC clone RP11-943J20 showing signals on normal X, and split signals on derivative X and derivative 20 (Figure 1D).

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

## Characterization of the ARHGEF9 gene

The breakpoint mapping results of the proband with $t(X ; 20)$ translocation revealed one gene in chromosome X breakpoint (ARHGEF9) and 5 genes in chromosome 20 breakpoint region. We studied ARHGEF9 expression in the patient RNA to study the existence of ARHGEF9 transcripts. Total RNA was isolated and cDNA synthesized. Multiple ARHGEF9 primers were designed for the 2
isoforms of the ARHGEF9 gene. Several combinations of primers were used on control and patient cDNA with exons 1aF-1R, 1-3, 2-4, 5-6, 6-$7,6-8$, and 9-11. The transcripts were present with all the combinations except $1 \mathrm{aF}-1 \mathrm{R}$ and $1-3$ exons. This indicated that the breakpoint lies between the exon 1 and exon 2 or upstream of $A R H$ GEF9 gene (Figure 1E and 1F).The expression of the other exons of ARHGEF9 gene might be due to the effect of a fused gene. The fused gene could be due to the result of fusion with other genes in the vicinity of the breakpoint region on the derivative chromosome 20 region. Hence we also checked for the fusion transcripts with 2 possible genes ( $P R N P$ and $P R N D$ ) on the chromosome 20 region. No products were obtained with this combination of genes using ARHGEF9 primers. We concluded that the ARHGEF9 gene was disrupted in our patient causing the phenotype. There are two more reports in literature of patients with similar phenotype in whom the breakpoints disrupted this gene.

Project II: Clinical, biochemical and molecular analysis of common lysosomal storage disorders.

Summary of work done until beginning of this reporting year (Upto March 31, 2012)

Lysosomal storage disorders are a heterogenous group of disorders associated with specific lysosomal enzyme deficiency. The diagnosis in most of these disorders is based on enzyme assay. There is a large amount of overlap in enzyme levels among carriers and normal people; hence it is very difficult to detect carriers by enzyme assay. Mutation detection is helpful for carrier detection and accurate prenatal diagnosis. Our study aims to characterize the clinical features, biochemical parameters and molecular defects in common lysosomal storage disorders.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

Over last three years we have been able to identify mutations in 159 patients with different lysosomal storage diseases (LSDs) (Table 1). This study has
revealed the mutation spectrum in patients with LSDs in the Indian population. Based on this work, the Indian Council of Medical Research has established a Task Force on LSDs and our centre had been chosen as one of the nodal centres to continue this work for a larger number of LSDs.


Figure 1. Breakpoint mapping in a patient with de novo translocation.(A) Clinical photograph of the patient.(B) Ideograms depicting the translocated chromosomes and their normal homologs. (C) Partial karyogram showing the translocation X and 20. (D) FISH with BAC clone RP11-943J20 showing signals on normal chromosome X and split signals on both derivative X and derivative 20. (E) Schematic diagram depicting ARHGEF9 with the location of the chromosome breakpoint (arrow) as determined by FISH, breakpoint cloning, and RT-PCR experiments. The breakpoint spanning BAC RP11-943J20 is shown below the gene. (F) ARHGEF9 transcript analysis in patient and control cell line RNAs.

Project III: Human exome sequencing to identify novel genes for Mendelian disorders
Summary of work done until beginning of this reporting year (Upto March 31, 2012)
Single gene disorders are rare by themselves but collectively they are an important cause of morbidity and mortality. The identification of genes for single gene disorders has value, not only in prenatal diagnosis and genetic counseling of affected families, but also in basic research towards
understanding gene functions and mechanisms of disease. This in turn can help to improve our knowledge regarding the function of the proteins involved and development of new therapeutic options for both single gene and common multifactorial disorders. In spite of the robustness of classical methods of gene identification like chromosomal breakpoint mapping, linkage analysis and homozygosity mapping, these methods are laborious and require multiple families with multiple affected individuals, thus they cannot
be used for single gene disorders occurring sporadically or exhibiting phenotypic heterogeneity. Alternatively, the availability of massively parallel high throughput sequencing technologies have made it
possible to identify gene for a particular disease using just a few affected individuals. This project is designed to identify mutations in such affected families using exome sequencing.

| Lysosomal storage disorder | Total <br> Patients | Number of <br> mutations | Number of novel <br> mutations |
| :---: | :---: | :---: | :---: |
| MPS I / Hurler syndrome | 30 | 10 | 2 |
| MPS II / Hunter syndrome | 38 | 9 | 2 |
| MPS VI / Maroteaux-Lamy syndrome | 30 | 16 | 13 |
| Niemann-Pick disease | 38 | 30 | 22 |
| Metachromatic leucodystrophy | 20 | 20 | 10 |
| Sialidosis | 3 | 3 | 3 |
| Total cases | $\mathbf{1 5 9}$ | $\mathbf{8 8}$ | $\mathbf{5 2}$ |

Table 1. Data sheet showing all the mutations detected in different patients.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
We report on preliminary results of exome sequencing in one patient with presumed single gene disorder. The patient, a 5 year old girl was
born out of consanguineous union and had similarly affected sibling. This patient presented with slowly progressive ataxia, optic atrophy, spasticity and normal cognition (Figure 2A).


Figure 2. (A) Pedigree of family. (B) Results of homozygosity mapping showing regions of overlapping homozygous variants (Coloured region containing the mutation). (C) Result of exome sequencing seen in Integrative Genomics Viewer (IGV) showing the single base deletion in GJC2 gene. (D) Sanger sequencing chromatograms along with the sequences showing no deletion in the control and homozygous deletion in the patient and heterozygous deletion in mother.

1. Array Comparative Genomic Hybridization analysis
Array Comparative Genomic Hybridization (CytoScan750K_SNP Array (Affymetrix, Santa Clara, CA, USA)) in the patient and affected sibling did not reveal any significant genomic deletion/ duplication. However there were many regions of loss of heterozygosity (homozygous alleles) which was helpful to narrow down the region of interest in variant identification (Figure 2B).

## 2. Exome sequencing analysis

Exome capture was performed on genomic DNA samples from patient using TargetSeq Exome Capture kit and sequencing was done on SOLiD $5500 x L$ platform. The reads obtained were analysed using Lifescope ${ }^{\text {TM }}$ software by mapping against Human Genome Build 19, followed by detection of single nucleotide variants and indels. The variants identified were compared with those in NCBI dbSNP database (GRCh37/hg19). Variant annotation was done using SeattleSeq for location and predicted function.
After filtering of variants, none of the SNP variants were found to be present in overlapping homozygous regions detected by homozygosity mapping. At the same time five potential Indel variants were identified to lie in overlapping homozygous region. Of these, only one variant; a single base pair (G) deletion in GJC2 gene, was found to be matching with the clinical phenotype (Figure 2C). This deletion at base number 228345527 on chromosome 1, causes a frameshift at the $23^{\text {rd }}$ amino acid creating a stop codon (TGA) at codon number 38 which is likely to result in a truncated protein. Further analysis for functional significance of this variant is in process.

## 3. Sanger sequencing analysis

The single base deletion identified in the patient by Exome sequencing was also confirmed by Sanger sequencing (Figure 2D) in both the affected siblings using ABI 3130 Genetic analyzer (Life Technologies, CA, USA). Both parents were found to be heterozygous for the variation.

## Publications

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3. Bashyam MD, Chaudhary AK, Reddy EC, Reddy V, Acharya V, Nagarajaram HA, Devi ARR, Bashyam L, Dalal AB, Gupta N, Kabra M, Agarwal M, Phadke SR, Tainwala R, Kumar R and Hariharan SV (2012). A founder ectodysplasin A receptor (EDAR) mutation results in a high frequency of the autosomal recessive form of hypohidrotic ectodermal dysplasia in India. British Journal of Dermatology 166: 819-829.
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9. Kumar R, Panigrahi I, Dalal A and Agarwal S (2012). Sickle cell anemia-Molecular diagnosis and prenatal counseling: SGPGI experience. Indian Journal of Pediatrics 79: 68-74.
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11. Patil SJ, Bhat V, Dalal A and Santosh JS (2012). Confirmation of the Zechi-Ceide syndrome. American Journal of Medical Genetics A 158A: 1467-1471.
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## शोध <br> Research

# LABORATORY OF MOLECULAR GENETICS Centre of Excellence (CoE) for Genetics and Genomics of Silkmoths 

| Faculty | *J Nagaraju <br> KP Arun Kumar |
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|  | M Muthulakshmi |
|  | S Annapurna Bhavani |
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|  | MJ Reddy |
|  | R Lakshmi Vaishna |
|  | Shantanu Shukla |
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|  | Saikat Chakraborty |
|  | S Srividya |
|  | Adarsh K Gupta |
|  | K Shree Rohit Raj |
|  | Shweta Anjan |
|  | S Babu |

Khorana Chair<br>Scientist<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Junior Research Fellow<br>Staff Scientist<br>Technical Officer<br>Technical Officer<br>Technical Officer<br>Technical Officer<br>Bioinformatician<br>Technical Assistant<br>Technical Assistant<br>Research Associate (Since Jul. 2012)<br>Project Associate(Since Sep. 2012)<br>Project-Junior Research Fellow<br>Project-Junior Research Fellow (Since Oct. 2012)<br>Project Assistant<br>Project Assistant<br>Project Assistant (Since May 2012)<br>CDFD-IKP Fellow

8. Understanding the evolutionary dynamics of $B$. mori Z chromosome in relation to autosomes and sex chromosomes of other species.

The progress made in the projects related to transgenic silkworms, miRNAs and genetic diversity of wild silkmoths is reported here.
Summary of work done until the beginning of this reporting year (Upto March 31, 2012)

* We have successfully developed transgenic silkworms resistant to baculovirus and also transferred the resistance property to a commercial high yielding baculovirus susceptible, diapausing silkworm strain through recurrent backcross strategy.
We have discovered several BmNPV-encoded miRNAs in B. mori and found that BmNPV suppresses the small RNA-mediated host defence to successfully proliferate in the host cells by employing bmnpv-miR- 1 .
* We have characterized partial sequences of silk genes namely Fibroin and Sericins in muga silkworm, Antheraea assama.
* We isolated microsatellite markers with a view to understand the phylogeography of ecoraces of tropical tasar silkworm, A. mylitta and muga silkworm, A. assama.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

Objective 1: Introduction of RNAi-based transgene to high yielding silkworm strains to construct baculovirus resistant strains followed by field trials and maintenance of transgenic silkworm strains.
The anti-baculoviral property in the Nistari genetic background was transferred to a high yielding, baculovirus susceptible bivoltine commercial silkworm strain, CSR2 through transgene (dsRed marker phenotype) selection coupled with microsatellite marker-assisted screening and repeated backcrossing. The various steps involved in the transfer of transgene from Nistari to CSR2 lines are pictorially represented in Figure 1. The recurrent backcrossed lines, which were at $\mathrm{BC}_{4} \mathrm{~F}_{13}$
were further advanced to $\mathrm{BC}_{4} \mathrm{~F}_{22}$ generation by rigorous selection for various traits such as fecundity, cocoon shape, cocoon weight, cocoon shell weight, cocoon shell ratio and silk filament length. Also, the introgressed lines were subjected to baculoviral infection to ascertain their resistance level vis-a-vis control lines. The CSR2 line incorporated with the transgene has commercial silk and cocoon traits ( $\sim 1.4 \mathrm{~g}$ cocoon weight; $>800$ m filament length) similar to the nontransgenic CSR2 line. The transgenic silkworm lines, the introgressed transgenic lines, other productive recipient silkworm lines and various genetic stocks are being monitored for the transgene stability, viral resistance and unique traits of the strains under laboratory conditions at the collaborating institutes namely Central Silk Board (CSB), Bangalore and Andhra Pradesh State Sericulture Research and Development Institute (APSSRDI), Hindupur. CDFD in coordination with Biotech Consortium India Limited (BCIL) approached the Review Committee on Genetic Manipulation (RCGM) by providing a road map for field trials of silkworms. As per the recommendations of RCGM, three transgenic lines will be tested in selected R\&D institutes of CSB and APSSRDI.


Figure 1. Schematic diagram showing introduction of transgene (dsRNA for multiple essential baculoviral genes) from the transgenic Nistari lines to high yielding CSR2 strain through recurrent backcross strategy. The recurrent backcrossed lines were further advanced to BC4F22 generation.

Objective 2: Studies on host-pathogen interaction as mediated by miRNAs

In recent years, the role of miRNAs in different biological processes including host-pathogen interaction is a topic of intensive research. Many of the recent studies have indicated the key role of virus-encoded miRNAs in regulating various host defense responses. In the present study, we have reported a BmNPV encoded miRNA, bmnpv-miR3 , that regulates the expression of its own late genes (cis targets) including the basic DNA binding protein (P6.9), which is important for the late replication of virus in the host, B. mori (Figure 2AC). We have performed both cell culture and in vivo experiments to demonstrate the role of bmnpv-miR3 in the infection cycle of BmNPV in the host. Our results showed that bmnpv-miR-3 expresses during early stage of infection and negatively regulates
the expression of P6.9 and other late genes, which are crucial for the virus in the later stage of infection. We noticed a remarkable increase in BmNPV load, when bmnpv-miR-3 was blocked by Locked Nucleic Acid (LNA) - modified oligonucleotides, implying the involvement of its target genes in BmNPV proliferation. Besides, knockdown of the viral RNA polymerase subunit resulted in a decrease in the expression of cis targets, but an increase in the expression of bmnpv-miR-3, suggesting that bmnpv-miR-3 is likely to be transcribed by host RNA polymerase (Figure 3A-C). Our results suggest that bmnpv-miR-3 mediated controlled regulation of BmNPV P6.9 and other late genes in the early stage of infection provides suitable environment for BmNPV to escape the early immune responses of the host.


Figure 2. Blocking of bmnpv-miR-3 using specific antagomir, LNA-3, resulted in the upregulation of bmnpv-miR-3 cis targets in B. mori larvae. (A) RT-PCR based expression analysis of bmnpv-miR-3 cis targets upon blocking of bmnpv-miR-3 by LNA-3, at 72 hours of post BmNPV infection, (B) RT-qPCR analysis showed more than 25 fold induction in the p6.9 expression upon blocking of bmnpv-miR-3, and (C) RT-PCR based expression analysis of bmnpv-miR-3 cis targets in LNA-3 administered larvae at different time points (24 to 96 hpi) post BmNPV infection. (hpi: hours post infection).


Figure 3. Knockdown of the RNA polymerase subunit gene, lef-9 of BmNPV, negatively affects expression of bmnpv-miR-3 cis-targets. (A) RT-PCR analysis showing knockdown of lef-9 in B. mori larvae, (B) Expression analysis of bmnpv-miR-3 cis-targets upon lef-9 knockdown, determined by RT-PCR, and (C) Enhanced bmnpv-miR-3 expression observed upon knockdown of lef-9, examined by Northern blot.

Objective 3: Characterization of silk genes in Indian golden silkmoth (A. assama)

The Indian golden silkmoth, popularly known as muga silkmoth, is a semi-domesticated silk producing insect confined to a narrow habitat range of the Northeastern region of India. The beautiful muga silk with golden luster is one of the most sought-after biomaterials for its indispensible properties like tensile strength, elasticity and luster. The characterization of the silk genes of $A$. assama is hampered by the unavailability of its genomic data. So we carried out transcriptome sequencing of posterior and middle silkgland tissues and filtered for Fibroin specific sequences based on the conserved 5' and 3' termini of other reported Fibroin sequences. The resulting Fibroin specific contigs were used to design primers specific to the 5' and 3' termini. Long PCR on genomic DNA resulted in the full length ( 9 kb ) amplification of Fibroin gene, which was cloned in to pWKS30, a low-copy plasmid. The full length Fibroin gene is inclusive of a short and highly conserved 5 ' non-repetitious
terminus of 407 bp and a shorter 3' non-repetitious terminus of 252 bp . The gene has one small intron of 131 bp in its 5 ' non-repetitious region and the rest $90 \%$ of the gene codes for the Alanine-Glycine rich highly repetitious crystalline portion of Fibroin protein. This repetitious nature deprecates the possibility of conventional primer-walking for progressive sequencing. Therefore, the full length sequencing was accomplished through a series of restriction digestions followed by sub-cloning and re-sequencing. As part of this full-length sequencing, the Fibroin insert was released and digested separately with specific restriction enzymes. The subclones, which were less than 1 kb , were sequenced directly; while the longer ones were digested again for further sequencing. The restriction map based on the overlapping sequences of the sub-clones was confirmed by the poly-Alanine and non poly-Alanine repeat motifs that resulted in the complete sequence of Fibroin transcript. Northern hybridization revealed the expression of Fibroin exclusively in the posterior silkgland (PSG).

Objective 4: Genetic diversity and population structure of $A$. assama

Owing to the prevailing socio-political problems, the muga silkworm habitats in the Northeastern region have not been accessible, hampering the phylogeography studies of this rare silkmoth. Recently, we have been successful in our attempt to collect muga cocoon samples, although to a limited extent, from their natural habitats (Figure 4). Out of 87 microsatellite markers developed previously for $A$. assama, 13 informative markers were employed to genotype 97 individuals from six populations to analyze their population structure and genetic variation. We observed highly significant genetic diversity in one of the populations (WWS-1, a population derived from West Garo Hills region of Meghalaya state). Further analysis with and without WWS-1 population revealed that dramatic genetic differentiation (global FST $=0.301$ ) was due to high genetic diversity
contributed by WWS-1 population. Analysis of the remaining five populations (excluding WWS-1) showed a marked reduction in the number of alleles at all the employed loci. Structure analysis showed the presence of only two clusters: one formed by WWS-1 population and the other included the remaining five populations, inferring that there is no significant genetic diversity within and between these five populations, and suggesting that these five populations are probably derived from a single population (Figure 4). Patterns of recent population bottlenecks were not evident in any of the six populations studied. A. assama inhabiting the WWS-1 region revealed very high genetic diversity, and was genetically divergent from the other five populations studied. These efforts should be continued to identify such populations from this region as well as other muga silkworm habitats. The information generated will be very useful in conservation of the dwindling muga culture in Northeast India.


Figure 4. Population structure of six $A$. assama populations.

# APEDA-CDFD CENTRE FOR BASMATI DNA ANALYSIS 

Faculty
Other Members

*J Nagaraju

Sabahat Noor
Manju Shukla

* deceased 31 Dec. 2012


## Objectives

1. Testing the purity of Basmati samples received from Export Inspection Council (EIC), Ministry of Commerce, Govt. of India, Basmati rice exporters from India and other countries; and
2. Fine mapping and characterization of the candidate genes governing grain appearance traits of Basmati rice.

Khorana Chair<br>Technical Officer<br>Project Assistant

Basmati samples analyzed at APEDA-CDFD Centre in the current reporting year (April 1, 2012 - March 31, 2013)
During the period under report, a total of 200 (EIC samples 194, Private samples 6) Basmati samples were analyzed and the number of samples indicating the percentage of adulteration with nonbasmati rice is shown in figure below.


Figure 5. Basmati samples analyzed at APEDA-CDFD centre in the current reporting year (2012-2013).

Project 1: Fine mapping and association study of candidate genes in a region on chromosome 5 possibly controlling grain appearance traits of Basmati rice.
Summary of work done until the beginning of this reporting year (Upto March 31, 2012)
Previously, 47 Quantitative Trait Loci (QTLs) governing 18 economically important traits of Basmati rice have been identified in a mapping
population of $189 \mathrm{~F}_{2}$ individuals of a cross between Basmati370 and Jaya by screening 134 polymorphic microsatellite markers. As $F_{2}$ is a primary mapping population wherein phenotypic data was recorded on a single plant without any replications, it was decided to confirm the same QTLs in a permanent population like Recombinant Inbred Lines (RILs). Accordingly, the $F_{2}$ material has been advanced to $\mathrm{F}_{7}$ generation comprising 155 RILs where phenotyping of 18 traits was carried out.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

During the period under report, the DNA was isolated from 155 RILs and is being screened with the same polymorphic SSR markers which have been used in our previous $F_{2}$ study. Out of 74 SSR markers screened so far, 52 microsatellites and 1 indel marker present in the region harbouring grain size QTLs, already identified in $F_{2}$ population between the markers RM289 and RM18600 on chromosome 5, were selected for marker analysis in the RILs. One microsatellite marker i.e., RM18582 was found to show close association with the grain size QTLs. This marker has the potential to be used in marker-assisted improvement of the grain size in Basmati rice.
The QTL qGL5.1 was identified for grain length by interval mapping in the marker interval of RM6024 and RM1237 with phenotypic variation explained (PVE) of $3.7 \%$, which may be a minor QTL or harboring a modifier gene. A major effect QTL, qGB5.1 was identified on chromosome 5 in the marker interval of RM1237-RM18582 with PVE of $3.58 \%$ by composite interval mapping (CIM) and $4.51 \%$ by interval mapping (IM) but located far away from the consistent QTL qGW offering room for further dissecting into a gene. A single QTL qLB5. 1 for length and breadth (LB) ratio in the marker interval of RM1237 and RM18582 was identified with PVE of $10.7 \%$ in the vicinity of the QTLs controlling grain length and grain breadth as LB ratio is a derived trait from the grain length and grain breadth. The genetic distance of the flanking markers harbouring QTL cluster in the previous study was 26.5 cM whereas in the present study it was possible to narrow it down to 15.7 cM . The physical distance also has come down from 11,128 kb to 685 kb .

Screening the mapping population of 155 RILs with the remaining 60 microsatellites is in progress for confirmation and fine mapping of the grain size QTL. Future work plan includes:

1. Further narrowing down of targeted QTL region through:
a. Association mapping;
b. Development of advanced backcross population and near isogenic lines (NILs);
c. Prediction of candidate genes and their structural and functional analysis.
2. Use of SoLiD data to align genomic sequence of the identified QTL region with rice reference genome sequences to check for the variations at genomic level.

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11. Surapaneni M, Vemireddy L, Begum H, Reddy P, Neetasri C, Nagaraju J, Anwar SY and Siddiq EA. Population structure and genetic analysis of different utility types of mango (Mangifera indica L.) germplasm of Andhra Pradesh state of India using microsatellite markers. Plant Systematics and Evolution (In press).

## Other publications

1. Arunkumar KP (2012). Review of: Annual Review of Genetics, 2011. Bonnie L. Bassler et al. (eds). Current Science 103: 947-949.

Patents

1. J Nagaraju et al. Single tube multiplex assay for detection of adulterants in Basmati rice samples.
Indian Patent Application No.: 662/CHE/2006
Indian Patent No.: 251825
Date of grant: April 10, 2012

# LABORATORY OF GENOMICS AND PROFILING APPLICATIONS 

Faculty<br>PhD Students<br>Other Members<br>Madhusudan Reddy Nandineni<br>Anujit Sarkar<br>Soumya Rao<br>Mugdha Singh<br>G Sreeja Reddy<br>Srujana Nagireddy<br>Yamini Sharma<br>Staff Scientist<br>Senior Research Fellow<br>Junior Research Fellow<br>Junior Research Fellow (Since Aug. 2012)<br>Project Assistant<br>Project Assistant (Till Aug. 2012)<br>Project-Junior Research Fellow

(Since Sep. 2012)

## Objectives

1. Development of novel strategies/methodologies for enrichment of human DNA from mixtures containing human and non-human DNAs for DNA profiling-based human identification; and
2. To study the human genetic diversity among various population groups in India.
Project 1: Development of novel strategies/ methodologies for enrichment of human DNA from mixtures containing human and non human DNAs for DNA profiling based human identification
Summary of work done until the beginning of this reporting year (Upto March 31, 2012)
Devising novel strategies for forensic human DNA identification is critical to circumvent the problems associated with the recalcitrant and challenging forensic samples such as those contaminated with non-human DNAs and PCR inhibitors. Enrichment of forensically relevant short tandem repeats (STRs), which showed promising results in our preliminary studies had been adapted and standardized for this purpose. In an effort to develop an STR-enrichment protocol for human identification (HID) purposes, three different methods of STR enrichment namely, primer extension capture (PEC), short hybridization and long hybridization were evaluated to test their efficiency to enrich STR regions from simulated forensic samples. Initial experiments with these simulated samples demonstrated that all the three methods were proficient in enriching the target loci in the presence of high amounts of non-human DNA contamination, whereas further experiments with PCR inhibitors revealed that PEC and short hybridization techniques were efficient (as compared to long hybridization protocol) in overcoming the effects of PCR inhibitors in DNA
profiling. As described in the previous report, the enrichment strategy demonstrated an increased tolerance to various potent PCR inhibitors such as hematin, humic acid and tannic acid up to 5 -fold when as little as 5 ng of initial template DNA was used to generate the STR profiles by employing the commercial multiplex STR kit (AmpFI STR ${ }^{\circledR}$ Identifiler ${ }^{\oplus}$ Plus, Life Technologies, Inc.).
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
Since the challenging forensic samples usually contain limiting amounts of human DNA admixed with various PCR contaminants, sensitivity is an important factor for any enrichment technique. Hence, the experiments with simulated samples contaminated with PCR inhibitors were also carried out with as scant as 2 ng of input DNA. Calcium, which is a component found abundantly in bone and teeth samples and considered as a potent PCR inhibitor was also included for the evaluation of enrichment method. It was observed that both PEC and short hybridization were able to generate the STR profiles from 2 ng of initial template DNA when mixed with up to 1 mM of hematin, $0.4 \mu \mathrm{~g} / \mu$ l of humic acid, $1 \mu \mathrm{~g} / \mu \mathrm{l}$ of tannic acid and 4 mM of calcium, which was $\sim 3-4$ fold increase in tolerance as compared to the STR profiles generated with the same commercial multiplex STR kit without enrichment (controls). Though, these two methods (PEC and short hybridization) have proved to be successful with comparable enrichment efficiencies, PEC method, where the primers got extended after hybridization with target templates, was assumed to be more stable in withstanding stringent washing conditions than partial double stranded molecules obtained after hybridization. Hence, further STR enrichment experiments with simulated samples were carried out employing PEC method.

To simulate the conditions of forensic samples as close as to their natural context, various concentrations of humic acid, tannic acid and calcium were mixed simultaneously with the human DNA along with bacterial DNA since the possibility of occurrence of all these inhibitors together with non-human DNAs in buried human remains such as skeletal bones is very high under normal circumstances. These simulated DNA samples, heavily infested with all possible PCR inhibitors were subjected to enrichment by PEC method for the recovery of respective STR loci, followed by multiplex PCR amplification of these loci employing AmpFISTR ${ }^{\circledR}$ Identifiler ${ }^{\circledR}$ plus PCR amplification kit. Analysis of the results revealed that DNA profiles could be successfully obtained from as little as 2 ng of template human DNA contaminated with 15\% of the maximum independent tolerance limits of each of the inhibitors (mentioned above) coupled with contamination of non-human DNA at 1:10000 ratio (human to bacterial DNA by weight).

Validation of the enrichment method on forensic samples

As the above-mentioned PEC enrichment technique proved to be successful in sequencespecific capture of STR regions in simulated samples, the strategy was tested with real life forensic samples for validation studies. DNA samples obtained from skeletal bones, teeth, fabric, etc. which generated either partial STR profile or no profile were subjected to enrichment by PEC method.

As observed from the STR profiles obtained from few of the highly contaminated forensic samples, multiplex STR PCR kits were unable to generate larger amplicons (as they are more vulnerable to inhibition), even though sufficient quantities of intact human DNA was detected. However, enrichment of STR loci by PEC method prior to STR amplification facilitated in minimizing the effect of the inhibitors and generating the complete STR profile (as represented in Figure 1).
In sample 2, qPCR assay was unable to detect any human DNA and even the commercial STR kit failed to generate any STR profile, but the same sample upon enrichment with PEC method prior to multiplex PCR showed the amplification of maximum number of STR loci, demonstrating the robustness and reliability of hybrid-capture method in enhancing the quality of template DNA during HID testing. This strategy was further used to
successfully generate STR profiles from various challenging forensic exhibits. Thus the application of sequence-specific STR capture and enrichment strategy for challenging and recalcitrant forensic samples would be of tremendous help to the forensic DNA profiling community in increasing the success rate of DNA profiling of these exhibits.
Project 2: To study human genetic diversity in various population groups in India.
Summary of work done until beginning of this reporting year (Upto March 31, 2012)

The second area of interest of the laboratory is to assess the genetic diversity among different population groups in India and to examine the phenotypic effects of genetic variation(s) within and between population groups. As part of the genotype-phenotype correlation studies, SNPs implicated in skin pigmentation were tested for association in different Indian populations. Further, as a step towards designing a SNP-based forensic panel for HID in Indian populations, Genplex panel (a 48-plex SNP-based genotyping system for HID proposed elsewhere for various global populations) was tested for its applicability in the Indian populations and was found to be less informative in these populations as only a subset of the panel (15-20 SNPs) was found to possess the desired characteristics (high heterozygosity and low $\mathrm{F}_{\mathrm{st}}$ ) for forensic HID purposes. A preliminary approach to shortlist additional SNPs for HID testing in Indian populations was described in the previous report.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

1. Evaluation of Kidd SNP panel in Indian populations.

Dr. Kenneth Kidd proposed a SNP-based panel comprising of 92 SNPs for HID testing. The panel had met the desired criteria for HID testing in various continental populations, but was not tested in Indian populations. Hence, it was decided to test its applicability in Indian population groups. In this study, the Illumina GoldenGate ${ }^{\circledR}$ Genotyping Assay system (96-plex) was employed to genotype more than 150 samples belonging to unrelated individuals residing in different geographical locations of India, like Andhra Pradesh, Assam, Jharkhand, Jammu, Tamil Nadu and West Bengal. The genotyping assay was carried out as per manufacturer's protocol. Statistical analyses revealed that the mean call rate of the assay was $97.65 \%$. Three of the 92

## Sample 1:



## Sample 2:



Figure 1. Samples 1 and 2 represent the electropherograms of forensic samples obtained after amplification using multiplex STR PCR amplification kit. Panels P1 and P2 represent STR profiles, without and with STR enrichment by PEC method, respectively.

SNPs tested, which had less than 90\% call rate, were discarded from further analysis. The genetic variation/ allele distribution studies were carried out
at two levels, viz., within population and between populations.

## (a) Within population studies

The within population study was carried out separately for each of the populations to determine the genetic variation and applicability of the Kidd panel. The summary of the within population study is shown in Table 1. As can be gleaned from the table, the mean minor allele frequency (MAF) of
the whole panel was within the acceptable limits for HID testing in accordance to similar studies reported elsewhere, however, for many of these loci when genotyped in Indian populations, the MAF was found to be low, which tended to decrease the overall discriminating power of the HID SNP panel in these populations and hence were discarded from the proposed SNP panel.

| Parameter | AP | Assam | Jharkhand | Jammu | Tamil <br> Nadu | West <br> Bengal |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Average MAF | 0.3854 | 0.3555 | 0.3846 | 0.3963 | 0.3709 | 0.3437 |
| Average Heterozygosity | 0.4436 | 0.4905 | 0.4962 | 0.4792 | 0.4585 | 0.465 |
| LD pairs | 5 | 2 | 4 | 5 | 4 | 2 |

Table 1. Summary statistics of each population. AP: Andhra Pradesh. MAF: Minor allele frequency. LD pairs: SNP pairs found to be in linkage disequilibrium

Further, many SNP pairs were also found to be in linkage disequilibrium (LD), perhaps owing to their physical linkage or other evolutionary factors, which has further reduced the number of informative SNPs for HID testing (due to absence of independent assortment of these loci).

## (b) Between population studies

The $\mathrm{F}_{\mathrm{st}}$ studies were mainly carried out across the six populations mentioned above. When SNPs satisfying the desired criteria i.e., heterozygosity $>0.4$ and $\mathrm{F}_{\text {st }}<0.02$, were sought to be identified, it was found that only 30 of the 96 SNPs tested satisfied these conditions; a number which is not sufficient to give high power of discrimination desirous for a HID SNP panel. In order to devise a SNP panel for HID testing for Indian populations (consisting of 60-80 SNPs), we wished to screen and incorporate additional SNPs appropriate for these populations. Hence additional SNPs were screened from public databases which could potentially form the identity-testing SNPs in Indian populations.
2. Screening SNPs from databases to obtain desired SNPs to be tested for Indian populations
The screening strategy reported in the previous report was modified slightly for SNP screening, wherein, instead of obtaining the SNPs using filters available through the SPSmart database alone, all the genotypes for the listed SNPs (Illumina_Human660Quad chip containing 592652 SNPs) typed in all the major SNP databases (HapMap data, Release \# 28 dated August 2010, 1000 Genome Project data, Phase I, May 2011,

CEPH U. Stanford HGDP (Human Genome Diversity Project) dataset and CEPH NIH-U, Michigan HGDP dataset) were downloaded and classified according to their chromosomal locations. Only unique unrelated samples ( $\mathrm{N}=2744$ ) were analyzed using Powermarker software. The samples were divided according to major geographical regions as mentioned in SPSmart and each region was considered as a single population for further analysis.

The allele distribution data available from the databases was downloaded and used to calculate the statistical parameters like heterozygosity and $F_{\text {st }}$ and the SNPs which successfully passed the desired filters were shortlisted and various statistical tools were employed to screen the SNPs with desired characteristics. A summary of the various filters utilized for this purpose is shown in Figure 2. At the end of this screening exercise, ~ 270 SNPs have been shortlisted and in future these would be genotyped in Indian populations employing the Illumina GoldenGate ${ }^{\circledR}$ Genotyping Assay system to test for their suitability for HID testing.
3. Studies on human salivary microbiome in Indian populations and its implications on human genetic diversity studies (New project)
Metagenomics aims at studying uncultured organisms to understand the true diversity of microbes, their functions, cooperation and evolution, in environments such as soil, water, ancient remains of animals and in sites on living organisms. In the context of human beings, the commensal microbial symbionts (microbiome) are estimated to be highly diverse and provide traits


Figure 2. Outline of the strategy for short listing SNPs for human identification in Indian populations.
that humans did not need to evolve on their own. Though there are several microbial habitats in the human body, but the oral cavity is unique as it provides immense possibilities for a diverse range of microbiota in the intraoral niches including dental surfaces, cheek, hard palate, tongue and saliva. The primary objective of the project is to study the human salivary microbiome to identify the various bacterial taxa in saliva that may be able to provide insights into human population structure and migrations.

The microbial diversity would be studied on the basis of the occurrence and prevalence of various bacterial genera in the saliva of the samples. It is planned to study the microbiome diversity in saliva samples sourced from various geographical locations (States) in India. The genera will be identified by amplification and sequencing of the 16S rRNA gene sequences and comparing them with the sequence entries in the ribosomal database project (RDP-II). A study of the variation
in the occurrence and prevalence of various bacterial genera would provide the microbial diversity of the salivary microbiome in Indian populations and will provide new insights into the population structure of various Indian populations.

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# LABORATORY OF FUNGAL PATHOGENESIS Understanding the Pathobiology of an Opportunistic Human Fungal Pathogen Candida glabrata 

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Staff Scientist<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Junior Research Fellow<br>Technical Officer<br>Project Assistant<br>Project Assistant (Since Sep. 2012)<br>Project Assistant (Since Nov. 2012)

Candida spp. are the leading cause of disseminated fungal infections and rank fourth among the most common nosocomial pathogens. C. glabrata, a regular commensal of human oral cavity and gastrointestinal tract, accounts for 12-20\% of total Candida blood stream infections. C. glabrata infections, which range from mild mucosal to severe life-threatening systemic infections in immunocompromised individuals, are difficult to treat, in part, owing to intrinsic low susceptibility of C. glabrata towards widely used azole antifungals. C. glabrata is a haploid budding yeast which exists in the blastoconidial form in both commensal and pathogenic states. Research in our laboratory is aimed at a better understanding of the interaction of $C$. glabrata with host immune cells, antifungal drug resistance mechanisms and iron uptake and homeostasis mechanisms operational in C. glabrata.
Project 1: Functional genomic analysis of $C$. glabrata-macrophage interaction.

## Objectives

1. Analysis of intracellular behavior;
2. Screening of a C. glabrata mutant library for altered survival profiles; and
3. Identification and analysis of genes required for survival in vitro.

Summary of work done until the beginning of this reporting year (upto March 31, 2012)
Using an in vitro system comprised of human monocytic cell line THP-1, we demonstrated that wild-type C. glabrata cells were able to inhibit maturation of phagolysosome, counteract/survive
the reactive oxygen species (ROS) generated and replicate in THP-1 macrophages. We further showed that $C$. glabrata cells, upon phagocytosis by THP-1 macrophages, modify their chromatin architecture to a closed, condensed form and mutants defective in chromatin remodeling and/or DNA damage repair are attenuated for virulence in the murine model of disseminated candidiasis. Based on global transcriptional profiling, biochemical and microscopy data, we proposed that C. glabrata response to THP-1 macrophage internal milieu is composed of three distinct phases: an Early-, a Mid- and a Late-phase. While the Early-phase ( $0-2 \mathrm{~h}$ ) is defined by activated DNA damage repair signaling, shut-down of translational machinery and remodeled carbon metabolism, the Mid-phase (3-12 h) represents the adaptive response of C. glabrata to macrophage environment and is characterized by heterochromatinization of the $C$. glabrata genome. Latephase $C$. glabrata cells symbolize proliferating cells with active transcriptional machinery. Additionally, by screening of a library of 18,350 C. glabrata Tn7 insertion mutants for altered survival profiles in THP-1 macrophages, via signaturetagged mutagenesis (STM) approach, we identified a set of 56 genes that are required to survive and/ or replicate in the intracellular milieu of THP-1 cells. Identified genes were implicated in diverse biological processes including chromatin and cell wall organization, signal transduction and golgi vesicle transport.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

Reconfigured carbon metabolism, epitomized by decreased glycolysis and increased
gluconeogenesis, glyoxylate cycle and fatty acid degradation is a characteristic signature of several macrophage-internalized fungal pathogens including Cryptococcus neoformans, C. albicans and $C$. glabrata. Thus, to investigate the possibility whether glucose limitation and presence of alternative carbon sources in the macrophage internal milieu act as cues for remodelling of chromatin, we performed three experiments. First, we checked the ability of mutants defective in chromatin organization to utilize different compounds as sole carbon sources and found Cgrsc3-as, Cgrsc3-ba, Cgchz1 and Cghfi1 mutants to be growth-attenuated in medium containing oleic acid, sodium acetate, citric acid and lactic acid as sole carbon sources (Figure 1A). CgRsc3-A and B are orthologs of S. cerevisiae Rsc3 which is a component of a 17 -subunit RSC (remodel the structure of chromatin) complex. $\mathrm{CgCHZ1}$ and CgHFl 1 code for histone chaperone for $\mathrm{Htz} 1 / \mathrm{H} 2 \mathrm{~A}-$ H2B dimer and an adaptor protein required for structural integrity of the SAGA (Spt-Ada-Gcn5-

Acetyltransferase) complex, respectively.
Second, we examined acetylation of histone H 3 on lysine 56 in C. glabrata cells grown in medium containing sodium acetate as the sole carbon source and found it to be diminished compared to dextrose-grown cells (Figure 1B). This is in accord with the reduced H3K56 acetylation observed in macrophage-internalized C. glabrata cells. Importantly, chromatin extracted from both midphase macrophage-ingested C. glabrata cells and sodium acetate-grown cells was resistant to micrococcal nuclease digestion indicating that cellular response to macrophage internalization and alternative carbon source utilization may involve similar chromatin modifications.
Third, we measured total cellular lysine deacetylase (KDAC) activity in macrophage-internalized and sodium acetate-grown cells as reversible protein lysine acetylation is pivotal to the regulation of cellular metabolism. We observed a 2 - to 3 -fold increase in KDAC activity when $C$. glabrata cells


Figure 1. Macrophage-internalized and sodium acetate-grown C. glabrata cells display elevated lysine deacetylase activity. (A) Serial dilution-spotting assay to assess the growth of C. glabrata strains in indicated medium. (B) Immunoblot analysis on whole-cell extracts of wild-type cells, grown for 6 h in YNB medium containing either 2\% dextrose (YNB-D) or 2\% sodium acetate (YNB-S) as the sole carbon source, with antibodies against indicated proteins/modifications. (C) Measurement of cellular lysine deacetylase activity in RPMI-grown and macrophageinternalized C. glabrata cells using trifluoroacetyl-lysine as a substrate. Trichostatin A (TSA, 10 nM ) treatment was used as the assay specificity control. Data represent the mean of three independent analyses ( $\pm$ SEM). (D) Measurement of cellular lysine deacetylase activity in C. glabrata cells grown either in dextrose- (YNB) or sodium acetate ( NaOAc )-supplemented medium. Data represent the mean of three independent analyses ( $\pm$ SEM).
were either ingested by macrophages or grown in medium containing sodium acetate as the sole carbon source (Figure 1C and D).

Intriguingly, Cgrsc3-as mutant did not show elevated KDAC activity under these conditions (Figure 1C and D) indicating an impaired metabolic regulation. Collectively, these data suggest that response of C. glabrata cells to macrophage environment could be a mimic of the cellular carbon starvation response and chromatin architecture reconfiguration plays an important role in the metabolic adaptation of macrophage-ingested $C$. glabrata cells.
Additionally, we screened C. glabrata mutants, identified through the STM screen for reduced survival in THP-1 macrophages, for their ability to prevent maturation of phagolysosome and identified three mutants which co-localized with acidified phagosomes. Experiments are currently underway to characterize these mutants and better understand the mechanisms that C. glabrata employs to modulate phagolysosomal acidification in macrophages.

Project 2: Innate resistance of C. glabrata to fluconazole

## Objectives

1. Understanding the molecular basis of low inherent susceptibility of $C$. glabrata towards fluconazole; and
2. Identification of targets for combinatorial therapy with azole antifungals.

Summary of work done until the beginning of this reporting year (upto March 31, 2012)

We have previously reported two components of RNA polymerase II mediator complex (CgMed2 and CgPgd1) and three players of Rho GTPasemediated signaling cascade (CgBem2, CgSlt2 and CgBnr1) to be essential for survival of stress imposed by the azole antifungal, fluconazole. Fluconazole targets an essential enzyme of the ergosterol biosynthesis pathway, lanosterol 14 $\alpha$ demethylase (CgErg11). CgBem2, CgSIt2 and CgBnr1 encode a RhoGAP (GTPase activating protein) domain-containing protein, a terminal MAP kinase of Protein kinase C (PKC)-mediated cell wall integrity (CWI) pathway and a formin protein, respectively. PKC signaling is known to be regulated by a small guanosine triphosphatase, Rho1. Further, we demonstrated that disruption of

CgBem2 resulted in bud-emergence defects, azole susceptibility, constitutive activation of PKCmediated CWI signaling and defective regulation of multidrug transporters upon fluconazole exposure. We also showed that genetic abrogation of CgSIt2 rendered C. glabrata cells unable to survive fluconazole stress and defects in the transcriptional up-regulation of multidrug efflux pumps partially accounted for the viability loss of Cgsilt2a mutant.

## Details of progress made in the current

 reporting year (April 1, 2012 - March 31, 2013)In the current study, we have focussed on CgMED2 which codes for a fungal-specific, tail-subunit of the multiprotein mediator complex. Mediator complex is composed of 3 modules, the head, the middle and the tail wherein the tail region interacts with gene-specific activators/repressors and the head and the middle modules interact with core RNA polymerase subunits to form the RNA polymerase II holoenzyme. Mediator complex is essential for activator-dependent transcription in eukaryotic cells. To investigate the molecular basis underlying increased sensitivity of Cgmed2:: Tn7 mutant to fluconazole, we first created a clean knock-out strain by deleting the CgMed2-encoding ORF, CAGLOC04477g, via homologous recombination-based strategy, using nourseothricin acetyltransferease (confers resistance to nourseothricin) as a selection marker. Serial dilution spotting assays verified the enhanced susceptibility of Cgmed24 deletion strain to fluconazole and this elevated sensitivity was complemented by expressing CgMED2 ectopically from a plasmid (Figure 2A).
To examine whether CgMed2 is involved in the transcriptional activation of multidrug transporters upon fluconazole exposure, we checked the expression of the CgCDR1 gene which codes for the major multidrug efflux pump in C. glabrata, via quantitative real-time PCR. Compared to 5 -fold elevated transcription of CgCDR1 in wild-type cells, Cgmed24 could upregulate CgCDR1 only by 1.5 fold in response to fluconazole treatment. Consistent with diminished CgCDR1 transcript levels, Cgmed $2 \Delta$ mutant also exhibited significantly reduced ATP-dependent efflux of rhodamine 6G (R6G) dye, a CgCdr1 substrate. Importantly, both wild-type and Cgmed $2 \Delta$ mutant displayed $\sim 5$-fold increase in CgERG11 transcript levels upon fluconazole exposure. Together, these data
indicate a specific role for CgMed 2 in the transcriptional regulation of multidrug efflux pumps.

Expression of genes encoding multidrug transporters in C . glabrata is regulated by a $\mathrm{Zn}_{2} \mathrm{Cys}_{6}$ zinc cluster-containing transcription factor, $\mathrm{Cg} 2 \mathrm{Pdr1}$, which is known to interact with the Med15 tailsubunit of the mediator complex. To examine if

CgMed2-mediated transcriptional activation of CgCDR1 is via its interaction with CgPdr1, we tagged CgMed 2 with the myc epitope at both N and C -terminal. Functionality of N - and C-terminally-myc-tagged CgMed2 was verified by their ability to complement the fluconazole sensitivity of Cgmed24 mutant (Figure 2A). Western blot and immuno-


Figure 2. Growth profiles of C. glabrata mutants with altered fluconazole susceptibility profiles. (A) Spot assay to assess the growth of indicated C. glabrata strains in YPD and YPD medium containing $16 \mu \mathrm{~g} / \mathrm{ml}$ fluconazole (FLC-16). (B) Representative plate images from the altered fluconazole susceptibility mutant screen. Green and yellow circles denote mutants which displayed sensitivity to $16 \mu \mathrm{~g} / \mathrm{ml}$ (FLC-16) and resistance to $64 \mu \mathrm{~g} / \mathrm{ml}$ (FLC64) fluconazole, respectively. (C) Heat map illustrating the growth of fluconazole-resistant and fluconazolesensitive mutants in the presence of diverse stresses. Growth conditions tested were rich medium (YPD), thermal stress ( $42^{\circ} \mathrm{C}$ ), sorbitol ( 1 M ), glycerol-containing medium (YPG, $3 \%$ ), cell wall stress (caffeine, 10 mM ), membrane stress (SDS, $0.05 \%$ ), antifungal stress (fluconazole (FLC, 16, 32 and $64 \mu \mathrm{~g} / \mathrm{ml}$ ) and clotrimazole (CTZ, $30 \mu \mathrm{~g} / \mathrm{ml}$ )) and cycloheximide (CHX, $1 \mu \mathrm{~g} / \mathrm{ml}$ ). Scaled growth scores are color-coded according to the legend at the bottom.
precipitation experiments are currently ongoing to examine the interaction between CgMed 2 and CgPdr1 upon fluconazole exposure.

Additionally, we screened 9,134 C. glabrata Tn7 insertional mutants for altered fluconazole susceptibility profiles using plate growth assays (Figure 2B) and identified a total of 200 and 231 mutants which displayed sensitivity and resistance to fluconazole, respectively. After three rounds of retesting, we selected a set of 54 mutants, which are likely to carry Tn 7 insertions in unique genes, for further analysis. Of these, 19 mutants were resistant to fluconazole while 35 mutants exhibited sensitivity to fluconazole (Figure 2C). Phenotypic profiling of these mutants revealed varied levels of susceptibility to azole antifungals (fluconazole and clotrimazole), CgCdr1 substrate (cycloheximide) and thermal and cell wall stress (Figure 2C). Mutants also differed in their ability to utilize glycerol as carbon source and sorbitol to rescue
thermal and cell wall stress-related growth defects (Figure 2C). Notably, mutants with dysfunctional mitochondria have been reported to be growthattenuated in glycerol-supplemented medium and highly resistant to azole antifungals. Mapping of the Tn7 insertions in this select set of 54 mutants is currently underway to deduce the identities of the genes disrupted.

## Publications

1. Rai MN, Balusu S, Gorityala N, Dandu L and Kaur R (2012). Functional genomic analysis of Candida glabrata-macrophage interaction: role of chromatin remodeling in virulence. PLoS Pathogens 8: e1002863.
2. Bairwa G, Balusu S and Kaur R. Aspartyl proteases in human pathogenic fungi: roles in physiology and virulence. The Fungal Cell Wall. Editor: Héctor M Mora-Montes. Nova Science Publishers (In press).

## LABORATORY OF IMMUNOLOGY

## Role of advanced glycation end products (AGE) in inducing various cellular activities related to diabetic responses and its regulation

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## Objectives

1. Understanding the biological effects of advanced glycation endproducts (AGE)mediated lipogenesis;
2. Understanding the molecular mechanism of autophagy; and
3. Studies on regulation of cytokine receptors to regulate tumorigenesis and inflammatory responses.

Summary of work done until the beginning of this reporting year (Upto March 31, 2012)

Several protein tyrosine kinase (PTK) inhibitors predominantly isoflavones, such as genistein, erbstatin, quercetin, daidzein, present in red clover, cabbage and alfalfa, show apoptotic effects against cancer cells. In this study, we found that biochanin, a methoxy form of genistein, inhibited IL-8-mediated activation of nuclear transcription factor kappaB (NF-кB), activator protein 1 (AP-1), and its dependent genes more potently than genistein. Both biochanin and genistein potently inhibited activity of Lck and Syk, but biochanin specifically inhibited activity of IKK. Genistein was unable to inhibit IL-8-induced IKK activity, but it blocked PVinduced IKK activity. The data showed that both biochanin and genistein are potent inhibitors of PTK, but biochanin is a potent inhibitor of serine/threonine kinase too. Biochanin inhibited NF-кB activation not only by blocking the upstream IKK, but also

PTK that phosphorylate tyrosine residues of $\mathrm{I}_{\mathrm{\kappa} \mathrm{~B} \alpha}$ Thus, the double-edged sword effect of inhibition of NF-kB via inhibition of both serine/threonine kinase and PTK by biochanin might show useful therapeutic value against activities of cells that lead to tumorigenesis and inflammation (Manna SK, Biochem. Pharmacol., 83: 1383-1392, 2012). Thus, the double-edged sword effect of biochanin to inhibit cellular kinases may be useful to regulate several biological responses that are deleterious to cells, and use this molecule as a therapeutic.
Advanced glycation end products (AGE) accumulate in diabetic patients due to high blood glucose levels and cause multiple deleterious effects. In this report we provided evidence that the AGE increased cell death, one such deleterious effect. Methyl glyoxal coupled human serum albumin (AGE-HSA) induced transcription factors like NF-кB, NF-AT, and AP-1. AGE acts through its cell surface receptor, RAGE and degranulates vesicular contents, including interleukin-8 (IL-8). Degranulated IL-8 acts through its receptors, IL8Rs and induces sequential events in cells increase in intracellular $\mathrm{Ca}^{2+}$, activation of calcineurin, dephosphorylation of cytoplasmic NFAT, nuclear translocation of NF-AT, and expression of FasL. Expressed FasL increases activity of caspases and induces cell death (Mahali et al. J. Biol. Chem., 286: 34903-34913, 2011). Thus, this study may be important in several age-related neuronal diseases where AGE-induced apoptosis is observed because of high amounts of AGE.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

1. Beta-D-glucoside protects against advanced glycation end products (AGEs)mediated diabetic responses by suppressing ERK and inducing PPAR gamma DNA binding
Advanced glycation end products (AGE) that accumulate, due to high amounts of 3 - or 4-carbon sugars derived from glucose, cause multiple consequences in diabetic patients and aged persons. The transcription factor, peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ), is often downregulated in the diabetic condition. Drugs targeting PPAR $\gamma$ for diabetes therapy were developed. We found that AGE inhibited PPAR $\gamma$ activity induced by PPAR $\gamma$ activators, like troglitazone (Figure 1A), rosiglitazone, oleamide, and anandamide. AGE induced translocation of

PPAR $\gamma$ from nucleus to cytoplasm (Figure1B), which was increased on activation of ERK in cells. Antioxidants that inhibit AGE-induced NF-кB activation via ROI generation were unable to protect AGE-mediated decrease in PPAR $\gamma$ activity. Only mangiferin, a $\beta$-D-glucoside, prevented AGEmediated decrease in PPAR $\gamma$ activity, and inhibited phosphorylation of ERK (Figure 1C) and cytoplasmic translocation of PPAR $\gamma$. Mangiferin interacts with PPAR $\gamma$ and enhanced its DNA binding activity as predicted by in silico studies (Figure 1D) and shown by in vitro DNA-binding activity (Figure 1E). Overall, our data suggest that (i) mangiferin inhibited AGE-induced ERK activation thereby inhibited PPAR $\gamma$ phosphorylation and cytoplasmic translocation; (ii) mangiferin interacts with PPAR $\gamma$ and enhances its DNA-binding ability. With these dual effects, mangiferin can be a potential candidate for developing therapeutic drug against diabetes.


Figure 1. Advanced glycation endproducts (AGE)-mediated downregulation of PPAR $\gamma$ activity induced by troglitazone is protected by mangiferin. Jurkat cells, stimulated without or with $10 \mu \mathrm{M}$ troglitazone for 6 h were treated with different concentrations of AGE-HSA for 12 h . Nuclear extracts (NE) were prepared and PPAR $\gamma$ DNA binding was assayed by gel shift assay (A1). Cells, stimulated without or with $10 \mu \mathrm{M}$ troglitazone for 6h were treated with different concentrations of AGE-HSA for 12 h . Whole cell extracts were prepared and amount of TGase was determined by Western blot from 100 ig of extract (A2). Jurkat cells, stimulated with troglitazone for 6 h were treated with $100 \mu \mathrm{~g} / \mathrm{ml}$ AGE-HSA for different times. Amount of PPAR $\gamma$ was determined from CE ( $100 \mu \mathrm{~g}$ ) and NE (50 $\mu \mathrm{g})$ as detected by Western blot (B). Jurkat cells were treated with NAC ( 5 mM ), mangiferin ( $10 \mu \mathrm{M}$ ), vitamin C ( 2 $\mathrm{mM})$, and PDTC $(100 \mu \mathrm{M})$ for 2 h and then stimulated with $100 \mu \mathrm{~g} / \mathrm{ml}$ AGE-HSA for 12 h . Whole cell extracts were used to assay the activity of ERK using MBP as substrate (C). $50 \mu \mathrm{~g}$ of whole cell extracts were used to detect ERK1 by Western blot and the same blot was reprobed for tubulin. Docking interaction of mangiferin with PPAR $\gamma$ (PDB ID: 2PRG) (D1). Mangiferin (PUBCHEM ID: 5281647) and Troglitazone (PUBCHEM ID: 5591) form strong H bond interaction with amino acids of PPAR $\gamma$ (D2). Different concentrations of NE proteins were incubated with mangiferin ( $10 \mu \mathrm{M}$ ), troglitazone $(5 \mu \mathrm{M})$, or both for 2 h and then assayed for PPAR $\gamma$ DNA binding (E).


Figure 2. Pulse treatment of oleandrin increases IL-8-mediated biological responses and suppresses cell death. U-937 cells were treated with different concentrations of oleandrin for 1 h and then cells were washed and cultured for 24 h . Cells were then treated with different concentrations of oleandrin for 24 h . Cell death was determined by PARP cleavage. The PARP (pro- and cleaved fragment) fragmentation was detected from whole cell extract (WCE) of cells with similar treatments and detected by Western blot (A). Total RNA was isolated from cells pulsed with oleandrin followed by cultured for 24 h . The amounts of CXCR4, CXCR2, IL-8, and actin were measured by RT-PCR (B). U-937 cells were pulsed with $100 \mathrm{ng} / \mathrm{ml}$ oleandrin for 1 h , then washed and treated with $1 \mu \mathrm{M}$ cycloheximide or $10 \mu \mathrm{M}$ cystamine for 1 h and then cultured for 24 h . The amount of IL-8Rs were determined from $100 \mu \mathrm{~g}$ of WCE and $50 \mu \mathrm{~g}$ of membrane extract by Western blot (C). The U-937 cells' migration was determined in Boyden chemotactic chamber and chemotactic index (cells number of induced migration/uninduced migration) is indicated in the figure as determined from oleandrin-pulsed and non-pulsed cells against different concentrations of IL-8 (D). Oleandrin-pulsed and non-pulsed cells were suspended in phenol-free DMEM and stimulated with different concentrations of IL-8 for 4 h . Activities of myeloperoxidase, $\beta$-D-glucuronidase and alkaline phosphatase were measured from the culture supernatant and absorbances were indicated in the figure (E). U-937 cells were pulsed with oleandrin and incubated with CsA $(2.5 \mu \mathrm{M})$ for 2 h and then stimulated with IL8. The amount of NF-AT DNA binding was measured by gel shift assay (F).Oleandrin-pulsed cells were stimulated with $\mathrm{IL}-8(100 \mathrm{ng} / \mathrm{ml})$ for 2 h and the calcineurin activity was determined by measuring inorganic phosphate released from phospho-peptide (RII peptide) (G). Oleandrin-pulsed and non-pulsed cells were stimulated with different concentrations of IL-8 for 4h. NE were prepared and NF-кB, AP-1, and Oct1 DNA binding was determined by gel shift assay (H).
2. Cardiac glycoside-pulse enhances IL-8mediated biological responses by increasing cell surface IL-8 receptors

Cardiac glycosides are potent inducers of cell death, but very toxic to cells. Use of these
molecules as therapeutics after reducing toxicity would be viable strategy. In this report we provide evidence that oleandrin alone induced cell death, but pulse treatment of it did not show any induction of cell death (Figure 2A). Pulse exposure of oleandrin, but not by azadirachtin, resveratrol,
thiadiazolidine, or benzofuran enhanced IL-8-, but not TNF-, IL-1-, EGF-, or LPS-mediated induction of NF-кB. This enhancement of NF-кB activation is not restricted in specific cell types. Increase in IL-8-mediated biological responses further proved in the oleandrin-pulsed cells upon overexpression of TRAF6. Oleandrin-pulsed cells did not show increase in NF-kB activation mediated by other ligands for G-protein-coupled receptors, except IL8. Oleandrin-pulse increased expression of IL-8Rs (CXCR1 and CXCR2) (Figure 2B \& 2C) thereby increased IL-8-induced biological responses like chemotaxis (Figure 2D), proteolytic enzymes release (Figure 2E), and activation of NF-кB and AP-1 (Figure 2H). Increase in IL-8Rs further enhanced IL-8-mediated intracellular $\mathrm{Ca}^{2+}$ release, calcineurin activation (Figure 2G), NF-AT activation (Figure 2F), and wound healing. Oleandrin pulse treatment decreased cell surface IL-8Rs by changing the microviscosity and further culturing compensated IL-8Rs by degranulation and expression of NF-AT-dependent transcription. Overall, for the first time we are providing data that the pulse exposure of toxic cardiac glycoside enhances biological activity in a typical manner by activating IL-8-mediated biologic responses. This study might be helpful to design oleandrin for therapy against those diseases where cell migration is required to improve the conditions of patients.

## Publications

1. Fialho AM, Salunkhe P, Manna S, Mahali S and Chakrabarty AM (2012). Glioblastoma multiforme: novel therapeutic approaches. ISRN Neurology 2012: Article ID 642345, doi:10.5402/2012/642345.
2. Mahali SK and Manna SK (2012). Beta-Dglucoside protects against advanced glycation end products (AGEs)-mediated diabetic responses by suppressing ERK and inducing PPAR gamma DNA binding. Biochemical Pharmacology 84: 1681-1690.
3. Manna SK (2012). Double-edged sword effect of biochanin to inhibit nuclear factor kappaB: suppression of serine/threonine and tyrosine kinases. Biochemical Pharmacology 83: 1383-1392.
4. Mulakayala C, Babajan B, Madhusudana P, Anuradha CM, Rao RM, Nune RP, Manna SK, Mulakayala N and Kumar CS. Synthesis and evaluation of resveratrol derivatives as new chemical entities for cancer. Journal of Molecular Graphics and Modelling (In press).

# LABORATORY OF BACTERIAL GENETICS <br> Studies on Gene Regulation, Transcription Termination, and Amino Acid and Ion-Transport in Escherichia coli 

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The work undertaken by the group in this reporting year is described below under the following objectives:

## Objectives

1. To understand the role of Rho-dependent transcription termination in avoidance of R-loops in Escherichia coli;
2. To characterize a novel pathway for potassium translocation in E. coli;
3. To determine mechanisms of export of basic amino acids in E. coli;
4. To examine the interplay between (p)ppGpp metabolism and tmRNA;
5. To test the role of (p)ppGpp in transcription elongation;
6. To delineate the role of transketolases in $E$. coli physiology; and
7. To develop new processes for dehairing of animal skins and hides during leather manufacture.

Summary of work done until the beginning of this reporting year (Upto March 31, 2012)
The work undertaken in earlier years on each of the objectives has been summarized in the first parts of the corresponding descriptions below.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

1. Role of Rho-dependent transcription termination in avoidance of R-loops in $E$. coli
It is well established that transcription and translation are coupled in all bacteria including $E$. coli. When transcription proceeds in the absence of translation, for example beyond the end of an open-reading frame, it is terminated by a process called Rho-dependent transcription termination (RDTT) in which the Rho protein binds the nascent untranslated transcript and mediates the release of RNA polymerase from DNA. Another protein that participates in RDTT is NusG, and both Rho and NusG are individually essential for viability in
E. coli. Compromised RDTT function, arising from missense mutations in rho or nusG, is associated with a phenotype of "polarity relief", that is, absence of premature transcription termination immediately downstream of a nonsense mutation in the promoter-proximal gene of an operon.
In last year's Annual Report, two lines of evidence emanating from the work of this laboratory were described to support the hypothesis that RDTT is essential for prevention of excessive occurrence of RNA-DNA hybrids or R-loops across the genome. One of the assays for detection of R-looped regions exploits the sensitivity to sodium bisulphite of the displaced non-template DNA strand that results in C-to-T changes in nucleotide sequence of the latter. With the aid of this assay, we could show that Rloops occur genome-wide at a basal level of around $5 \%$ in wild-type E. coli, and that the frequency of their occurrence is elevated in a nusG missense mutant deficient for RDTT. The second line of evidence was that the lethality associated with complete absence of Rho or NusG in wild-type $E$. coli could be rescued (on defined medium but not rich medium) by ectopic expression of UvsW, an R-loop helicase from phage T4, thereby establishing that it is indeed excessive R-loops that are probably responsible for inviability of these mutants.
In the current year, additional experiments were undertaken with the UvsW-expressing strains to exclude alternative explanations for the finding that such strains retain viability on defined medium even in absence of Rho and NusG proteins. For example, it has been shown earlier that UvsW is able not only to unwind R-loops but also to catalyze the regression of blocked replication forks. We therefore considered the possibility that suppression by UvsW of lethality associated with compromised RDTT is because of an enhanced ability to restart replication following double-strand breaks at blocked forks in these mutants. We found, however, that UvsW-mediated rescue of $\Delta$ rho lethality occurs even in strains that are deficient for RecA, RecB, or PriA, which are some of the other proteins that are crucially involved in fork restoration and restart. Thus, it appears that UvsW is acting more to prevent fork blockage by R-loop unwinding than to resolve the blocked forks through its fork regression activity.

The additional results that were obtained in the current year in this regard were the following: (i) The rich-medium sensitivity of $\Delta r h o$ mutants expressing UvsW was elicited not only on LB
medium but also on nutrient agar and on defined medium supplemented with $5 \%$ (but not $0.5 \%$ ) (wt/ vol) Casamino acids, indicating that it is the growth rate and not any specific component of the growth medium that modulates UvsW's ability to rescue $\Delta r h o$ lethality. (ii) UvsW expression did not alter the nonsense polarity-relief phenotype conferred by a rho missense mutation at two different loci that were tested, and even the $\Delta$ rho strain expressing UvsW was polarity-relieved; these data indicate that UvsW was not merely substituting for Rho's termination function while mediating the rescue of $\Delta r h o$ lethality. (iii) The E. coli enzymes RNases HI and HII catalyze degradation of RNA in RNA-DNA hybrids, and we found that the Ts growth phenotype of a mutant doubly defective for these enzymes can be partially rescued by UvsW; this observation supports the notion that UvsW acts in vivo to reduce toxicity associated with excessive RNA-DNA hybids. (iv)We also found that the Ts phenotype of a mutant deficient for both RNases HI and HII is manifested only in rich medium, suggesting that R-loops act to impede replication fork movement. (v) We have previously shown that rho or nusG missense mutations can suppress lethality associated with defective RNase E function, and in the present year it was demonstrated that such suppression can also be achieved with $\Delta r h o$ mutation in presence of UvsW. (vi) Finally, UvsW expression could also suppress inviability arising from exposure of a wild-type strain to either of two Rho inhibitors, namely the antibiotic bicyclomycin or the protein Psu from phage P4.
Future work in the laboratory on this project is being directed towards (i) employment of additional assays for R-loop demonstration in RDTT-defective mutants; (ii) deriving the relationships between RDTT and DNA supercoiling in R-loop avoidance and generation; (iii) identifying the determinants of R-loop formation in particular regions of the genome, such as antisense transcription, nucleotide sequence preferences, and so on; and (iv) delineating the mechanisms of R-loop toxicity, for example, whether it is through transcriptionreplication conflicts.
2. Evidence for a novel cryptic pathway for potassium translocation in E. coli

In earlier studies, we have found that the commonly used E.coli strain MC4100, lacking the nucleoid protein H-NS in combination with either a deficiency for thioredoxin 1 or thioredoxin reductase (collectively designated Trx) is rendered sensitive
to an extracellular $\mathrm{K}^{+}$concentration $\left(\left[\mathrm{K}^{+}\right]_{\mathrm{e}}\right) \geq 20 \mathrm{mM}$, a phenotype that has also been reported for a strain deleted for ptsN. PtsN is the terminal phosphoacceptor protein of a paralogous phosphotransferase pathway comprising PtsP-PtsO-PtsN and the phosphorylation substrate of PtsN is unknown. The $\mathrm{K}^{+}$sensitive ( $\mathrm{K}^{\mathrm{S}}$ ) phenotype of the trx hns (TH) and ptsN mutants persisted in the absence of known $\mathrm{K}^{+}$uptake systems, indicating that the $K^{s}$ phenotype may result due to the activity of an as yet uncharacterized $\mathrm{K}^{+}$transport system. To delineate the mechanistic basis of the $\mathrm{K}^{s}$ phenotype we have earlier isolated and characterized genetic suppressors of the $\mathrm{K}^{s}$ phenotype. We have found that the $K^{s}$ phenotype of the trx hns (TH) and the ptsN mutants was dependent upon elevated ppGpp pools contributed by the spoT1 allele of MC4100. Accordingly presence of the spo $T^{+}$allele suppressed the $\mathrm{K}^{\mathrm{s}}$ phenotype of the TH and the ptsN mutants and did so by mediating a reduction in cellular ppGpp level. In addition the $\mathrm{K}^{s}$ phenotype of the TH and pts $N$ mutants was also suppressed by expression of the $\mathrm{K}^{+}$carrier proteins TetA, and a truncated version of the KdpA (KdpA') K+ translocator subunit or by a null mutation in $y c g O$, that encodes an inner membrane protein of unknown function. The $\mathrm{K}^{\mathrm{s}}$ phenotype of the TH but not of the ptsNmutant was suppressed by null mutations in ptsP and ptsO, indicating that the absence of dephospho-PtsN is the common feature of the $\mathrm{K}^{+}$sensitivity displayed by the TH and ptsNmutants. In addition, barring the effect of spo $T^{+}$, all genetic suppressors of $\mathrm{K}^{+}$ sensitivity exerted their effects without causing significant alterations of cellular ppGpp level.
In this year we assessed the effects of heterologous overexpression of $y c g O$ on the growth of the parent in media of varying $\left[\mathrm{K}^{+}\right]_{\mathrm{e}}$ and found that overproduction of YcgO led to a $\mathrm{K}^{+}$sensitive phenotype in the parent and in a strain that lacked known $\mathrm{K}^{+}$uptake systems, that was suppressed by expression of tet $A$ and by the presence of $s p o T^{+}$, but not by other suppressors of the $\mathrm{K}^{\mathrm{S}}$ of the TH and pts $N$ mutants, indicating that the activity of YcgO may be causal to their $\mathrm{K}^{\mathrm{S}}$ phenotypes and that ppGpp may modulate the activity of YcgO. Overall our studies indicate that the perturbations caused by the trx hns and the ptsN mutations may act in distinct ways to limit the amount of dephospho-PtsN leading to unfettering of a novel ppGpp modulated $\mathrm{K}^{+}$translocation pathway in a medium of high $\left[\mathrm{K}^{+}\right]_{e}$ specified by the activity of YcgO. Based on the genetic suppressor studies,
we propose that the trx, hns mutations may exert their effects at a step(s) upstream of PtsN and influence its phosphorylation status, to limit the amount of dephospho-PtsN and that dephosphoPtsN may fetter the activity of YcgO , rendering its activity cryptic in the parent. The genetic relationship between the trx hns and the ptsN mutations can also be explained if one considers that TrxA/B and H-NS stimulate (directly or indirectly) expression of ptsN. We have discounted this possibility because expression of a chromosomal ptsN-lac transcriptional fusion (located in ptsN) was not affected in strains lacking H-NS or TrxA. Our current studies are directed towards examining the effect of overexpression of $y c g O$ on cellular $\mathrm{K}^{+}$levels and testing the notion that dephospho- PtsN may directly interact with YcgO and fetter its activity.

Recently it has been reported by other workers that besides displaying a ${ }^{\text {S}}$ phenotype, growth of a strain lacking PtsN is impaired in a medium of intermediate ( 22 mM ) $\left[\mathrm{K}^{+}\right]_{\mathrm{e}}$ containing $5 \mathrm{mM} \mathrm{L-}$ leucine (Leu), leading to a Leu sensitive (Leus) phenotype that is suppressed by isoleucine and valine (ILV) supplementation. This phenotype is thought to arise due to a combination of altered intracellular $\mathrm{K}^{+}$levels and L-leucine in the ptsN mutant that perturbs cellular AHAS (acetohydroxyacid synthase) activity synergistically in a medium of intermediate $\left[\mathrm{K}^{+}\right]_{\mathrm{e}}$, giving rise to a state of isoleucine pseudoauxotrophy. In E. coli K-12 the activities of the isoenzymes AHASI and AHASIII constitute the first committed step in the biosynthesis of branched chain amino acids leucine, isoleucine and valine. The activity of a third AHAS namely the valine insensitive AHASII encoded by ilvGM is absent in E. coli K-12 owing to the presence of a naturally occurring frameshift mutation in ilvG. In addition based on a recent report it appears that in a certain genetic background the pathology of the $\mathrm{K}^{+}$ sensitivity of the ptsN mutant in E. coli K-12 may be solely related to $\mathrm{K}^{+}$mediated perturbations of cellular AHAS activity based on the observation that both the Leus and the $\mathrm{K}^{s}$ phenotypes associated with a pts $N$ mutant are reversed in a strain that is ilvG ${ }^{+}$, thought to encode with ilvM an AHAS that may be insensitive to $\mathrm{K}^{+}$. Consistent with recent reports we found that a deficiency of PtsN in MC4100 led to a Leus ${ }^{\text {p }}$ phenotype in medium of a $\left[\mathrm{K}^{+}\right]_{\mathrm{e}}$ of 22 mM supplemented with 5 mM Leu, which was suppressed by exogenous ILV at a concentration of 5 mM . However the presence of
ilvG ${ }^{+}$(or exogenous ILV) suppressed the Leus but not the $K^{s}$ phenotype of the pts $N$ mutant. Our studies thus indicate that $\mathrm{K}^{+}$mediated perturbation of ILV biosynthesis constitutes one but not the only casualty of the $\mathrm{K}^{+}$mediated physiological perturbation in the $p t s N$ mutant and that a strain specific variation(s) is perhaps responsible for the ilvG+ mediated suppression of $\mathrm{K}^{\mathrm{s}}$ of the pts $N$ mutant.
3. Studies on basic amino acid export and exploiting ArgP-ArgO regulation to obtain L-arginine overproduction in E. coli
Earlier work from this laboratory had identified an anonymous ORF yggA (subsequently redesignated as $\arg O$ ) to encode a novel arginine (Arg) exporter ArgO in E.coli. These studies had also shown that argO expression is regulated by a transcription factor ArgP, and that certain gain-offunction variants of $\operatorname{ArgP}$ (designated $\operatorname{Arg}{ }^{d}$ ) conferred high and constitutive expression of ArgO leading to increased excretion of Arg into the culture medium. These findings have been circumscribed as an inventive process for microbial production of Arg, and patented by CDFD in several countries.

Towards understanding the mechanism of Arg export mediated by ArgO we have earlier reported the identification of amino acid residues in ArgO, critical for mediating canavanine (Can) export and its topology in the inner membrane using alkaline phosphatase fusions to ArgO. Furthermore we obtained evidence for functional inter/intrahelical interactions in ArgO based on the isolation of second site suppressor mutations of primary mutations that were critical for mediating export of Can. The canavanine sensitive (Cans) phenotype of the ArgO V 118 E substitution variant was suppressed by the V132A and A60P amino acid substitutions individually whereas that of the S156F substitution was suppressed by the I51T substitution. Overall these studies are indicative of existence of putative functional interactions between transmembrane (TM) segments TM2-TM4 and TM2-TM5 of ArgO. The V118E/V132A suppressor pair may represent an intrahelical interaction. We have extended these studies by constructing combinations of the second site suppressor substitutions A60P, I51T and V132A with other primary substitutions in ArgO that impaired ArgO activity and our results suggest that the suppressive effects of the second site amino acid substitutions in ArgO are specific to a given primary ArgO substitution, namely the V118E and
the S156F substitutions. We are currently exploring alternate approaches to validate the indicated TM interactions in ArgO.

So far in E. coli, ArgO represents the only example of a solute exporter involved in the export of a basic amino acid. In order to identify novel genes whose products may promote export of Arg or L-lysine (Lys) we have earlier reported the isolation of a plasmid from a multicopy E. coli genomic library, whose presence in a strain lacking ArgO suppresses its $\mathrm{Can}^{s}$ phenotype. Our studies have shown that canavanine resistance is mediated by the presence on the plasmid of a novel ORF $y b j E$, predicted to encode an inner membrane protein. Elimination of YbjE exacerbated the Cans phenotype of an argO null mutant but its deficiency on its own did not lead to a Cans phenotype in the parent. On the other hand a ybjE but not an argO null mutant was rendered hypersensitive to the toxic analogue of Lys, thialysine. Furthermore, the $y b j E$ mutant was impaired for growth in a medium containing Lys-Ala but not in media containing ArgAla or His-Ala dipeptides indicating that growth inhibition may result to due toxic intracellular build up of Lys. A strain lacking ArgO grew in all the above mentioned media and the argO mutation did not exacerbate the impaired growth of a ybjE null mutant in Lys-Ala medium. Accordingly ybjE has been renamed as lysO, encoding a putative novel Lys exporter. Our results therefore suggest that $E$. coli appears to employ distinct exporters for the export of the two basic amino acids Arg and Lys.

In continued studies on LysO, we have obtained evidence that growth inhibition of a lysO null mutant in a medium containing the Lys-Ala dipeptide correlated with increased cellular Lys content. In studies directed towards understanding the regulation of $l y s O$, we examined the lysO promoter region and located two binding sites for the ArgR repressor, positioned 102 bp and 123 bp upstream of the translational initiation codon of lysO and a 32 bp inverted repeat located 248 bp upstream of the first ARG box. A deletion of the inverted repeat did not alter the expression of a lysO-lac transcriptional fusion whereas lysO-lac expression was impaired in Arg supplemented minimal media in an ArgR dependent manner. By primer extension the transcription start site of lysO was found to be located 36 bases upstream of the translation initiation codon of /ysO. Currently we are examining the binding of purified ArgR to the lysO regulatory region, to understand its repressive effect on lysO expression.

As mentioned above, an argO mutant was not impaired for growth in a medium containing the Arg-Ala dipeptide and to understand the basis of its resistance to Arg-Ala, we have isolated transposon generated mutants, bearing secondary lesions in an argO background that are rendered hypersensitive to Arg-Ala and have found that null mutations in another ORF ydhE encoding an inner membrane protein orthologous to the multidrug efflux protein NorM of $V$. parahaemolyticus render an argO mutant hypersensitive to Arg-Ala. An argO $y d h E$ double mutant was also hypersensitive to other arginine containing dipeptides such as ArgVal, Arg-Leu and Arg-lle. Loss of YdhE in the parent led to a modest impairment of its growth in ArgAla medium but a strain lacking YdhE was not rendered sensitive to the toxic Arg analogue Can. Our studies indicate that YdhE may function as a dedicated Arg exporter whereas ArgO activity serves mainly to export Can and coincidentally exports Arg.

Finally, in work completed this year under a program of co-operation with the IKP Knowledge Park, Hyderabad, improvements were undertaken on the patented process to achieve Arg excretion in shake flask culture supernatants, up to values of 420 mg per litre. The CDFD-IKP Fellow who had performed this work has registered a start-up company to which the technology has been licensed by CDFD for further development and commercialization, and the company has been successful in securing funding from the Small Business Innovation Research Initiative (SBIRI) for this task.
4. Understanding the genetic interaction between (p)ppGpp and tmRNA(ssrA)/smpB system
In work described in earlier reports, the lethal phenotype arising from the combined deficiency of (p)ppGpp and tmRNA (synthetic lethality) was genetically characterized and the following were inferred:
a) Absence of (p)ppGpp - mediated modulation of transcription contributes to synthetic lethality.
b) Genetic suppression studies using biochemically characterized RNA polymerase (RNAP) mutants defective for elongation properties indicated that the synthetic lethal phenotype could be a consequence of
elongation defect and implied a possible role for (p)ppGpp in the modulation of transcription elongation in vivo.
c) Studies done using various mutant alleles of ssrA indicated that its ribosome rescue function but not that of peptide-tag addition is necessary for supporting cell survival in a (p)ppGpp deficient strain.
ssrA codes for small stable RNA that functions like an alanyl-tRNA and a messenger RNA in order to rescue stalled ribosomes. It is generally believed that SsrA-mediated rescue occurs on ribosomes that contain a 3'-mRNA end at or very near a vacant A site (non-stop mRNA). The alanyl-tRNA like activity of SsrA results in the incorporation of an alanine residue to the growing polypeptide chain and this is followed by the addition of a 10aminoacid tag and the termination of translation due its mRNA like activity. Our earlier studies had shown that slow and fast moving RNAP mutants suppressed and accentuated respectively the ppGpp ${ }^{0}$ ssrA synthetic lethality. Based on this finding we reasoned that the faster movement of RNAP in the ppGpp ${ }^{0}$ strain could result in lesions that require the ribosome rescue function of $\operatorname{ssr} A$ for survival. Generation of non-stop mRNAribosome complex in vivo can be mediated by the action of certain mRNA endoribonucleases that make up the toxin component of the toxin-antitoxin systems. We obtained a strain lacking five toxinantitoxin (AT) loci and tested for ppGpp ${ }^{0}$ ssrA synthetic lethality in this genetic background. We expected not to observe synthetic lethality in this background assuming that one or more of the toxins (mRNA endoribonucleases) would be responsible for mRNA cleavage necessitating SsrA-mediated ribosome rescue in the $\mathrm{ppGpp}^{0}$ strain. Contrary to our expectation the strain continued to exhibit synthetic lethality, indicating that atleast the 5-AT loci tested were not important for the generation of mRNA truncations and SsrA-mediated rescue of ribosomes.

The functionality of the ssrA mediated ribosome rescue system can also be tested through an artificial generation of non-stop mRNA (3'- end without a stop codon) and studying the effect on growth. Using an IPTG inducible non-stop mRNA generating plasmid (kind gift from Prof. Sue Lin Chao) it was observed that the $\mathrm{ppGpp}{ }^{0}$ strain was 100 -fold more sensitive to the presence of IPTG than a wild-type strain and this increased sensitivity was not observed when the plasmid also carried
the ssrA gene. We interpret this as an indication of the SsrA-ribosome rescue machinery working close to saturation in strains lacking (p)ppGpp.
Using an E. coli genomic library generated in the medium-copy number plasmid pACYC184 we identified gene(s) capable of conferring multi-copy suppression of the $\operatorname{ssrA}$-ppGpp ${ }^{0}$ synthetic lethality. A clone carrying the complete ydaW and $r z p R$ genes and 5' truncated ydaV and 3 ' truncated trk $G$ genes conferred suppression. By a series of subcloning and recombineering experiments we have identified that the deletion of the $y$ daWopen reading frame is required to observe suppression. The function of this gene and the mechanism of suppression need investigation.
We also made a serendipitous observation that the ssrA ppGpp ${ }^{0}$ synthetic lethality is not observed during growth in minimal A media containing glucose and casaminoacids or only casaminoacids as carbon source. This brings up the question if specific component(s) in LB media or the physiology of the strain during growth in rich media such as LB is responsible for eliciting the synthetic lethal phenotype.

It has been reported that ssrA/smpB genes aid in cell survival when stable DNA-protein complexes are artificially induced and possibly also during the formation of stalled RNAP elongation complexes. The protein factors that help in the clearance of stalled RNAP elongation compelex are Mfd, Rho, GreA and GreB. We find suppression of ssrA $p p G p p^{0}$ synthetic lethality when the wild type allele of rho is replaced with the rho-4 allele which encodes a hypomorphic Rho protein that exhibits termination defect. In E. coli, the Rho protein causes termination of transcription especially under conditions when the nascent RNA is not translated. The Rho protein binds nascent RNA and the RNAP to cause termination of transcription. We are currently trying to develop a model to explain our findings.

## 5. (p)ppGpp - a role in transcription elongation?

E. coli has two well studied elongation factors called GreA and GreB. When RNAP encounters a block during elongation and backtracks, the transcription factors GreA and GreB suppress pausing by stimulating the intrinsic nucleolytic activity of RNAP. These factors exhibit structural homology to DksA that can positively or negatively modulate the effect of (p)ppGpp on transcription initiation at the promoters of amino acid
biosynthetic genes and ribosomal RNA respectively. However studies have not been performed to study the relationship between (p)ppGpp and the Gre factors. Our preliminary genetic data indicate that a phenotype seen in the greA greB double mutant can be altered through the modulation of intracellular (p)ppGpp pool. The greA gre $B$ mutant exhibits temperature sensitive growth phenotype in LB, presumably because prolonged RNAP pausing prevents replication and/ or transcription and this is suppressed in genetic backgrounds carrying mutant alleles of relA and spoT consistent with an idea that the elevation of intracellular (p)ppGpp content can suppress the temperature sensitivity.

## 6. Role of transketolases in E. coli

 physiologyTwo isoforms of transketolase enzyme, namely, TktA and TktB have been identified in E. coli. Expression of $t k t B$ is regulated by (p)ppGpp/Rpos and comes up during stationary phase while TktA is the major transketolase during exponential growth. Transketolases are required for the transfer of two carbon units between sugar molecules, and are part of the non-oxidative pentose phosphate pathway that connects glycolysis and the pentose phosphate pathway.
Genetic studies carried out in transketolase deficient strain i.e., a $t k t A t k t B$ double mutant and previously reported, revealed that:

1. Transketolase activity is essential to sustain the growth of $E$. coli in LB media and that the growth defect can be partially compensated by: a) elimination of the purine/pyrimidine salvage pathways through inactivation of DeoB (phosphopentomutase) resulting in reduced ribose-5-phosphate pool; b) activation of $\mathrm{g} / \mathrm{p} K$, coding for glycerol-3-phospate kinase; c) presence of the $p n t A B$ genes, coding for the subunits of the membrane bound pyridine nucleotide trans-hydrogenase on multi-copy plasmid and presumed to increase its intracellular activity and d) glucose supplementation to LB media.
2. The suppression of growth defect observed in all the suppressor strains excepting that of multi-copy pntAB is contingent on the presence of the wild-type chromosomal copy of $p n t A$ gene. We interpret this result to indicate that restoration of growth in the various suppressor backgrounds is through the
modulation of intracellular pyridine co-factor levels. It needs to be studied if the suppressor mutations modulate directly the expression of the pntAB genes or that the PntAB activity independently contributes to the net cellular pyridine cofactor pool.

We measured the pyridine cofactor levels in various strain backgrounds and the results are presented in Table 1. Since the $t k t A t k t B$ double mutant is inviable we constructed a strain wherein growth is sustained by the expression of TktB from a conditional $\mathrm{Amp}^{R}$ plasmid whose replication is
mutations largely restore the cofactor levels to that observed under permissive growth condition in the $t k t A t k t B$ double mutant.
7. Studies on biological approaches to the dehairing of animal skins and hides during leather manufacture

For leather manufacture, skins and hides from animal carcasses are commonly preserved from putrefaction during storage and transport by the application of salt (salting); and their subsequent dehairing is achieved (after removal of salt by extensive washing) by the application either of

| Relevant genotype (growth condition) | NAD ${ }^{+}$ | NADH | NADP+ | NADPH |
| :---: | :---: | :---: | :---: | :---: |
| Wild type | $189.5 \pm 2.69$ | $37.05 \pm 15.49$ | $15.78 \pm 0.68$ | $22.96 \pm 11.29$ |
| $t k t A t k t B$ (LB) | $7.8 \pm 1.36$ | $0.31 \pm 0.62$ | $10.34 \pm 1.34$ | Not detectable |
| $t k t A$ tktB (LB amp IPTG) | $151.01 \pm 14.14$ | $14.08 \pm 5.58$ | $21.82 \pm 2.37$ | $14.89 \pm 1.13$ |
| $t k t A$ tktB (LB glucose) | $67.3 \pm 13.93$ | $6.71 \pm 2.32$ | $8.59 \pm 1.04$ | $0.27 \pm 0.29$ |
| tktA tktB deoB (LB) | $142.97 \pm 13.56$ | $17.82 \pm 2.76$ | $15.61 \pm 1.19$ | $6.72 \pm 3.79$ |
| tktA tktB deoB (LB amp IPTG) | $148.36 \pm 12.2$ | $14.48 \pm 7.35$ | $12.82 \pm 1.68$ | $7.29 \pm 2.81$ |
| tkt tktB deoB (0.2\% ribose in LB) | 9.03 | 5.22 | 8.03 | 2.27 |
| $t k t A$ tktB deoB sup8 (LB) | $206.56 \pm 6.14$ | $22.54 \pm 12.83$ | $33.01 \pm 2.04$ | $4.45 \pm 0.26$ |
| tktA tktB deoB sup8 (LB amp IPTG) | $201 \pm 40.49$ | $28.83 \pm 10.23$ | $24.43 \pm 7.41$ | $5.99 \pm 2.27$ |
| $t k t A$ tktB/pACYC184 (LB) | $7.26 \pm 2.17$ | $1.88 \pm 0.32$ | $9.33 \pm 0.92$ | $0.22 \pm 0.38$ |
| tktA tktB/pACYC184 (LB amp IPTG) | 93.33 | 30.36 | 17.06 | 3.41 |
| tktA tktB/pACYC184-pntAB (LB) | $37.9 \pm 3.44$ | $8.69 \pm 1.35$ | $7.29 \pm 1.54$ | $1.06 \pm 0.27$ |
| tktA tktB/pACYC184-pntAB (LB amp IPTG) | $158.19 \pm 27.47$ | $24.65 \pm 2.11$ | $7.88 \pm 4.69$ | $13.13 \pm 8.07$ |

Table 1. Pyridine cofactor levels.

IPTG-dependent. The cultures were grown in permissive conditions (LB amp IPTG) overnight and sub-cultured (1:1500) and growth continued under permissive or non-permissive (LB) growth conditions to mid-log phase. Cells were harvested and normalized (using $A_{600}$ ) and the extraction performed for pyridine cofactor analyses. The results clearly indicate that with the exception of NADP ${ }^{+}$the intracellular levels of the other cofactors are dramatically lowered in the transketolase deficient strain and the presence of the suppressor
chemicals such as lime and sulphide (chemical process) or of enzyme preparations from any of a variety of sources (biological or enzymatic process), or by a suitable combination of chemical and enzymatic processes (enzyme-assisted dehairing). Both salting and the chemical dehairing processes contribute substantially to environmental pollution. Accordingly, the efforts to use enzymes to substitute, either partially or completely, for chemicals in the dehairing process has gained considerable importance.

Commercially available enzyme preparations (proteases, lipases, etc., usually from bacterial or fungal sources) of varying degrees of purity are used in dehairing. The two surfaces of a skin or hide are referred to as the grain side and the flesh side, respectively, and the enzyme preparations are commonly applied to the flesh side during the dehairing process. Two important considerations are (i) an absolute need to preserve the integrity of collagen in the skin or hide, and (ii) the desirability to retain also the integrity of the hairs (hair-saving), since the latter represent a commercially valuable by-product of the dehairing process.
In work undertaken a few years ago in collaboration with the Central Leather Research Institute, Chennai, we had shown that live cultures of the yoghurt bacterium Lactobacillus plantarum grown in milk are doubly effective, first as a substitute to salting in the protection of skins and hides of goat, sheep and buffalo from putrefaction (for up to several months), and also to achieve their satisfactory dehairing in a hair-saving process. It appears that the quality of soft leather obtained using this process may be suitable for the manufacture of products such as gloves or chamois leathers.

The time taken to achieve dehairing by the chemical or enzymatic processes is typically of the order of several hours to a day, and is around two to three days with the use of live Lactobacillus culture preparations. Any reduction in the time taken for dehairing will be of value to industry since it would result in, among other features, faster turnaround times, less inventory costs, and less likelihood of undesirable events such as skin putrefaction. Furthermore, in the enzymatic or enzyme-assisted methods for dehairing, it would be advantageous
in terms of input costs if satisfactory dehairing can be achieved with smaller amounts of enzyme preparation to be applied.
In work completed during the present year, we have developed a process by which the dehairing of animal skins and hides is achieved within a matter of two to ten minutes, that is, much faster than that obtained by any of the methods in current practice. In this simple yet novel process, a suitable source of enzyme is placed on the flesh side of a skin or hide and direct electric current of appropriate polarity is then applied across the thickness of the skin for a few minutes; this is achieved by placing the skin between a pair of metal surfaces that are then connected to a source of electric current such as a set of dry cells, a leadacid battery, or an electrophoresis power pack. After this treatment, the skin is gently scraped to result in a hair-saving mode of dehairing (Figure 1).

An important consideration is the polarity of voltage applied and the consequent direction of electric current flow through the skin (that is, whether it is from the flesh side to grain side or the reverse) that is suitable for the dehairing process; our experiments indicate that this is determined by the nature of the enzyme source as well as by the pH , and that for any particular combination the dehairing is achieved only with one polarity and not the other. These observations provide strong support to the notion that it is electrophoretic enzyme delivery to the hair follicles which is responsible for the extreme rapidity of the dehairing process. An additional advantage with this method is that the quantity of enzyme needed for dehairing is substantially less than that in the absence of any applied voltage.


Figure 1. Example of dehairing of goat skin following application of enzyme and direct electric current for 10 minutes. Images shown at left and right are, respectively, of skin before and after treatment; in the latter, one part of the skin has been gently scraped to depict the ease of hair removal.

We have shown that this method is effective for dehairing of skins or hides from a variety of animals, as well as with different commercial enzyme preparations. Dehairing could also be achieved with a sonicated extract of cells of L. plantarum, which is classified as a GRAS (Generally Regarded as Safe) microorganism in industry. The method has also successfully been applied to achieve dehairing of a stack of several skins, in which enzyme has been applied to the flesh side of each skin, the skins are stacked one above the other in the same orientation, and electric current is then passed across the entire stack.

It is expected that the electrophoretic process may also be useful in other steps of leather manufacture that employ enzymes, such as soaking, liming, bating, and degreasing. We are at present trying to engage with suitable industries to undertake further testing and development of applications of the enzyme electrophoretic process in leather processing and manufacture.

## Publications

1. Marbaniang CN and Gowrishankar J (2012). Transcriptional cross-regulation between gramnegative and gram-positive bacteria, demonstrated using ArgP-argO of Escherichia coli and LysG-lysE of Corynebacterium glutamicum. Journal of Bacteriology 194: 5657-5666.
2. Leela JK, Syeda AH, Anupama K and Gowrishankar J (2013). Rho-dependent transcription termination is essential to prevent excessive genome-wide R-loops in Escherichia coli. Proceedings of the National Academy of Sciences of the USA 110: 258-263.

## Other Publications

1. Gowrishankar $J$ (2012). Public funding for research projects: roles of experts and finance officials in decision-making. Current Science 102: 1499.

# LABORATORY OF COMPUTATIONAL BIOLOGY <br> Computational Studies on Protein Structure, Function and Interactions 

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## Objectives

Studies on protein-protein interaction networks (PPIN):

1. Structural and functional characterization of hubs in human PPIN;
2. Studies on spatio-temporal dynamics of human PPIN; and
3. Analysis of Human-Virus PPI (HU-Vir PPI) network.

Summary of work done until beginning of this reporting year (Upto March 31, 2012)

## 1. Studies on hubs in human PPIN

Studies were carried out to examine the relationship between the extent of splice variation and the average unstructuredness of protein products. The nodes with top $5 \%$ of the splice variant count showed a considerably higher propensity for disorderedness than the rest of the nodes. Similar studies were performed for nodes with top 10\% and $15 \%$ of the splice variant count. We had also analyzed the combined effect of splice variation, domain composition and disorderedness on the degree of the nodes. The nodes with high structural disorderedness showed high degree difference as compared to the nodes with low disorderedness suggesting that the propensity of a node for large number of interactions arises substantially from its structurally disordered splice variants.
2. Studies on tissue-specific human PPINs

We first curated a dataset of human PPIs using BIOGRID, DIP, HPRD, IntAct and MINT. This dataset comprises of 78356 unique undirected interactions for 12142 human proteins. To get tissue-specific PPI network we integrated the PPI physical interaction data with the gene expression data available from microarray (for 70 normal tissues). The construction of tissue-specific network relies on the very fact that protein products of two genes can only interact when both the genes are expressed in the same concerned tissue.

Staff Scientist<br>Senior Research Fellow<br>Senior Research Fellow Senior Research Fellow Junior Research Fellow Junior Research Fellow (Since Jul. 2012)

## 3. Studies on human-virus PPIs

We studied the role of intrinsically disordered proteins (IDPs) in human-viral PPI networks. We found that about $70 \%$ of the human proteins interacting with viral proteins are disordered. We also found that these IDPs are involved in vital cellular processes such as cell cycle regulation, apoptosis, DNA and RNA binding, transcription, translation, protein trafficking, protein degradation pathway and signaling. We merged Human-Virus PPI (Hu-Vir PPI) with Human PPI (Hu-PPI) network and generated a network referred to as Bridged Human Virus PPI network (BHVN). Analyses on BHVN showed that viral proteins act as articulation points i.e. they connect previously unconnected components in the Hu-PPI network. 12 viruses belonging to ssRNA and dsDNA classes formed articulation points. Preliminary studies on viral articulation points indicated that there is similarity between viral articulation points from related viruses.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

Project 1:Structural and functional characterization of hubs in human PPI network

1. As stated in the previous report we examined the relation between the degree of genes/nodes in human protein-protein interaction network with respect to the number of splice variants. We found that on average, hubs in HPRD PPI network have greater number of splice variants than the non-hubs. This observation was confirmed across multiple PPI databases like IntAct, Reactome etc.
2. We further extended our studies to other eukaryotic organisms such as Caenorhabditis elegans, Drosophila melanogaster, Mus musculus and Rattus norvegicus. We found that in all the organisms (Figure 1) except C. elegans hubs have higher number of splice variants.


Figure 1. Splice Variant count in hubs and nonhubs of the Mouse (Mus musculus) PPI Network. Hubs are characterized by the presence of large number of isofroms as compared to nonhubs (Wilcox ranksum test $\mathrm{P}=2.509 \mathrm{e}-05$ ).
3. The existence of a large number of proteins (splice variants) contained in the ENSEMBL database (release 62) has been verified experimentally at the level of the transcripts (mRNA sequences) but not at the level of amino acid sequences. Hence, we repeated our studies with a list of proteins (splice variants) whose existence has been experimentally


Figure 2. Splice Variant count in hubs and nonhubs of the HPRD PPI Network. Only those Splice Variants (proteins) were used for calculation whose existence was experimentally validated. Presence of the proteins in the PRIDE database was considered as a proxy for experimental validation. Hubs are characterized by the presence of large number of isoforms as compared to Non-hubs (Wilcoxon ranksum test $\mathrm{P}=1.612 \mathrm{e}-10$
verified. Figure 2 shows the splice variant count distribution for Hubs and Non-hubs in the HPRD PPI network. As can be seen Hubs have significantly higher number of splice variants than Non-Hubs (Wilcoxon ranksum test $\mathrm{P}=1.612 \mathrm{e}-10$ ). Similar trends were observed for the BioGrid, Intact, HOMOMINT and Reactome databases.
Project 2: Studies on spatio-temporal dynamics of human PPI networks

1. Mapping of tissue-wise expression data pertaining to 70 tissues onto human global PPI resulted in 70 tissue-specific networks. Proteins in the tissue-specific networks were grouped into five distinct classes on the basis of their degree and expression breadth (EB; the number of tissues they are expressed) (Figure 3). They are: 1) House-keeping hubs (HKH): Proteins expressed in at least 60 tissues and also form hubs in all the tissues they are expressed 2) Tissue-preferred hubs (TPH): Proteins expressed in at least 60 tissues but are hubs in at most 10 tissues; 3) Tissue-specific hubs (TSH): Proteins expressed in $<10$ tissues and hubs in all those tissues 4) Housekeeping non-hubs (HKNH): Proteins expressed in $>60$ tissues and nonhubs in all of them and 5) Tissue-specific nonhubs (TSNH) Proteins expressed in <10 tissues and are non-hubs in those tissues. Of the total of 1979 hubs 908 were HKH, 220 were TSH and only 138 were TPH. Among 7610 non hubs 1903 were TSNH and the 3663 were HKNH. Comparative analysis of TSH, TPH and HKH revealed that TSH and HKH exhibit distinct properties as discussed below while, TPH exhibit properties similar to HKH.
2. In-depth analysis of TSH and HKH revealed significant differences between these two groups at sequence, structural and functional levels. TSHs are longer proteins enriched with more disordered regions as compared to HKHs. TSHs are also evolving at faster rates than HKHs. We found that HKHs contain more number of charged and more exposed residues than TSHs. We also looked into the \% of residues in the loop regions and found that TSHs have higher fraction of residues in loop regions as compared to HKHs. We found HKHs have higher fraction of residues in LCRs as compared to TSHs. Despite having similar number of binding surfaces TSHs and HKHs distinctly differ in the number of interactions
they make with other proteins; TSHs are associated with lower degree centrality as compared to HKHs suggesting that TSH are "unsaturated" with regard to their binding capability and are perhaps evolving with regard to their interactions. TSHs are less expressed both at transcript and protein level and also enriched with PEST motifs indicating their easy degradation and tight regulation. TSHs are
we rebuilt our dataset by including them. We further noted some redundancies and duplicate entries in the dataset and hence the entire dataset was subjected to rigorous filtering process to remove obsolete entries, duplicate entries etc. The resultant data comprise of 3392 unique interactions between 270 viral proteins from 74 different viruses and 1736 human proteins. This dataset was used for all our further studies.


Figure 3. Schematic representation of tissue-specific networks and various kind of hubs and non hubs. Hubs are shaded with red colour. Tissue-specific proteins are colored in violet and housekeeping proteins in yellow colour.
mostly secreted, transporters, signaling proteins in contrast to HKHs which are involved in transcription, translation and complex formation. Moreover, HKHs are subjected to more number of protein translational modifications than TSHs which can affect protein conformational or functional specificity facilitating its multi-specificity with partners.

Project 3: Analysis of human and virus proteinprotein interaction (Hu-Vir PPI) networks.

1. As mentioned in the previous report we had identified viral articulation points in Bridged HuVir PPI Network (BHVN) for different viruses. As the protein-protein interaction data became available for Dengue virus, HTLV1 and HTLV2
2. Global survey was conducted on Hu-PPI and Hu-Vir PPIs. 176 human proteins interacting with viral proteins did not have any known human interaction partners in Hu-PPI (Figure 4). This prompted us to look into functions of those 176 peripheral proteins and compare them with functions of peripheral proteins in Hu-PPI network using Gene Ontology (GO). We observed that peripheral components of Hu-PPI network are actually involved in metabolic processes where as those interacting with viral proteins were enriched in functions related to chromatin remodeling, ion binding, stress related pathways and transcription ( $p$-value $<0.05$ ).


Figure 4. Venn diagram showing distribution of human proteins in $\mathrm{Hu}-\mathrm{PPI}$ and Hu -Vir PPI data.
3. The BHVN constructured using the new revised dataset was used for identifying viral articulation points. Articulation points are the nodes whose removal results in an increase in the number of components in the network. They act as bridging elements between two components and add to the complexity of the network. These articulation points are conserved i.e., similar proteins from related viruses form articulation points when mapped on to Hu-PPI network. Functional domain analysis of such similar proteins suggested that they have same functional domains. Functional annotation studies showed that viruses connect metabolic pathways to PPI network and hence seem to take over the regulation of metabolic pathways.
4. HIV1 was found to be connecting highest number ( 101 proteins) of peripheral nodes to the Hu-PPI network hence further analysis was carried out on HIV1. For any functional connection to occur between two proteins, they should be in close proximity with each other and this is achieved by their co-expression in the same sub-cellular localization. In order to study functional relevance of bridged connections via viral articulation points, gene expression data was downloaded from NCBI Gene Expression Omnibus (GEO). The median of all the expression values was used as the cut off for determing expression or no expression of various genes. All possible pairs of proteins bridged by HIV1 articulation points were made between bi-connected nodes which are part of giant component and articulation point connected peripheral nodes. Then each pair was given score 1 if the proteins in the pair are co-expressed in a tissue. All the coexpressed protein pairs were were taken and their subcellular localization was predicted using LOCATE (http://locate.imb.uq.edu.au/). Further studies are underway.

## Future plans and directions

1. Integration and analysis of human nsSNP data on protein-protein interaction networks.
2. Further analysis of viral-human bridge PPI network.
3. Studies on tissue-wise PPI networks integrated with drug-protein interaction data.
4. Further studies on structural and functional characterization of hubs in HPPIN.

## Publications

1. Acharya $V$ and Nagarajaram HA (2012). Hansa: an automated method for discriminating disease and neutral human nsSNPs. Human Mutation 33: 332-337.
2. Bashyam MD, Chaudhary AK, Reddy EC, Reddy V, Acharya V, Nagarajaram HA, Devi ARR, Bashyam L, Dalal AB, Gupta N, Kabra M, Agarwal M, Phadke SR, Tainwala R, Kumar R and Hariharan SV (2012). A founder ectodysplasin A receptor (EDAR) mutation results in a high frequency of the autosomal recessive form of hypohidrotic ectodermal dysplasia in India. British Journal of Dermatology 166: 819-829.
3. Bashyam MD, Chaudhary AK, Sinha M, Nagarajaram HA, Devi ARR, Bashyam L, Reddy EC and Dalal A (2012). Molecular genetic analysis of MSUD from India reveals mutations causing altered protein truncation affecting the C-termini of E1 $\alpha$ and E1 $\beta$. Journal of Cellular Biochemistry 113: 3122-3132.
4. Bashyam MD, Purushotham G, Chaudhary AK, Rao KM, Acharya V, Mohammad TA, Nagarajaram HA, Hariram V and Narasimhan C (2012). A low prevalence of $M Y H 7 / M Y B P C 3$ mutations among familial hypertrophic cardiomyopathy patients in India. Molecular and Cellular Biochemistry 360:373-382.
5. Kumar P and Nagarajaram HA (2012). A study on mutational dynamics of simple sequence repeats in relation to mismatch repair system in prokaryotic genomes. Journal of Molecular Evolution 74: 127-139.
6. Sinha A and Nagarajaram HA (2013). Effect of alternative splicing on the degree centrality of nodes in protein-protein interaction networks of Homo sapiens. Journal of Proteome Research 12: 1980-1988.

## Other publications

1. Acharya V and Nagarajaram HA (2013). Response to: Statistical analysis of missense mutation classifiers. Human Mutation 34:407.

## LABORATORY OF MOLECULAR CELL BIOLOGY

Signal Transduction Pathways in Macrophages and Host-Pathogen Interaction in Tuberculosis

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Staff Scientist<br>Senior Research Fellow (Till Apr. 2012)<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Junior Research Fellow<br>Junior Research Fellow<br>Scientist<br>Senior Technical Officer<br>Research Associate<br>Research Associate<br>Research Associate<br>Project-Junior Research Fellow (Since Aug. 2012)<br>Project Assistant (Since May 2012)<br>NCCS, Pune<br>NIN, Hyderabad<br>Mahavir Hospital, Hyderabad \&<br>BPRC, Hyderabad<br>IIT, Delhi

## Objectives

1. Signal transduction pathways in macrophages regulating its innate-effector immune responses; and
2. Studying how various candidate proteins of Mycobacterium tuberculosis interfere with macrophage signaling cascades to modulate host's protective responses against the bacilli.
Project I: Role of PPE18 protein in intracellular survival and pathogenicity of $M$. tuberculosis
Summary of work done until the beginning of this reporting year (upto March 31, 2012)

PPE18 (Rv1196), also known as Mtb39a, a member of the PPE family, is expressed more in Mycobacterium tuberculosis (Mtb) as compared to $M$. bovis. Also, comparative genome analyses of the avirulent H37Ra strain vs virulent H37Rv strain revealed presence of 53 insertions and 21 deletions in H37Ra relative to H37Rv. Interestingly, PPE18 harbored one of those deletions in H37Ra, indicating that this gene may be pathophysiologically important. Previous work by us (Nair et
al. [2009] J. Immunol. 183: 6269; Nair et al. [2011] J. Immunol. 186: 5413-5424) documented that PPE18 binds to toll like receptor (TLR) 2 on macrophages and upregulates IL-10 cytokine production which favors a Th2-type response. Also, its interaction with TLR2 leads to phosphorylation of the SOCS3 protein which then physically interacts with the $\mathrm{I}_{\mathrm{K} B} \alpha-\mathrm{NF}-\kappa \mathrm{B} / \mathrm{c}-\mathrm{rel}$ complex preventing nuclear translocation of p50 and p65 NF-кB and c-rel transcription factors. As a consequence, there is a downregulation of transcription of NF-кB-regulated genes like IL-12 and TNF- $\alpha$. We now aim to understand whether PPE18 plays any role in the survival and multiplication of Mtb bacilli during infection.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

PPE18 confers a growth advantage to Mtb in vivo in mouse model
To understand the role of PPE18 in Mtb virulence in vivo, C57BI/6 mice were infected with either wildtype (WT) or ppe18knock-out (KO) strains of Mtb via the aerosol route and the bacterial burden was
estimated in lung, liver and spleen of infected animals at 3 different time points ( 3 weeks, 6 weeks and 9 weeks) after infection. The aerosol infection deposited 80~130 colony forming units (CFUs) per lung (as assessed by counting CFUs in two infected mice per Mtb strain at day 1 postinfection). Infection through aerosol deposits Mtb directly into the lungs and hence is considered to be closest to the physiological mode of infection. Lungs being the primary site of infection showed maximum CFUs at all the time points examined (Figure 1A). In mouse model upon infection, bacteria are known to disseminate from lungs to liver and spleen. We observed a steady rise in the bacterial load in all the organs at 3 week after aerosol infection and then a decrease at 6 week and 9 week post-infection. Interestingly, the number of ppe18 KO bacteria remained significantly less in all the organs at almost all the time points investigated (Figure 1A). In the lungs of ppe18KOinfected mice, the mean bacterial counts ( $\pm$ SEM) were significantly lower at 3 weeks post-infection as compared to those of infected with WT Mtb strain and this trend continued to later time points also ( 6 weeks and 9 weeks) (Figure 1A). Similar observations were made in liver as well as in spleen (Figure 1A). PPE18 has previously been reported to be non essential for bacterial growth in vitro. Our results indicate that PPE18 probably plays a role in replication and survival of Mtb in vivo and thus, may be a candidate virulent factor.

Mice infected with ppe18 KO strain show a reduced degree of inflammation and tissue damage and tuberculosis induced fatality as compared to mice infected with WT strain
We next examined the tissue damage in lung, liver and spleen in mice infected with WT and ppe18 KO Mtb strains in vivo by histopathological analyses. The extent of inflammation and tissue damage due to infection as seen in the hematoxylin and eosin (H\&E) stained sections of lung and liver from mice infected with WT Mtb was found to be markedly pronounced than that observed in mice infected with the ppe18 KO Mtb (Figure 1, B-D). Mice infected with ppe18 KO had more intact alveolar spaces while mice infected with WT Mtb almost had none, especially at 21 and 60 weeks post-infection (Figure 1B). The lesions and tissue damage observed in the WT Mtb-infected animals were graded 4 (marked with 51-75\% tissue affected) and 5 (severe with 76-100\% tissue affected) and those in the ppe18 KO-infected animals were graded 3 (moderate with 26-50\% tissue affected),

60 weeks post-infection (Figure 1B). A similar trend was observed in the liver (Figure 1C). Effect of infection was not observed in spleen of both WTand ppe 18 KO Mtb-infected mice sacrificed at 3 weeks. Histiocytosis or accumulation of macrophages in spleen was observed only at 21 and 60 weeks post-infection in mice infected with WT Mtb strain, however, the spleen tissue structure of ppe18 KO strain-infected mice appeared to be normal (Figure 1D).
Our observations thus indicated that in comparison to the WT, the ppe18 KO strain elicited a reduced and delayed inflammatory response in lung, liver and spleen of the infected mice. To understand the total effect of in vivo growth and inflammation, survival of mice infected with WT and ppe18 KO strains of Mtb was monitored over a prolonged period of time. No deaths were registered in the group of mice infected with the ppe18 KO strain during the entire study period of 60 weeks, however, in the group of mice infected with the WT Mtb, survival rate had dropped to $25 \%$, 60 weeks postinfection (Figure 1E). Also, mice infected with ppe18 KO strain visibly appeared healthier. The percentage increase in the weight of mice infected with ppe18 KO strain 9 weeks after infection was $55 \pm 2 \%$ compared to the $31.9 \pm 5 \%$ increase in the weight of mice infected with the WT strain. We are presently characterizing PPE18-induced modulation of the immune responses that contribute to Mtb virulence in mice.

Project II: Understanding the role of $M$. tuberculosis hsp60 as Th1/Th2 immunomodulator

Summary of work done until the beginning of this reporting year (upto March 31, 2012)

Macrophage is known to regulate T-cell effector responses as Th1 or Th2. While Th1 is protective, Th2 favors Mtb survival. We have earlier demonstrated a novel role of the M. tuberculosis heat shock protein 60 (Mtbhsp60, Cpn60.1) protein to favor Th2-environment by modulating the surface TLR2 population in macrophages (Khan et al. [2008] Cell Microbiol. 10: 1711). To understand in detail how Mtb proteins influence the TLR-signaling to modulate macrophage effector-APC functions and the Th balance, we have demonstrated that Mtbhsp60 interacts with both TLR2 and TLR4, but its interaction with TLR2 leads to clathrin-dependent endocytosis resulting in an increased activation of p38 MAPK and IL-10 cytokine that favors Th2. In contrast, upon interaction with TLR4, Mtbhsp60


Figure 1. PPE18 protein plays an important role in intracellular survival and pathogenicity of Mycobacterium tuberculosis in mice. C57BL/6 mice were infected aerogenically with a low dose of either WT or ppe18 KO strains of Mtb. At different time points post infection, mice were sacrificed and CFU counts were measured in lung, liver and spleen (A). Data are mean $\pm$ SEM of results for five mice per group for each time point. Liver (B), Lung (C) and Spleen (D) sections from mice infected with either WT (left panel) or ppe18 KO (right panels) strains of Mtb were stained with Hematoxylin and eosin (H\&E) at different time points post infection. Photographs of representative sections from 2 mice visualized at 40X magnification are shown. Arrows indicate the sites of lymphocytic infiltration. In another set of experiment, survival of C57BL/6 mice ( $n=8$ ) following a lowdose aerosol infection with either WT or ppe18 KO strains of Mtb was monitored for 420 days post infection (E).
remains predominantly localized on the cell-surface due to reduced endocytosis of the protein, that leads to p38 MAPK activation and poorer IL-10 production but triggers ERK $1 / 2$ and TNF- $\alpha$ production. These results were further confirmed using macrophages from TLR2 and TLR4 knockout mice. Inhibition of endocytosis by MDC increased cell surface accumulation of Mtbhsp60 and compromised its ability to induce IL-10. In such situation, we observed an increase in the TNF- $\alpha$ production.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

The E. coli heat shock protein 60 (Ecolihsp60) is retained mainly on the macrophage surface upon interaction with either TLR2 or TLR4 and triggers induction of TNF- $\alpha$
Although the protein sequence of Ecolihsp60 is significantly similar to that of Mtbhsp60, the biochemical features of Mtbhsp60 deviate significantly from the characteristic properties of
the Ecolihsp60. The Mtbhsp60 exists in a lower oligomeric state as compared to its E. coli counterpart due to substitutions in some crucial interface residues required to stabilize its intersubunit interactions and also lacks ATPase activity. Also at the structural level, Mtbhsp60 significantly deviates from Ecolihsp60 (Figure 2A). Thus, in the next experiment we investigated whether Ecolihsp60 modulates TNF- $\alpha / \mathrm{IL}-10$ cytokines post TLR interaction in a different fashion. We observed that although Ecolihsp60 interacted with both TLR2 and TLR4 (Figure 2B), but unlike Mtbhsp60, it failed to undergo endocytosis through TLR2 (Figure 2C) and preferentially induced TNF- $\alpha$ through both TLR2
and TLR4 (Figure 2D) but very little IL-10. In the presence of isotype-matched antibodies, the level of TNF-a was significantly higher as compared to that of TLR2 or TLR4 alone and almost summed up the levels of TNF- $\alpha$ produced together by these receptors (Figure 2D). These observations led us to speculate that the cellular localization of Mtbhsp60 post-binding to TLRs activates different signaling cascades that finally dictate the type of inflammatory response to be produced in macrophages. In the near future we will focus on identifying how the Mtbhsp60 protein targets the TLR-signaling to influence macrophage APC functions and T-cell immune responses.


Figure 2. Interaction of Ecolihsp60 either with TLR2 or TLR4 results in induction of TNF- $\alpha$. The secondary structure of the Mtbhsp60 protein model, predicted based on homology modeling with Modeller software is shown (Green) after its energy minimization, the solved crystal structure of Ecolihsp60 obtained from Protein Data bank (chain-A of PDB-code:2EU1) is displayed (Blue) and the superimposed structures of Mtbhsp60 and Ecolihsp60 is shown (A). In the next experiment, PMA-differentiated THP-1 macrophages were pre-treated with anti-TLR2 mAb or anti-TLR4 mAb or isotype-matched control Ab and then incubated with $10 \mathrm{mg} / \mathrm{ml}$ of FITC labeled Ecolihsp60 at $4^{\circ} \mathrm{C}$ for 30 min . The cells were fixed and the fluorescence was measured by flow cytometry (B). In another experiment, PMA-differentiated THP-1 macrophages were pre-treated with anti-TLR2 mAb or anti-TLR4 mAb or isotype-matched control antibody for 1 h , followed by incubation with Ecolihsp60-FITC ( $10 \mathrm{mg} / \mathrm{ml}$ ) at $37^{\circ} \mathrm{C}$ for 15 min . The cells were fixed and endocytosis of the protein was assessed by confocal laser-scanner microscopy (C). Also, PMAdifferentiated THP-1 macrophages were pre-treated with $10 \mu \mathrm{~g} / \mathrm{ml}$ of anti-TLR2 mAb or anti-TLR4 mAb or isotypematched control antibody for 1 h and then incubated with $3 \mu \mathrm{~g} / \mathrm{ml}$ Ecolihsp60. TNF- $\alpha$ levels were quantified after 48 $h$ in different culture supernatants by EIA (D). Data are representative of Mean $\pm$ SD of three different experiments.

## Publications

1. Akhter Y, Ehebauer MT, Mukhopadhyay S and Hasnain SE (2012). The PE/PPE multigene family codes for virulence factors and is a possible source of mycobacterial antigenic variation: perhaps more? Biochimie 94: 110116.
2. Bhat KH, Ahmed A, Kumar S, Sharma $P$ and Mukhopadhyay S (2012). Role of PPE18 protein in intracellular survival and pathogenicity of Mycobacterium tuberculosis in mice. PLoS One 7: e52601.
3. Bhat KH, Chaitanya CK, Parveen N, Varman R, Ghosh S and Mukhopadhyay S (2012).

Proline-Proline-Glutamic Acid (PPE) protein Rv1168c of Mycobacterium tuberculosis augments transcription from HIV-1 Long Terminal Repeat promoter. Journal of Biological Chemistry 287: 16930-16946.
4. Mukhopadhyay S, Nair S and Ghosh S (2012). Pathogenesis in tuberculosis: transcriptomic approaches to unraveling virulence mechanisms and finding new drug targets. FEMS Microbiology Reviews 36: 463-485.
5. Bhat KH, Das A, Srikantam A and Mukhopadhyay S. PPE2 protein of Mycobacterium tuberculosis may inhibit nitric oxide in activated macrophages. Annals of the New York Academy of Sciences (In press).

## LABORATORY OF NEUROSPORA GENETICS

## What keeps the Neurospora genome repeat-free?

Faculty<br>Other Members<br>DP Kasbekar<br>A Sheeba<br>K Sreethi Reddy

Haldane Chair (Since Jul. 2012)<br>Technical Officer<br>Technical Assistant (Since Oct. 2012)

## Objectives

Meiotic silencing by unpaired DNA (MSUD) is a presumed RNAi-mediated elimination of the transcripts of any Neurospora crassa gene that is not properly paired with a homolog in meiosis. Recent results from our laboratory, obtained in the course of constructing a recombinant inbred line (RIL), have suggested that inbreeding can affect MSUD in a genotype-independent manner. The major objective of the research in the past year was to verify this result, and to complete making the RIL.

In crosses of the standard laboratory Oak Ridge (OR) wild type strains with the $:: B m /$ and ::mei-3 tester strains, MSUD of the bml ( $\beta$-tubulin) and mei-3 genes causes dramatic ascus-development abnormalities. MSUD does not occur in homozygous tester $A \times$ tester a crosses, nor in crosses of the testers with the semi-dominant suppressors of MSUD (e.g., Sad-1, Sad-2), and the asci develop normally. Presumably, the suppressor alleles prevent the proper pairing of their wild-type homologues, and thus induce them to autogenously silence themselves. Wild-isolated $N$. crassa strains were classified into three types based on the phenotype of their crosses with the testers. In crosses with "OR" and "Sad" type strains the $\mathrm{Bm} / \mathrm{l}$ and mei-3 genes are, or are not, silenced. Whereas in crosses with "Esm" type, bml was silenced but not mei-3+ We proposed that $\mathrm{bm} /$ is more sensitive to silencing than mei-3, and that sequence polymorphisms between the OR-derived tester and Sad and Esm genomes might cause one or more genes essential for meiotic silencing to become unpaired and silence itself, thus shortening the duration of silencing. Thus, MSUD could become very fleeting in the cross with Sad type, of intermediate duration in the cross with Esm type, and persist throughout the cross only with OR type (Figure 1). In which case, if a new tester is made in the genetic background of a Sad type strain, and a cross is performed that is heterozygous for the tester allele but isogenic for
the rest of the genome, then we would expect to see MSUD. To test this prediction we decided to construct an isogenic recombinant inbred line from the Sad type wild strains Bichpuri-1 a and Spurger3 A.
The Sad type wild strains, Bichpuri-1 a (B) and Spurger-3 $A(S)$, were crossed with each other to produce generation f 1 . In the f 1 , and in each successive generation, pairs of sibling strains of opposite mating type were crossed to produce the next generation. We confirmed that in a line the later generations are more isogenic than the earlier generations, and that different lines become isogenic for different genomic segments from the $B$ and $S$ strains. After 10 generations of sibling crosses we generated a pair of isogenic mat $A$ and mat a strains that is now ready to be used to make the tester strain. Although Bichpuri-1 $a$ and Spurger-3 A, and most of their f1 progeny were consistently and reproducibly Sad type in crosses with the $:: B m l^{r}$ and $::$ mei-3 testers, the later generation strains of each line showed Sad, Esm, or Sad / Esm types with variable expressivity. Since all later generation genotypes are, in principle, obtainable in the f1, the observed transition from an apparently stable Sad phenotype to an apparently unstable Sad/Esm phenotype is probably not due to genotype differences between the generations. Therefore, it appears that the Sad versus Esm difference can have a genotypeindependent basis.
Summary of work done (prior to the faculty member joining CDFD i.e., before July 2012)
Only one exceptional Neurospora crassa strain contains transposons. This strain, isolated from Adiopodoume in West Africa, contains the retrotransposon Tad, whereas all the other (>1000) Neurospora strains examined (by J.A. Kinsey and colleagues) contained only relics of Tadinactivated by RIP. No other transposon is known in Neurospora. RIP is a mutational process that acts during a sexual cross and targets $\mathrm{G}: \mathrm{C}$ to $\mathrm{A}: T$


Figure 1. Two wild-isolated $N$. crassa strains induce different phenotypes in their crosses with the ascusdominant Dip-1 mutant. A rosette of asci from the cross of the Dip-1 a strain (FGSC 9536) with the Roanoke-1m A (FGSC 2227) strain (A) shows the Dip-1 mutant phenotype, namely, several asci with two to four large ascospores instead of the normal eight, but in the rosette from the cross of the Dip-1 a strain with the wild-isolated Klong Rangsit (FGSC 6488) strain (B) almost all asci are eight-spored.
hypermutation to repeated DNA sequences. We developed an assay for RIP, and using it we showed (1) the RIP machinery is titrable by chromosome segment duplications $(D p)>300 \mathrm{kbp}$; and (2) the Adiopodoume strain is one of seven wild-isolated Neurospora strains identified to exert a dominant RIP suppressor phenotype. The Adiopodoume strain contains $\sim 40$ copies of Tad, each $\sim 7 \mathrm{~kb}$, therefore this ~280 kbp of duplicated DNA might contribute to titration of the RIP machinery, whereas Tad would remain vulnerable to RIP in low copy strains. The studies leading to 1 also defined the breakpoints of several $D p$-generating chromosome rearrangements onto the genome sequence. We pioneered transformation of a closely related pseudohomothallic species, $N$. tetrasperma, to initiate the genetic analysis of diplophase-specific processes such as RIP and MSUD. In related work, we identified Fmf-1p as a master regulator of sexual differentiation and mapped the dow mutation.
In other studies, we found that LBR (a vertebrate nuclear membrane protein that tethers chromatin to nuclear lamina) has sterol biosynthetic activity, and we identified its essential amino acid residues by site-directed mutagenesis. Our laboratory is unique in having studied the response of dictyostelids (free-living soil amoebae that feed on bacteria) to antimicrobial isoflavonoids made by leguminous plants. Based on these studies we
proposed a novel plant-microbe interaction wherein leguminous plants use isoflavonoids to recruit dictyostelids to clear bacteria from the vicinity of root lesions.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
The fraction of Esm type progeny increases with inbreeding: Both the $B$ and $S$ strains consistently and reproducibly displayed the Sad phenotype. Seventy-six progeny strains in the f1, and 10-20 progeny strains in each succeeding generation were crossed with the testers. Only two ( $2.6 \%$ ) of the f1 strains tested were Esm type, and the rest were Sad type. In contrast, a larger fraction of progeny were Esm type in the later generations. Esm types represented $14.5 \%$ of the f2+f3+f4 ( $\mathrm{N}=191$ ), and $57.5 \%$ in the $\mathrm{f} 5+\mathrm{f} 6+\mathrm{f} 7+\mathrm{f} 8$ ( $\mathrm{N}=94$ ). None of the 357 strains examined was OR type. To rule out the possibility that the observed variation in phenotype is due to a failure to control for variations in temperature, age of the crosses at analysis, variation in media formulation, and clerical errors; we performed a blind experiment in which 48 f 1 strains and 49 f 5 strains were examined afresh in parallel. The frequency of Esm types was $4 / 48$ in the f 1 and $10 / 49$ in the 55 . These results allow us to reject the null hypothesis that the f 1 and $f 5$ have the same Esm frequencies ( p < 0.045 , one-tailed $z$-test).

Later generation strains also show an apparently unstable Sad/Esm phenotype: Twenty-five Sad type f1 progeny were re-examined by re-crossing them with the testers and all rescored as Sad type. However, re-examination of progeny from the later generations showed that a subset that had scored as Sad type, could subsequently retest as Esm type, and others that scored as Esm type, could retest as Sad type. These results suggested that although the $B$ and $S$ wild strains and most of their f1 progeny were stably Sad type, the later generations have a more variable phenotype and upon re-assay at least a subset of strains can switch between the Sad or Esm types.

Significance of our results: The change from an apparently stable Sad phenotype in the f1 to an apparently unstable Sad/Esm phenotype in later generations is not easily attributed to genetic differences between the strains of the different generations. First, the f1 and later generations (f2, f 3 , etc.) are all genetically equivalent, in that, they contain about equal contributions of the B and S genomes, and any genotype obtained in a later generation is, in principle, obtainable in the f1. Second, the later generations are more isogenic than the earlier ones (e.g., f1 siblings differ in $50 \%$ of their genomes, whereas f8 siblings differ in $<0.5 \%$ ), therefore one would expect a more uniform phenotype among the f8, yet they appeared more variable.

Near-isogenic crosses are likely to be an exception in $N$. crassa since it is an out-crossing heterothallic species. Therefore when such a cross is made in the laboratory, it might experience exceptionally little (or no) meiotic unpairing, and could trigger positive feedback mechanisms to enhance the detection of unpairing and induce silencing. For instance, assembly of the Sad-2 -containing perinuclear complex, in which aberrant RNA molecules derived from unpaired genes are converted into dsRNA may be made more efficient by having the proto-structures for assembling such complexes become long-lived and remain associated with the zygote nuclei and their descendents. That is, the proto-complex might undergo perdurance along with the progeny nuclei. During vegetative growth of the progeny, a subset of mitotic nuclei might retain these proto-structures all the way through to the next cross, whereas they might be lost from other nuclei. Consequently, crosses involving nuclei that retain such structures would show an increase in meiotic silencing
strength, and those involving nuclei that lost these structures would show a relative decrease in meiotic silencing strength. Thus the variable perdurance might account for the variable expressivity in meiotic silencing strength in crosses with the testers. An alternative model to account for the apparently genotype-independent change from a stable Sad phenotype to an unstable Sad/ Esm phenotype is that bml expression levels (or stability levels, turnover rates, etc.) might decrease with inbreeding, independent of effects of meiotic silencing, thereby making the $\mathrm{bm} /$ meiotic silencing test more sensitive in crosses with these strains (since the inbred strains have lower $\mathrm{bm} /$ levels, silencing of $\mathrm{bm} /$ is easier). As for the unstable Esm/ Sad phenotype, it could be that $\mathrm{bm} /$ expression levels have decreased to near a key threshold in the inbred strains. Below this threshold, one would observe the phenotype as Esm, above this threshold; one would observe the Sad phenotype. Because any one of these 'bm/ near threshold' strains may have slightly different bm/ expression levels from cross to cross, these strains could be observed to switch from Esm to Sad (or vice versa) from cross to cross. The latter model raises the question of why expression of $\mathrm{bm} /$ is depressed by inbreeding.

## Publications

1. Kasbekar DP (2013). Neurospora duplications and genome defense by RIP and meiotic silencing. Neurospora: Genomics and Molecular Biology. Editors: DP Kasbekar and K McCluskey, Caister Academic Press, Norfolk, UK. Pages 109-127.

## Other Publications

1. *Kasbekar DP (2012). Lymphohematopoietic licence: sterol C-14 reductase activity of lamin B receptor (Lbr) is essential for neutrophil differentiation. Journal of Biosciences 37: 199-201.
2. Kasbekar DP (2012). Green-carding the referee and Haldane's spell. Journal of Biosciences 37: 579.
3. Kasbekar DP (2012). The Sad paradox: mutations with dominant and recessive phenotypes. Journal of Biosciences 37: 933936.
4. Kasbekar DP (2013). Myth versus mutant: story of 0 . Journal of Biosciences 38: 1 .

* Work done elsewhere


# LABORATORY OF MAMMALIAN GENETICS 

## Epigenetic Mechanisms Underlying Developmental Pathways

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|  | Amitava Basu |
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| Collaborators | Gayatri Ramakrishna |
|  | Shekhar Mande |
|  | Rakesh Mishra |
|  | Vinay K Nandicoori |

Project 1: DNMT3L: Role in development
Summary of work done until the beginning of this reporting year (upto March 31, 2012)

We had previously reported analysis of the regulatory mechanisms underlying the transcription of DNMT3L in Drosophila. Transgene reporter assay in Drosophila was performed wherein the promoter region flanked by loxP sites, was inserted upstream of the hsp70 promoter driven mini-white reporter gene containing P -element vector pCaSpeR . The analysis of the reporter gene transcription showed that the presence of DNMT3L promoter/Exon1 region in the reporter construct causes repression of the GFP expression.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

Role of DNMT3L promoter in regulation of its transcription
For functional analysis of the DNMT3L promoterExon 1 CpG island that had previously been shown to be hypomethylated in cervical and ocular cancer samples, we performed transient transfection assay in mammalian HeK cell line with two overlapping fragments from this region. Approximately 70\% decrease in expression of the GFP reporter gene in the mammalian cells was observed for both the versions of the DNMT3L Promoter-Exon 1 CpG island and in both orientations. The extent of the inhibition of GFP expression in presence of this region was similar to that observed for H19 ICR, a

Staff Scientist<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Senior Research Fellow<br>Junior Research Fellow<br>Technical Officer<br>Project Associate<br>Project-Junior Research Fellow<br>Project-Junior Research Fellow (Till Jan. 2013)<br>CDFD, Hyderabad \& ILBS, New Delhi<br>NCCS, Pune<br>CCMB, Hyderabad<br>NII, New Delhi

known transcriptional repressor (Figure 1). The inhibitory nature of this CpG island was found to be due to its interaction with Polycomb proteins that are known to inhibit transcription. Concordant with the observation of its interaction with Polycomb proteins, our results also showed that this region adopts an inactive chromatin conformation in both the Drosophila transgene reporter gene assay and the mammalian transient transfection assay.
Project 2: Host epigenetic response to infection
Summary of work done until the beginning of this reporting year (upto March 31, 2012)
During the interaction with Mycobacterium tuberculosis, not only does the host cell reprogram its epigenetic markings at several loci in the genome so that the affected gene may be appropriately modulated but also the mycobacterium can produce molecules that interact or influence the effectors of host epigenetic modifications. We have previously initiated studies to identify (i) putative DNA methyltransferases in M. tuberculosis and (ii) DNA methylation changes in the host genome.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

To influence the host epigenetic circuitry, the mycobacterial factor could be (i) a DNA methyltransferase; (ii) a histone modifier (histone methyltransferase, acetyltransferase, etc.); (iii) a protein that interacts with DNA; (iv) a protein that


Figure 1. Functional analysis of the DNMT3L promoter-Exon 1 CpG island by reporter gene assay in transiently transfected mammalian cells. A. Graphical representation of the various constructs transfected into HEK293 cells. Shown for each construct is the reporter AcGFP gene, its CMV promoter, the selection Kan ${ }^{\text {R/ }}$ $\mathrm{Neo}^{R}$ marker, the various DNMT3L promoter-Exon 1 CpG island fragments and the control Chr1 fragment and the H19 ICR inserted upstream of the CMV promoter. B. Relative transcriptional level analysis of the reporter AcGFP gene in the various constructs by Real-Time PCR. C. A representative Western Blot analysis showing AcGFP protein expression for HEK293 cells transfected with the various constructs mentioned in A. Luciferase activity for the pG5luc vector co-trasfected with the above mentioned constructs is shown below the â-TUBULIN panel. D. Relative AcGFP protein expression levels for the cells transfected with the various constructs. Error bars represent standard error of mean. Asterisks indicate significant difference (Student's $t$ test, * $-\mathrm{p}<0.05$, ** $-\mathrm{p}<$ 0.01 , *** $-\mathrm{p}<0.001$ ).
interacts with histones, or (v) a protein that interacts with chromatin modifiers. Based on a combination of bioinformatic analysis and Co-Immunoprecipitation assay we have identified a few mycobacterial genes that could be putative DNA demethylase or Histone methyltransferases. Further experiments are being carried out to characterize their role in modulating the host epigenetic circuitry.
We had previously reported identification of mycobacterial proteins that can methylate DNA. One of these proteins was found to be secreted out of BCG and our transient transfection assay showed that it can localize to the Thp1 nucleus. By performing deletion experiments a nuclear localization signal has been identified in the Cterminus of this protein (Figure 2). Site-directed mutagenesis experiments are underway to pin point the NLS sequence motif. We plan to identify the role of this protein during infection and its correlation with the host epigenetic circuitry.


Figure 2. Subcellular localization of the putative DNA methyltransferase Mtbmeth1 in mammalian cells after transient transfection. Constructs of Mtbmeth1 and its deletion mutant in fusion with GFP were transiently transfected in HeK cells and the subcellular localization was examined under a confocal microscope. Mtbmeth1 14 and Mtbmeth $1 \Delta \mathrm{~N}$ denote the C -and N -terminal deletion of Mtbmeth1 respectively. Bar is $\sim 5 \mu \mathrm{M}$.


Figure 3. Expression analysis of the various mammalian Histone methyltransferases and demethylases upon infection with BCG. Semi-quantitative RT-PCR for the various Histone methyltransferases (KMTs) and demethylases (KDMs) was performed on uninfected and BCG-infected Thp1 cells. Time points mentioned in the labels on the top refer to the number of hours the cells were left in culture after the BCG infection. As a control Thp1 cells were also maintained in culture without infection for the same time.

In order to respond to the infection by $M$. tuberculosis, the host cells would have to reprogram the epigenetic markings at several loci in the genome so that the affected gene may be appropriately modulated. It has been our endeavor to examine these epigenetic changes and identify the genetic loci where these changes are brought about. This, we believe, will provide us important evidence about the genes in the host that might be participating in a response to mycobacterial infection. Previously, we had performed examination of the DNA methylation changes upon infection in the treated macrophage cell line, Thp1. The loci that showed changes in DNA methylation upon infection are being validated at present. We
are also in process of examining the expression level of several Histone methyltransferases (KMTs) and Histone Demethylases (KDMs) by RT-PCR and Western in BCG infected Thp1 cells. As can be seen from Figure 2, a few histone methyltransferases and demethylases do show changed expression in BCG infected Thp1 cells. The validation of the changed expression level and its correlation with mycobacterial infection is being studied at present.

## Publications

1. Gokul G and Khosla S (2012). DNA methylation and cancer. Subcellular Biochemistry 61: 597-625.

## LABORATORY OF MOLECULAR ONCOLOGY Genomics and Molecular Genetics of Cancer and Genetic Disorders

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## Objectives

1. Identification and characterization of important deregulated genes/pathways in cancers prevalent in India; and
2. Identification and characterization of disease causing mutations in genetic disorders.

Summary of work done until the beginning of this reporting year (Upto March 31, 2012)

Colorectal Cancer (CRC): 50\% of early onset rectal (but not colon) cancer samples did not

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AIMS, Cochin
CHG, Bengaluru
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harbour deregulated canonical Wnt signalling or mismatch repair (MMR) inactivation. Interestingly, despite the absence of canonical Wnt activation, the early-onset sporadic rectal cancer (EOSRC) samples exhibited significant chromosomal aberrations. One such aberration at 5 p 11.2 (including $h T E R T$ ) presented as a gain in Wnt- and loss in Wnt+ samples.
Pancreatic Cancer (PaCa): We characterized a possible tumour suppressor function for ARID1B encoding a member of the SWI/SNF chromatin
remodelling complex, which was found to be deleted in several PaCa cell lines and xenografts. Permanent transfectancts generated using ARID1B cDNA in MiaPaCa2 PaCa cell line (harbouring ARID1B homozygous deletion) exhibited reduced colony formation ability in liquid media and soft agar, though there was no difference in conventional growth, apoptosis and cell cycle analyses. In addition, preliminary analysis revealed hypermethylation of ARID1B promoter CpG island in PaCa cell lines. ARID1B exhibited significantly reduced expression in tumour when compared to matched normal tissue, as determined by immunohistochemistry ( $\mathrm{IHC} \mathrm{)} \mathrm{on} \mathrm{a} \mathrm{PaCa} \mathrm{tissue}$ microarray (TMA).

Phenylketonuria (PKU): Molecular genetic analysis of PKU in seven Indian patients revealed complete absence of Phenylalanine hydroxylase ( $P A H$ ) missense mutations; four novel $P A H$ mutations were identified.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

CRC: We performed genome-wide DNA copy number and transcript profiling on additional microsatellite stable (MSS) EOSRC samples, stratified for Wnt status. Interestingly, several distinct copy number alterations, validated by quantitative PCR (Q-PCR), were identified in the Wnt- vis-à-vis Wnt+ samples. Analysis of array transcript profiles generated for 36 Wnt- and 16 Wnt+ EOSRC samples using Significance Analysis of Microarrays and Gene Set Enrichment Analysis (GSEA) validated enrichment of the Wnt/ $\beta$-Catenin gene set in Wnt+ samples. Surprisingly, non-canonical Wnt pathway genes were enriched in a subset of Wnt- samples (Figure 1A); this observation was validated using quantitative-reverse transcription PCR (Q-RT-PCR) (Figure 1B). This is the first report of possible presence of non-canonical Wht driven tumours in rectal cancer.


Figure 1. Gene set enrichment analysis identifies significant upregulation of non-canonical Wht pathway genes specifically in Wnt- EOSRC (Panel A). The differential expression of non-canonical Wnt pathway genes is validated using Q-RT-PCR (Panel B).

PaCa: Azacytidine and TSA treatment resulted in significant elevation in ARID1B transcript levels only in PaCa cell lines that exhibited reduced expression, corroborated by identification of extensive methylation of promoter CpG island using bisulphite sequencing (Figure 2A). ARID1B expressing PaCa cells exhibited significantly increased senescence-associated â-galactosidase activity (Figure 2B). A pool of several ARID1B permanent MiaPaCa2 clones exhibited significantly reduced colony formation ability in liquid culture when compared to pooled vector clones, a difference
not observed with clones generated in Panc1 (a PaCa cell line harboring elevated ARID1B levels); thus validating results obtained with individual MiaPaCa 2 clones. The loss of ARID1B expression in PaCa (determined by IHC on a TMA) was associated significantly with advanced tumour stage ( $p=0.0185$, Fisher's exact test) indicating perhaps it could be a late event (Figure 2C). The results therefore strongly support a tumour suppressor role for ARID1B in PaCa akin to a similar role of other SWI/SNF components in many cancers.


Figure 2. ARID1B CpG island (Panel A , top) methylation profile (Panel A , bottom) in PaCa cell lines harbouring low (SW1990, CFPAC1 and PANC10.05) or high (HPAFII and PANC8.13) ARID1B transcript levels and in primary human pancreatic ductal epithelial line (HPDE) also harbouring high ARID1B levels. Each horizontal row of squares (Panel A, bottom) represents result for one cell line; each square represents one CpG dinucleotide and percent Cytosine methylation is denoted by a colour code (white, $<10 \%$; green, $10-33 \%$; orange, $34-66 \%$; red, $>66 \%$ ). The total number of clones analyzed for each cell line is given at the end of each row. Methylation status following Azacytidine $(8 \mu \mathrm{M})$ treatment was evaluated only in SW1990. Panel B depicts results for senescence-associated $\beta$-galactosidase staining in permanent ARID1B or vector MiaPaCa2 clones. Panel Chows result of IHC-based detection of ARID1B in pancreatic tumour samples stratified for tumour stage.

PKU: We extended PAH mutation analysis to twenty six suspected Indian PKU families; disease causing mutations were detected in twenty four. A total of twenty different mutations were identified of which eight 'unique' India-specific mutations accounted for fourteen of twenty four mutation positive families (Figure 3A). Interestingly, only five were missense mutations while five were splice and four were nonsense mutations, respectively (Figure 3A). Two nonsense mutations were characterized to confirm significant reduction in mutant transcript levels possibly through activation of nonsense mediated decay (Figure 3B). All missense mutations affected conserved amino acid residues and sequence and structure analyses suggested significant perturbations in enzyme activity of respective mutant
proteins. This is the first report of identification of a significantly low proportion of missense PAH mutations in PKU families and together with the presence of a high proportion of splice and nonsense mutations, points to a unique $P A H$ mutation profile in Indian PKU patients.

## Future plans

1. Identification of genes/pathways that drive oncogenesis in Wnt- MSS EOSRC.
2. Characterization of $A R I D 1 B$ transcriptional targets with respect to PaCa.
3. Characterization of selected mutations affecting transcript stability/processing identified through screening of various genetic disorders.


Figure 3. Analyses of PAH mutations identified from Indian PKU patients. Panel A (top) shows the location of mutations in the PAH gene. Red, splice; blue, nonsense; green, in/del; orange, missense; black, silent and 3'UTR. India-specific mutations are denoted by '*'. A bar diagram (bottom) depicts three important PAH domains. Panel B shows the result of quantitative reverse transcription PCR carried out on RNA isolated from proband lymphoblasts (for mutation p.Y206X and p.R243X) and from a normal lymphoblast sample.

## Publications

1. Bashyam MD, Chaudhary AK and Bhat V (2012). The IVS2+837T $>G$ appears to be a relatively common 'rare' $\beta$-globin gene mutation among $\beta$-Thalassemia patients in the South Indian state of Karnataka. Hemoglobin 36: 497-503.
2. Bashyam MD, Chaudhary AK, Reddy EC, Reddy V, Acharya V, Nagarajaram HA, Devi ARR, Bashyam L, Dalal AB, Gupta N, Kabra M, Agarwal M, Phadke SR, Tainwala R, Kumar R and Hariharan SV (2012). A founder ectodysplasin A receptor (EDAR) mutation results in a high frequency of the autosomal recessive form of hypohidrotic ectodermal dysplasia in India. British Journal of Dermatology 166: 819-829.
3. Bashyam MD, Chaudhary AK, Sinha M, Nagarajaram HA, Devi ARR, Bashyam L, Reddy EC and Dalal A (2012). Molecular genetic analysis of MSUD from India reveals mutations causing altered protein truncation affecting the C-termini of E1 $\alpha$ and E1 $\beta$. Journal of Cellular Biochemistry 113:3122-3132.
4. Bashyam MD, Purushotham G, Chaudhary AK, Rao KM, Acharya V, Mohammad TA, Nagarajaram HA, Hariram V and Narasimhan C (2012). A low prevalence of $M Y H 7 / M Y B P C 3$ mutations among familial hypertrophic cardiomyopathy patients in India. Molecular and Cellular Biochemistry 360: 373-382.
5. Muranjan M, Agarwal S, Lahiri K and Bashyam M (2012). Novel biochemical abnormalities and genotype in Farber disease. Indian Pediatrics 49: 320-322.
6. *Shain AH, Giacomini CP, Matsukuma K, Karikari CA, Bashyam MD, Hidalgo M, Maitra A and Pollack JR (2012). Convergent structural alterations define SWItch/Sucrose NonFermentable (SWI/SNF) chromatin remodeler as a central tumor suppressive complex in pancreatic cancer. Proceedings of the National Academy of Sciences of the USA 109: E252-E259.
7. Kavela S, Shinde SR, Ratheesh R, Viswakalyan K, Bashyam MD, Gowrishankar S, Vamsy M, Pattnaik S, Rao S, Sastry RA, Srinivasulu M, Chen J and Maddika S (2013). PNUTS functions as a proto-oncogene by sequestering PTEN. Cancer Research 73: 205-214.
8. Raman R, Kotapalli V, Adduri R, Gowrishankar S, Bashyam L, Chaudhary A, Vamsy M, Patnaik S, Srinivasulu M, Sastry R, Rao S, Vasala A, Kalidindi N, Pollack J, Murthy S and Bashyam M. Evidence for possible noncanonical pathway(s) driven early-onset colorectal cancer in India. Molecular Carcinogenesis (In press).

* Work done elsewhere


# LABORATORY OF CANCER BIOLOGY Cellular Senescence and Sirtuin Biology, and Cancer Cervix Progression 

Faculty

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## Objectives

The major focus of our research includes:

1. Understanding the process of cellular senescence; and
2. Role of serine threonine phosphatase, calcineurin, during cervix progression.
Project 1: Understanding the mechanism of cellular senescence
Telomere attrition is a well known cause for cellular senescence. However, oxidative damage can accelerate ageing leading to premature senescence. In this context we had earlier proposed a role for wild type Ras in growth arrest (Singh et al., FASEB 2005, and Bose et al., 2011). In fact, senescence is now considered an important growth arrest mechanism in context of neoplastic transformation. We are currently focusing on two main aspects (a) Role of sirtuins in cellular senescence, and (b) effect of peroxovanadates as redox modulators in accelerating the process of senescence.
Summary of work done until beginning of this reporting year (Upto March 31, 2012)
We had earlier reported that expression of nucleolar SIRT7 in young fibroblast was very prominent, decreased during late passages and became undetectable in the senescent cells. We had also evaluated the role of SIRT1 in cervical neoplasia

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and reported its overexpression in cervical intraepithelial lesions. In addition, we reported the use of catalase resistant peroxovanadate compound, as an alternate tool to induce premature cellular senescence.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

1a. Understanding the biology of Sirtuins in context of senescence

Besides the genetic makeup of the cell, the epigenome plays a crucial role in gene regulation. The epigenome in turn is maintained by acetylation and methylation of the chromatin and its associated proteins. Chromatin modifications are brought about by DNA methyl transferases (DNMTs) and Histone deacetylases (HDACs). Amongst the various histonedeacetylases known, members of the silent information regulator 2 (Sir2) family are conserved from yeast to humans and regulate lifespan in various organisms. Some of the recent reports point to role of Sirtuins as critical regulators at the crossroads between cancer and aging. However, the exact function of the various isoforms in the context of cell proliferation and ageing is still unclear in higher organisms. In the previous report, using fibroblast cultures we had shown that SIRT7 levels decline following replicative senescence. Intriguingly, we also found the loss of nucleolar SIRT7 during replicative senescence


Figure 1. Induction of senescence in adrimaycin treated cells. (A) Senescence associated $\beta$-galactosidase (SA $\beta$-Gal) staining in U2OS cells treated with adriamycin (1 $1 \mu \mathrm{M}$ ) (B) Percentage SA $\beta$ - Gal positive cells (C) Immunoblot analysis of growth arrest markers in control and adriamycin treated U2OS cells.
in primary fibroblasts (TIG, WI38). In the present reporting year we undertook a detailed study on standardizing conditions for stress induced premature senescence by the DNA-damage agent adriamycin a widely used anticancer agent that acts by stabilizing "cleavable complexes" of DNA with topoisomerase II. Premature cellular senescence was induced in osteosarcoma cells, U2OS, by treating with low dose of adriamycin / doxorubicin.

The adriamycin treated cells showed enlarged morphology starting at $3^{\text {rd }}$ day and by $8^{\text {th }}$ day more than $90 \%$ of the cells exhibited enlarged and flattened morphology indicative of accelerated senescence. Most of the enlarged cells stained positive for senescence associated $\beta$-galactosidase activity (SA-beta Gal), as detected by 5-bromo-4-chloro-3-indolyl $\beta$-D-galactoside (X-gal) staining at pH 6.0 (Figure 1A, B). We also checked for the general growth arrest markers like p53, p21/WAF1 and p27/Kip1 in the adriamycin treated cells ( $8^{\text {th }}$
day). We found upregulation of p53 and p21/WAF1 in the senescent cells but there was no change in the expression of p27/Kip1 in the senescent cells as compared to control cells. Senescent cells also exhibited higher expression of Plasminogen Activator Inhibitor 1 or PAI-1, which is considered to be a marker for senescence (Figure 1C). The expression level of all the different isoforms of Sirtuins (SIRT1-7) was checked by quantitative Real Time-PCR in proliferating (control) and senescent U2OS cells. A significant increase in the expression of SIRT4 (3 fold) and SIRT6 in the senescent cells was noted (Figure 2). Additionally we also performed the subcellular localization of various Sirtuin isoforms. Intriguingly, SIRT2 which is mostly cytoplasmic showed a nuclear expression in senescent cells. Unlike the replicative senescence where a loss of nucleoloar SIRT7 was observed, no alterations in its localization pattern was seen in adriamycin induced premature senescence. In brief, we found that a low dose of adrimaycin


Figure 2. Expression of various Sirtuin isoforms in senescent U2OS cells normalized to control cells.
induces characteristic features of senescence in U2OS cells and this is accompanied with increased levels of SIRT4, SIRT6 and shuttling of cytoplasmic SIRT2 to nucleus.
1b. Role of Sirtuins in cervical neoplasia
Recent studies point to a close connection between cancer and ageing with Sirtuins at the crossroads. However, the role of human Sirtuin isoforms in malignancies is still controversial. We therefore designed a study to evaluate the correlation of SIRT1 expression with proliferation marker Ki-67, and growth arrest/senescence marker p27, during cervical cancer progression. The expression was evaluated by immunohistochemistry in formalin fixed archival human cervical samples: normal/ ASCUS, preneoplastic squamous intraepithelial lesions (SIL) and invasive squamous cell carcinoma (SCC). Expression of SIRT2 and SIRT7 was found to be higher in progressive grades of cancer in the following order: Normal<SIL<SCC and correlated well with the proliferative index of Ki-67. Intriguingly, SIRT1 levels were higher only in the benign stages of squamous intraepithelial lesions and correlated with the growth arrest marker p27 (Figure 3).

1c. Chemical tools which change the cellular redox states to study premature senescence

In continuation with the previous studies on Sirtuins and ageing, we are also trying to evaluate the role of peroxovanadate compounds to induce stress
induced premature senescence (SIPS). Hydrogen peroxide is the most preferred oxidative agent to study SIPS. However, a very high dose of peroxide ( $100-500 \mu \mathrm{M}$ ) is needed to induce SIPS in vitro, as the cells are abundantly equipped with catalase which effectively destroys the peroxides. The present study was designed to evaluate the role of peroxovanadium compounds in inducing SIPS as they are resistant to catalase activity. Our results indicated that diperoxovanadate (DPV) can induce features of senescence viz. flattened morphology, upregulation of p21, PAI-1 and HMGA2 in mouse fibroblasts ( NIH 3 T 3 ) at much less dose $(25 \mu \mathrm{M})$ compared to $\mathrm{H}_{2} \mathrm{O}_{2}(150 \mu \mathrm{M})$. In addition, we report altered localization of cyclin-D1 to cytoplasm in the senescent cells. However, our attempt to induce senescence in lung carcinoma cell line A549 using the similar doses of DPV was not successful. Hence, another peroxovanadium compound polyacrylic acid sodium salt peroxovanadate (PAAV), which is more catalase resistant and a stronger oxidant than DPV was tried. PAAV treatment resulted in growth arrest of A549 with features of premature senescence and SAbetagalactosidase positivity. Cytomorphological changes including cytoskeletal reorganization was a marked feature of DPV and PAAV treated cells. We surmise a role of an early activation of Rac1GTPase as a necessary event triggering the peroxovanadium mediated premature senescence.


Figure 3. Expression pattern of SIRT1 in normal, dysplastic (SIL) and squamous cell carcinoma of cervix.

Project 2: Pathways in cancer cervix progression
Cervical cancer is a leading cause of mortality among women especially in rural India. Our research group's current focus is on: cancer cervix prevention strategies in rural population and role of candidate biochemical pathways and genetic/ epigenetic changes during cancer cervix progression.

Summary of work done until beginning of this reporting year (Upto March 31, 2012)
We had earlier reported that activity of calcineurin, a serine theroine phosphatase, is higher in cervical cancer cell lines (SiHa, Hela, C33A) compared to their normal immortalized counterpart ( $\mathrm{HaCaT} \mathrm{)}$. Cyclosporine A inhibited the growth and foci formation in SiHa cells and serendipitously we made an unusual observation that cyclosporine-A treatment leads to massive cellular vacuolation.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
Immunosuppressant, cyclosporine A, results in non-apoptotic cell death in cervical cancer cells

Calcium ( $\mathrm{Ca}^{2+}$ ) has been known for long as an almost universal intracellular messenger, controlling a diverse range of cellular processes, such as gene
transcription, cell proliferation and cell death. Calcium signaling is mediated through many signal transduction cascades involving calcium interacting proteins and one such important signaling event is activation of protein phosphatase calcineurin and its downstream effector NFAT (Nuclear Factor Activated in T cells). The role of calcineurin signaling is well established in immune cells, cardiac cells and certain neuronal cells. However, there is still a lacuna in understanding of calcineurin mediated pathways with regard to other epithelial cell types. Earlier we reported a significant increase in calcineurin activity in cervical cancer cell lines. To test if calcineurin inhibition also results in growth alterations in cervical cancer cell lines, we used cyclosporine A (CsA) a well known immunosuppressant and pharmacological inhibitor of calcineurin. CsA not only inhibited the growth of cancer cervix cells but also induced massive dilation in the endoplasmic reticulum (ER). An increase in ER-Unfolded protein response (UPR) pathway was noted in CsA treated cells, which culminated in a caspase independent cell death.

## Publications

1. Ramakrishna G, Anwar T, Angara RK, Chatterjee N, Kiran S and Singh S (2012). Role of cellular senescence in hepatic wound healing and carcinogenesis. European Journal of Cell Biology 91: 739-747.

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Project I: Genome analysis and functional characterization of the genomes of microbial organisms

Characterization of the promoter and transcription factor binding sites in Mycobacterium tuberculosis

Adaptation to the various conditions encountered by the pathogen during the establishment of an infection is thought to require strict gene expression control. In prokaryotes, much of this control is at the level of transcription. There are thirteen sigma factors encoded in the genome of $M$. tuberculosis. Although some of these have been characterized, many remain to be characterized in terms of the promoter recognition specificities and their physiological roles. Furthermore, over 140 putative transcriptional regulators are presumably involved in gene expression modulation in this pathogen.

## Objectives

1. To study gene expression and regulation in mycobacteria, with special emphasis to pathogenesis, using the non-pathogenic and relatively fast growing Mycobacterium smegmatis as a model organism; and
2. To study the promoter context of mycobacterial transcription factors in order to further understand and expand their regulons.
Summary of work done until the beginning of this reporting year (upto March 31, 2012)

A Machine Learning (ML) approach was applied to
the problem of promoter prediction in $M$. tuberculosis. Different ML algorithms were evaluated and Naive Bayesian was determined to best suit the available data.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

Promoter probability densities were calculated on the basis of the Naive Bayesian model. It is believed that DNA characteristics such as GC content, Tm, DNA bending, etc., may play a role in RNA Polymerase binding beyond the canonical sequence information. Therefore, these DNA features were incorporated into the promoter prediction model. Figure 1 shows the promoter probabilities within the intergenic region of Rv0040 and Rv0041, which are situated on opposite strands. Regions of higher promoter probabilities around 43.5 kb indicate greater likelihood of RNA Polymerase binding sites, which are currently unannotated for this genomic region.

Project II: Genome analysis and functional characterization of Plasmodium falciparum

1. A machine learning approach for the classification and prediction of exons and introns in Plasmodium falciparum 3D7
Plasmodium falciparum is the causative organism of the most deadly form of malaria, which led to a high morbidity and mortality in the last few decades. It belongs to the phylum of Apicomplexa, which includes parasites of many tropical diseases. P. falciparum 3D7 has been a genome of interest for


Figure 1. Graph of promoter probability in the integenic region between Rv0040 \& Rv0041. (A) Genic organization. (B) Promoter probabilities obtained from Machine Learning. (C) GC content. Higher probability scores around 43.5 kb may represent a possible RNA Polymerase binding region, which is indicated by the red arrow.
its AT- biasness, which is $\sim 80 \%$ in the exons and $\sim 90 \%$ in the introns and intergenic regions. Since a large percentage of its genome is unannotated, the gene models generated are predictive and incomplete. Work based on the full length cDNA analysis for the genomes of apicomplexa, revealed that there exist many inconsistencies in the gene models reported for this organism and suggested a scope for improvement of the same. The present work addresses the problem of classification and prediction of exons and introns of $P$. falciparum 3D7 using the machine learning classifiers of the WEKA software (http://www.cs.waikato.ac.nz/ml/ weka), and develop an efficient method of classification and prediction for the same.

## Objective

1. To develop an improved method of classification and prediction of the intron and exon sequences in P. falciparum 3D7.
Summary of work done until the beginning of this reporting year (upto March 31, 2012)
We have shown that the numeric data transformation by correlation method has performed better as a feature for the classification and prediction of the exon and intron sequences in $P$. falciparum. We also showed that a window size of 60 , which can accommodate an average exon and intron in any gene, has worked best for the model.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

In the present study, we have considered the experimentally validated datasets of $P$. falciparum 3D7 for generating a gold standard dataset, which was divided into training ( $80 \%$ of the gold standard data) test ( $10 \%$ of the data) and validation set ( $10 \%$ of the data). We have divided the sequences in each of the dataset of different window sizes starting from 9-300 in size which were numerically transformed using their dinucleotide frequency and correlation. Models were generated on the training set applying the RandomForest (RF), machine learning classifier (MLC) of the WEKA software for the dinucleotide frequency and correlation data. The performance accuracy of the model generated was evaluated on the independent test set, which showed high precision ( 0.82 for exons and 0.84 for introns), recall values ( 0.86 for exons and 0.80 for introns) and Receiver Operating Characteristic (ROC) ( 0.88 for both exons and introns) for the window size 60 (Figure 2) for the dinucleotide correlation data. The sensitivity (86.70), specificity (80.37), accuracy (83.67) and Matthews Correlation Coefficient (MCC) (0.67) values calculated for the same, were also reasonably high. The ROC curves for each of the window sizes were plotted as the threshold curves of exons and introns for the frequency and correlation data, which showed that the window size 60, showed better ROC for the


Figure 2. Performance of the training model generated by RandomForest (RF) classifier on the test sets of different window sizes. The precision (a) recall (b) and ROC-AUC (c) values were obtained for the exon and intron sequences, which shows that the values increase with increase in the window size for the frequency data and the values increase till window size 60 and then either decrease or remain stable for the correlation data.


Figure 3. ROC curves obtained by evaluating the test set applying the model generated by Random Forest classifier for different window sizes. The ROC curves for each of the window sizes were plotted as the threshold curves of exons and introns for the frequency ( $\mathbf{a}$ and $\mathbf{b}$ ) and correlation ( $\mathbf{c}$ and $\mathbf{d}$ ) data. We can observe that the ROC curves are better for the larger window size for the frequency data whereas they are better for window size 60 for the correlation data.
dinucleotide correlation data (Figure 3). We have observed that all the values calculated increased with an increase in the window size for the frequency data, whereas the values increased and were highest for window size 60 for the correlation data. The models generated were used for the prediction of exons and introns in an independent validation set of $P$. falciparum 3D7.
2. Analytical study of Plasmodium falciparum Acyl CoA binding protein.
Acyl-CoA Binding Protein (ACBP) is a low to medium molecular weight protein (MW: 10-55 KDa) which is relatively well conserved in eukaryotic organisms. This protein functions in fatty acid biosynthesis pathway and play an important role in regulation of intracellular acyl-CoA pool size modulation, transport of acyl-CoA for betaoxidation, vesicular trafficking, complex lipid biosynthesis, and gene regulation. Though fatty acids are essential bio-macromolecules, their entry into metabolic pathways are restricted unless they are activated by thioesterification with CoenzymeA (Co-A) to form Acyl-CoA esters. Activated fatty acids undergo metabolic utilization, or storage mediated by ACBPs. Among the four isoforms of $P$. falciparum ACBP, structure of one isoform is solved by X-ray crystallography but we are yet to understand complete functional relevance of ACBPs in lower eukaryotic organisms like plasmodia. ACBP have been shown to be essential proteins for Trypanosoma brucei in its host cell dependent
blood-stream stage, it would thus be interesting to study its role in biology of $P$. falciparum.

## Objective

1. Comparative analysis of PfACBPs and Human ACBPs.

Summary of work done until the beginning of this reporting year (upto March 31, 2012)
We have done comparative analysis of Pf ACBPs and human ACBPs. We have expressed all the four ACBPs as histidine-tagged recombinant proteins.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

The maximum expression of ACBPs occurs at merozoite stage in which the lipid changes are observed. In order to know whether expression of PfACBP has anything to do with lipid changes in merozoite stage or not, we started Plasmodium falciparum culture and are in the process of further standardization.

Further, our studies show that one of PfACBP, PfACBP15 (PF3D7_1001100.1), significantly differs from other ACBPs (Figure 4). A hydropathy index plot for ACBP 15 (PF3D7_1001100.1) using Prot Scale analysis is shown in Figure 5. In some organisms, like Arabidopsis thaliana and Cryptosporidium parvum, ACBPs have been reported with additional membrane bound Ankyrin repeats.



Figure 5. Hydropathy index of ACBP-15 (PF3D7_1001100.1) using Prot Scale analysis.

## Publications

1. Ali J, Thummala SR and Ranjan A (2012). The parasite specific substitution matrices improve the annotation of apicomplexan proteins. BMC Genomics 13 (Suppl. 7): S19.
2. Muley VY and Ranjan A (2012). Effect of reference genome selection on the performance
of computational methods for genome-wide protein-protein interaction prediction. PLoS One 7: e42057.
3. Muley VY and Ranjan A (2013). Evaluation of physical and functional protein-protein interaction prediction methods for detecting biological pathways. PLoS One 8: e54325.

## LABORATORY OF TRANSCRIPTION Mechanism of Transcription Termination and Antitermination in Escherichia coli

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Staff Scientist
Senior Research Fellow
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Research Associate (Since Jun. 2012)
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Technical Officer
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## Objectives

Mechanisms of bacterial transcription termination and antitermination processes are still not very clear and offer an exciting subject for study. In our laboratory, studies in the following areas are in progress:

1. Mechanism of action of transcription termination factor, Rho;
2. Molecular basis of Rho-NusG interaction;
3. Mechanism of conversion of NusA into an antiterminator by N ;
4. Mechanism of action of transcription antitermination of Rho-dependent termination by an anti-rho factor, Psu;
5. Physiological significance of Rho dependent termination; and
6. Designing transcription modulators using synthetic biology approaches.
Summary of work done until the beginning of this reporting year (Upto March 31, 2012)
7. We established the existence of in vivo kinetic coupling between the two molecular motors, Rho and RNA polymerase using suppression of the defects of different Rho, NusG and RNAP mutants (Microbiology, 2012).
8. We proposed a multi-pronged strategy employed by the transcription antiterminator, N , to overcome the factor-dependent transcription termination (Nucl. Acids Res., 2012).
9. In collaboration with a crystallography group, we have solved the structure of the Rhoinhibitor, Psu (J. Biol. Chem, 2012).
Details of the progress made in the current reporting year (April 1, 2012- March 31, 2013)
10. The interaction surface of a bacterial transcription elongation factor required for complex formation with an antiterminator during transcription antitermination
The bacterial transcription elongation factor, NusA, functions as an antiterminator when it is bound to the antiterminator protein, N. Mode of the N-NusA interaction is unknown, knowledge of which is essential to understand the antitermination process. It was reported earlier that, outside the elongation complex (EC), N interacts with the Cterminal, AR1 domain of NusA. However, the functional significance of this interaction is obscure. We identified mutations in NusA-N-terminal domain (NTD), specifically defective for N -mediated antitermination. These are located at a convex
surface of the NusA-NTD, opposite to its concave RNA polymerase (RNAP)-binding surface. These NusA mutants disrupted the N -nut site interactions on the nascent RNA, emerging out of a stalled EC. In the N/NusA-modified EC, a Cys-53(S53C) from this convex surface of the NusA-NTD formed a specific disulfide bridge with a Cys-39 (S39C) of the NusA-binding region of the N protein. We concluded that, when bound to the EC, the N interaction surface of NusA shifts from the AR1
domain to its NTD domain. This occurred due to a massive away-movement of the adjacent AR2 domain of NusA upon binding to the EC. We propose that, the close proximity of this altered N interaction site of NusA to its RNAP-binding surface, enables $N$ to influence the NusA-RNAP interaction during transcription antitermination that facilitates the conversion of NusA into an antiterminator (Figure 1).


Figure 1. A possible model of the N-NusA NTD-EC ternary complex. Exiting RNA (red), b-flap (dark grey), NusANTD (cyan) are highlighted. Rest of the $\beta / \beta^{\prime}$ are shown in the light grey. $N$ is shown as a cartoon. $N$-and RNAPbinding residues of the NusA-NTD are shown in red and green spheres, respectively. The RNA outside the EC is shown as dotted line.
2. Structural and mechanistic basis of antitermination of Rho-dependent transcription termination by a bacteriophage capsid protein.
The conserved bacterial transcription terminator, Rho, is a potent target for bactericidal agents. Psu, a bacteriophage P4 capsid protein, is capable of inducing antitermination to the Rho-dependent transcription termination. Knowledge of structural and mechanistic basis of this antitermination is required to design peptide-inhibitor(s) of Rho derived from Psu. Using suppressor genetics, crosslinking, protein foot-printing, and FRET analyses, we describe a conserved disordered structure, encompassing 139-153 amino acids of Rho, as the
primary docking site for Psu. Also a neighbouring helical structure, comprised of 347-354 amino acids, lining its central channel, plays a supportive role in the Rho-Psu complex formation. Based on the crystal structure of Psu, its conformation in the capsid of the P4 phage, and its interacting regions on Rho, we have built an energy-minimized structural model of the Rho:Psu complex. In this model, a V-shaped dimer of Psu interacts with the two diagonally opposite subunits of a hexameric Rho, enabling Psu to form a "lid" on the central channel of the latter (Figure 2). We show that, this configuration of Psu makes the central channel of Rho inaccessible, and causes a mechanical impediment to its translocase activity (Figure 2).


Figure 2. Structural model of a Rho:Psu complex. Proposed mechanism of Psu-induced mechanical impediment to the translocase activity of Rho.

## Future plans/directions

The following projects, being pursued in our laboratory, are in different stages of completion: (i) Role of NusA in Rho-dependent termination (ii) Importance of Rho-nascent RNA interactions in vivo (iii) Mechanism of NusG mediated stimulation of Rho, and (iv) Characterization of predicted Rhobinding proteins.

## Publications

1. Banerjee R, Nath S, Ranjan A, Khamrui S, Pani B, Sen R and Sen U (2012). The first structure of polarity suppression protein, Psu from Enterobacteria phage P4, reveals a novel
fold and a knotted dimer. Journal of Biological Chemistry 287: 44667-44675.
2. Muteeb G, Dey D, Mishra S and Sen R (2012). A multipronged strategy of an anti-terminator protein to overcome Rho-dependent transcription termination. Nucleic Acids Research 40: 11213-11228.
3. Shashni R, Mishra S, Kalayani BS and Sen R (2012). Suppression of in vivo Rho-dependent transcription termination defects: evidence for kinetically controlled steps. Microbiology 158: 1468-1481.

# LABORATORY OF CELL SIGNALLING <br> Investigating the Role of Inositol Pyrophosphates in Eukaryotic Cell Physiology 

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## Objectives

Inositol pyrophosphates are derivatives of inositol that contain pyrophosphate or diphosphate moieties in addition to monophosphates. They include diphosphoinositol pentakisphosphate (PP$\mathrm{IP}_{5}$, or $\mathrm{IP}_{7}$ ) and bis-diphosphoinositol tetrakisphosphate ( $[\mathrm{PP}]_{2}-\mathrm{IP}_{4}$ or $\mathrm{IP}_{8}$ ), which participate in diverse biological functions, including DNA recombination, vesicular trafficking, apoptosis and osmotic regulation. The beta phosphate group of inositol pyrophosphates can be transferred to prephosphorylated serine residues on proteins to form pyrophosphoserine. Pyrophosphorylation occurs on several proteins within the cell, including proteins involved in ribosome biogenesis, vesicular trafficking and glycolysis. 5PP-IP ${ }_{5}\left(\mathrm{IP}_{7}\right)$ is synthesised from inositol hexakisphosphate $\left(\mathrm{IP}_{6}\right)$ and ATP by $\mathrm{IP}_{6}$ kinases. Mammals have three isoforms of IP ${ }_{6}$ kinase, IP6K1, IP6K2 and IP6K3, whereas Saccharomyces cerevisiae have a single $\mathrm{IP}_{6}$ kinase, KCS1.

Our aim is to understand the biochemical link between protein pyrophosphorylation and cellular phenomena regulated by inositol pyrophosphates. We utilise S. cerevisiae, mammalian cell lines, and knockout mouse strains as model systems to investigate the signalling and metabolic pathways that are altered when inositol pyrophosphate levels are perturbed. In particular, we focus on the following objectives:

1. Examine the role of inositol pyrophosphates in yeast physiology;
2. Understand the cellular functions of mammalian inositol hexakisphosphate kinase 1 (IP6K1);
3. Study the role of inositol pyrophosphates in whole animal physiology.

Summary of work done until the beginning of this reporting year (Upto March 31, 2012)

We observed that S. cerevisiae strains lacking KCS1 display slow growth, reduced ribosome levels, and lower rates of protein synthesis. Steady state levels of 35 S precursor rRNA, and the rate of rRNA transcription were reduced. These data suggested that inositol pyrophosphates regulate ribosome biogenesis in yeast by participating in RNA polymerase I mediated transcription of rRNA. To determine whether $\mathrm{IP}_{7}$ regulates rRNA transcription via protein pyrophosphorylation, we tested RNA polymerase I components in an in vitro pyrophosphorylation assay using radiolabelled $\mathrm{IP}_{7}$, and determined that RPA34, RPA43 and RPA190, subunits of the RNA polymerase I elongation complex, are pyrophosphorylated by $\mathrm{IP}_{7}$.
To understand the cellular functions of $\mathrm{IP}_{7}$ in mammals, we use mouse embryonic fibroblasts (MEFs) derived from IP6K1 knockout ( $/ \mathrm{p} 6 \mathrm{k} 1^{1-}$ ) embryos, which have $70 \%$ reduced levels of $\mathrm{IP}_{7}$ compared with wild type (WT) MEFs. These cells provide an excellent model to study specific cellular functions of inositol pyrophosphates that may be
biochemically linked with protein pyrophosphorylation. We monitored WT and Ip6kt MEFs for their response to hydroxyurea (HU), a DNA damage agent which causes replication stress, inducing double strand DNA breaks and triggering repair by homologous recombination (HR). HU treated Ip6k $1^{-1}$ MEFs arrested at the G1/S boundary, indicating that checkpoint activation following DNA damage is intact in the absence of IP6K1, but displayed decreased viability and reduced recovery compared with WT cells. Markers associated with DNA repair, including the RecQ family helicase BLM, were recruited to DNA damage sites, indicating that HR repair is initiated in $1 \mathrm{p} 6 \mathrm{k}^{-1}$ MEFs. However, nuclear BLM foci persisted long
after drug removal, suggesting that repair did not proceed to completion. Expression of catalytically active but not inactive IP6K1 could restore the repair process in knockout MEFs, implying that inositol pyrophosphates are required for HR mediated DNA repair.

To study the role of inositol pyrophosphates in whole animals, we have established a colony of $106 \mathrm{kl}^{+1}$ heterozygous mice and are breeding them to obtain wild type and knockout litter-mates. Our preliminary phenotypic characterisation of these mice revealed that $/ \mathrm{p} 6 \mathrm{k} 1^{-1}$ male mice are infertile. We determined that testes of $1 \mathrm{p} 6 \mathrm{k} 1^{-1}$ mice have reduced number of elongated spermatids, which


Figure 1. Yeast RNA polymerase I subunits are pyrophosphorylated by $I_{7}$. GST or GST-tagged RNA polymerase I subunits RPA34 (A), RPA43 (B) and RPA190 (C) were expressed in S. cerevisiae, immobilized on glutathione beads, incubated with radiolabelled $\mathrm{IP}_{7}$, resolved by NuPAGE, and transferred to PVDF membranes. Blots were subjected to autoradiography to monitor pyrophosphorylation, followed by immunoblotting with a tagspecific antibody. The protein sequence used in each case is indicated on the right, and deletions or point mutations introduced in each protein are underlined.
display misshapen heads and bent tails. This may explain the absence of spermiation in $1 \mathrm{p} 6 \mathrm{kl}^{-1 /}$ testes, which is reflected in the absence of mature spermatozoa in the epididymides.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
Project 1: Regulation of yeast ribosome biogenesis by $\mathrm{IP}_{7}$
Having identified three RNA polymerase I subunits as targets for $\mathrm{IP}_{7}$-mediated pyrophosphorylation, we conducted site-directed mutagenesis to map the serine residues targeted by $\mathrm{IP}_{7}$. The C -terminal lysine-rich unstructured domain of RPA34, which contains two serine residues interspersed with glutamates (S205 and S206), was mapped as the site pyrophosphorylated by $\mathrm{IP}_{7}$ (Figure 1A). Interestingly, we ruled out reported phosphorylated serines, S10, S12 and S14, as the sites for $\mathrm{IP}_{7}$ pyrophosphorylation on RPA34 (data not shown). On RPA43 we identified the C-terminal tail, which contains five serine residues, as the site for pyrophosphorylation (Figure 1B). Here again, we did not identify any of the five reported phosphorylated serines as targets for $\mathrm{IP}_{7}$. RPA190 is the largest component of RNA polymerase I, making up the active site of the enzyme. We mapped IP 7 phosphorylation sites to S1413, S1415 and S1417, which lie in the unstructured part of the Jaw domain of RPA190, close to the active site (Figure 1C).
It is possible that $\mathrm{IP}_{7}$-mediated pyrophosphorylation of one or more serine residues that we have identified is required for optimal transcription activity of RNA polymerase I. We are currently conducting assays to monitor rRNA transcription and protein synthesis in cells expressing these mutant RNA polymerase subunits that cannot be pyrophosphorylated.

Project 2: Cellular functions of mammalian inositol hexakisphosphate kinase 1 (IP6K1): Role of inositol pyrophosphates in homologous DNA recombination

Following up on our observation that MEFs lacking IP6K1 display incomplete HR mediated DNA repair, we wondered whether repair is delayed, but eventually complete. We therefore monitored DNA repair subsequent to HU removal by conducting a TUNEL assay to measure DNA damage. While the level of TUNEL staining fell to baseline 12 h after drug removal in WT MEFs, DNA damage continued to persist in $1 p 6 \mathrm{k}^{1--}$ MEFs (Figure 2A). To determine whether Ip6k $1^{-1}$ MEFs enter mitosis despite the
persistence of damaged DNA, we stained cells for histone H 3 phosphorylation at Ser10, a marker for the initiation of mitosis. An increase in the mitotic population in WT MEFs followed the timeline of DNA repair (Figure 2B and C). Ip6k1-N MEFs displayed delayed entry into mitosis 10 h after HU removal (Figure 2C), despite the persistence of DNA damage (Figure 2A). When proliferation of MEFs was monitored up to 4 days after HU removal, there was an eventual increase in viable $1 \mathrm{p} 6 \mathrm{kt}^{-1-}$ MEFs, although it was still lower than WT MEFs (Figure 2D). Proliferation of cells by continuing DNA replication without repairing damage can result in the accumulation of chromosomal lesions. Defects in HR can lead to increased sensitivity to mitomycin C, a drug that induces DNA interstrand crosslinks, blocking DNA replication. Analysis of metaphase spreads following mitomycin C treatment may therefore be used to probe defects in DNA repair. When treated with mitomycin C, Ip6k1- MEFs show more chromosomal abnormalities such as triradial and quadriradial chromosomes, and chromatid breaks, compared to WT MEFs (Figure 2E and F).

In summary, our data reveal a role for inositol pyrophosphates synthesised by IP6K1 in HRmediated repair of DNA double strand breaks in mammalian cells. A research paper describing this work has recently been published (Jadav et al., Journal of Biological Chemistry, 2013). At a mechanistic level, inositol pyrophosphates may act by binding or pyrophosphorylating one or more proteins involved in HR. In the near future we will focus on identifying molecular targets that mediate the involvement of IP6K1 in HR. Our observation that $I p 6 k 1^{-1}$ MEFs accumulate chromosomal aberrations raises the possibility that these cells possess a higher tumourigenic potential compared with their wild type counterparts. We will therefore conduct studies to determine the effects of altered IP6K1 activity on tumourigenesis and cancer chemotherapy.

Project 3: Physiological role of $\mathrm{IP}_{7}$ in mice: Regulation of platelet function by IP6K1

This is a new activity. This project explores the link between inositol pyrophosphates and inorganic polyphosphate (polyP), a linear polymer of orthophosphate moieties linked by phosphoanhydride bonds. PolyP of chain length 60-100 phosphate units is present in dense granules of mammalian platelets, and regulates the blood clotting cascade at multiple stages. Budding yeast lacking the $\mathrm{IP}_{6}$ kinase KCS1 display
substantially lowered levels of polyP, prompting us examine whether $\operatorname{lp} 6 k 1^{-1}$ mice, which have 70\% reduced levels of $\mathrm{IP}_{7}$, have any defects in polyP accumulation and platelet function.
Western blot analysis confirmed that extracts from WT mouse platelets contain IP6K1, but there is no detectable band in $/ p 6 k 1^{-1 /}$ platelets (Figure 3A). Earlier studies have shown that the DNA binding
fluorophore DAPI stains polyP present in human platelets. DAPI staining of platelets isolated from WT mice revealed polyP accumulation, whereas very low polyP levels were observed in isolated lp $6 \mathrm{kt}^{-1}$ platelets (Figure 3B, C). The fluorescent lipophilic dye DiOC6 which stains platelet membranes was used to detect isolated platelets, and showed no difference in staining intensity between WT and Ip6k1-/ platelets.


Figure 2. Persistence of DNA damage in Ip6ktres. (A) Detection of DNA double strand breaks (DSBs) by TUNEL staining and flow cytometry analysis after HU ( $12 \mathrm{~h}, 0.5 \mathrm{mM}$ ) treatment of MEFs and recovery for the indicated time; bars represent mean $\pm$ range of two independent experiments. (B) Representative immunofluorescence images of histone H3 Ser10 phosphorylation in MEFs after HU ( $12 \mathrm{~h}, 0.5 \mathrm{mM}$ ) treatment and recovery for the indicated time. (C) Quantitation of (B); bars indicate the percentage of histone H3 Ser10 positive cells ( $n=140$; representative of two experiments). (D) Cell viability measurement by MTT assay following treatment with HU (12 $h, 0.5 \mathrm{mM}$ ), and recovery for the indicated time; bars represent mean $\pm$ range of two independent experiments. (E) Representative images showing chromosomal lesions (marked by arrows) found in metaphase spreads from MEFs treated with mitomycin C (12 h, 300 nM ). (F) Quantitation of (G); data (mean $\pm$ s.e.m. $\mathrm{n}=41$ ) are representative of two experiments. P values are from a two-tailed Student's $t$-test (**pd $\leq 0.01$ ).

Analysis of haematologic parameters in WT and $106 \mathrm{k}^{-1 /}$ mice revealed no difference in platelet count, platelet size or other blood parameters between these groups (data not shown). To monitor platelet activation by thrombin we measured the
levels of surface P -selectin, a cell adhesion molecule released from platelet $\alpha$-granules (Figure 3D). No alteration in P-selectin surface expression in $I p 6 \mathrm{k}^{1-}$ platelets implies that IP6K1 does not influence platelet $\alpha$-granule content or its thrombin


Figure 3. Altered platelet function in Ip6kTres (A) Western blot analysis of lysates prepared from WT and $/ p 6 \mathrm{kl}^{-1}$ platelets pooled from 3 mice of each genotype, using an antibody against IP6K1. GAPDH was used as a loading control. The blot is representative of 3 independent experiments. (B) Representative confocal fluorescence micrographs of platelets isolated from WT and Ip6k 1-1 mice, stained with DAPI (red) to detect polyP and DiOC6 (green) to visualize platelets. (C) Quantification of images in (B), using ImageJ software. Data are mean $\pm$ s.e.m. ( $\mathrm{n}=8$ mice of each genotype, with 20 platelets analyzed per mouse). (D) Surface P -selectin expression in resting and thrombin stimulated WT and $/ \mathrm{p} 6 \mathrm{k}^{1 /}$ platelets analysed by flow cytometry. Data are median fluorescence intensity (MFI), mean $\pm$ s.e.m. $\left(\mathrm{n}=3\right.$ ). ( E ) Thrombin stimulated aggregation of washed platelets from WT and $/ \mathrm{p} 6 \mathrm{k} 1^{-1}$ mice measured spectrophotometrically as a decrease in percent light transmission over a period of 10 min . Samples were pooled from 3 mice of each genotype for the analysis. Data are mean $\pm$ s.e.m. from 3 independent experiments. (F) Change in turbidity as a function of time was monitored spectrophotometrically at 405 nm in recalcified platelet releasates (PR) mixed with platelet poor plasma (PPP). Clotting time, the time taken to reach maximum absorbance, was measured in WT and $l p 6 \mathrm{k}^{-1}$ samples of PPP+PR or PPP alone. Data are mean $\pm$ s.e.m. $(\mathrm{n}=8)$. $p$ values are from a two-tailed Student's t test ( ${ }^{*} p \mathrm{~d} \leq 0.05$; ** $p \mathrm{~d} 0.01$; n .s., not significant, $p>0.05$ ).
stimulated release. Following activation by different agonists, platelets adhere and aggregate to form a plug at the site of injury, leading to primary haemostasis. Platelet aggregation upon thrombin stimulation was measured in washed platelets isolated from WT or $1 p 6 k^{-1}$ mice. We observe a significant decrease in the extent of aggregation of lp6k1-1 platelets (Figure 3E). This could be attributed to compromised von Willebrand factor function, which is regulated by polyP present in $\alpha$ granules. To examine the effect of polyP reduction on plasma clotting time, we added platelet releasates from WT and Ip6k1-1 mice to their autologous citrated platelet poor plasma (PPP), prior to recalcification and clot turbidity measurement. Total clotting time, the time taken to reach maximum turbidity following recalcification, is significantly lengthened in Ip $6 k 1^{-1 /}$ samples (Figure 3F). On the other hand, clotting time of recalcified PPP alone was unaltered in $1 \mathrm{p} 6 \mathrm{k} 1^{-1}$ compared with WT. This clarifies that changes in
platelet derived factor(s), and not plasma components, are responsible for the prolonged clotting time observed in $/ p 6 \mathrm{kl}^{-1 /}$ samples.

Our data so far reveal that the metabolic link between inositol pyrophosphates and polyP is conserved between yeast and mammals. Reduction in platelet polyP in Ip6k1-- mice leads to compromised platelet aggregation and lengthened clotting time. We are currently examining whether these changes lead to haemostasis defects in Ip6k1-1 mice. Our finding that IP6K1 is a novel player in platelet aggregation and blood clotting assumes clinical significance in the context of thrombotic and bleeding disorders.

## Publications

1. Jadav RS, Chanduri MVL, Sengupta S and Bhandari R (2013). Inositol pyrophosphate synthesis by inositol hexakisphosphate kinase 1 is required for homologous recombination repair. Journal of Biological Chemistry 288: 3312-3321.

# LABORATORY OF PLANT MICROBE INTERACTIONS <br> Understanding Virulence Mechanisms of Xanthomonas Plant Pathogens and Interaction with Host Plants 

Faculty

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Subhadeep Chatterjee

## Objectives

1. Identification and characterization of virulence factors of Xanthomonas;
2. Role of cell-cell communication in Xanthomonas colonization and virulence;
3. Function of protein secretion system in Xanthomonas and its role in virulence; and
4. Role of PAMP in pathogen recognition and plant defense response.
Summary of work done until beginning of this reporting year (Upto March 31, 2012)
We are trying to understand the virulence mechanisms of important Xanthomonas pathogens like, Xanthomonas campestris pv. campestris (Xcc; a pathogen of crucifers), Xanthomonas oryzae pv. oryzae, Xanthomonas oryzae pv. oryzicola (Xoo, Xola; pathogens of rice) and Xanthomonas axonopodis pv. citri (Xac; pathogen of citrus). In Xanthomonas oryzae pv. oryzae (Xoo), a pathogen of rice, we have previously identified several virulence associated functions which are regulated by Diffusible Signal Factor (DSF). Xoo exhibits atypical regulation of virulence associated functions, in contrast to closely related Xanthomonas. We have also proposed a model for the role of Xoo DSF in coordinating the switch between the planktonic and biofilm lifestyle of this bacterium. We are presently studying the role of chemotaxis, motility and components of cell-cell signaling in virulence of Xoo. We have also initiated studies to understand the role of DSF in regulating virulence associated function in Xanthomonas oryzae pv. oryzicola (Xcola) which causes a serious disease of rice known as Bacterial Leaf Streak (BLS).

Staff Scientist
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Senior Research Fellow
Senior Research Fellow
Junior Research Fellow
Junior Research Fellow (Since Jul. 2012)
Technical Officer
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Previously we have isolated a novel adhesin of Xoo, XadM, which is required for virulence. In this study we have further characterize the role of XadM in entry, attachment and colonization of Xoo.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
Project 1: Role of DSF in virulence of Xanthomonas oryzae pv. oryzicola (Xcola)
Since DSF exhibits atypical regulation of virulence associated functions in closely related Xanthomonas, we wanted to understand the role of DSF in Xanthomonas oryzae pv. oryzicola (Xcola). Xcola has an atypical lifestyle as compare to Xoo (a xylem dwelling pathogen), as it infects the rice plant by gaining entry through stomata and grows in the parenchyma tissue. Xcola exhibits streak like symptoms as opposed to long lesions exhibited by Xoo on the mid vein of rice leaves. We have made deletion mutants of rpfF and components of DSF mediated cell-cell signalling in Xcola. Phenotypic characterization of $\Delta r p f F$ mutant of Xcola indicated that DSF is required for virulence and in planta growth of Xoo (Figure 1). To understand why DSF mutants of Xcola exhibits in planta growth deficiency, we have performed microarray analysis of transcriptional changes between wild type, DSF deficient mutants with or without exogenous supplementation of DSF of Xcola along with phenotypic characterization of DSF regulated traits. Transcriptional studies indicated that several genes involved in iron metabolism are altered in the DSF deficient $\Delta r p f F$ mutant. Growth assay under low iron conditions, biochemical analysis of iron content, iron uptake assay and in planta iron supplementation
experiments indicated that the in planta growth deficiency of the "rpfF mutant of Xcola is due to deficiency in ferric $\left(\mathrm{Fe}^{+3}\right)$ uptake. Along with iron, we have also identified several other virulence associated functions that are regulated by DSF such as Type III secretion system and its effectors,
attach and form biofilms. Furthermore, we show that XadM is exposed on the cell surface and its expression is regulated by growth conditions and plays an important role in the early attachment and entry inside rice leaves (Figure 2). We have also proposed a model for the role of XadM in the


Figure 1. DSF deficient mutant of Xanthomonas oryzae pv. oryzicola exhibits virulence deficiency on rice. Rice leaves were inoculated with the YT6 (wild type), "rpfF (DSF) and "rpfF/pSC9 (rpfF mutant harboring the complementing plasmid). Typical symptoms of Bacterial Leaf Streak (BLS) are seen after 14 days post inoculation. (B) DSF is required for growth under low iron conditions. Different strains of Xcola were grown in rich media (Peptone sucrose; PS), PS + 2,2'-dipyridyl (Dp; iron chelator) with or without exogenous iron ( $\mathrm{FeSO}_{4}$ ) supplementation.
components involved in biofilm formation and motility. In future, we want to do detail study to understand the role of DSF in regulating these virulence associated function.
Project 2: Understanding the mechanism of attachment and biofilm formation in Xanthomonas
By screening a transposon induced mutant library of Xanthomonas oryzae pv. oryzae, the bacterial blight pathogen of rice, we have identified a novel 5.241 kb open reading frame (ORF) named xadM that is required for optimum virulence and colonization. This ORF encodes a protein, XadM, of 1746 amino-acids that exhibits significant similarity to Rhs family proteins. The XadM protein contains several repeat domains similar to a WallAssociated Surface Protein (WASP) of Bacillus subtilis, which has been proposed to be involved in carbohydrate binding. The role of XadM in $X$. oryzae pv. oryzae adhesion was demonstrated by the impaired ability of a xadM mutant strain to
cell-cell and cell-host attachment. In our proposed model, XadM plays an important role in the first step of colonization, the early attachment of Xoo at the hydathodal entry points on the rice leaf. XadM then promotes attachment of Xoo cells with the host cell-wall and with secreted extracellular polysaccharide, promoting stable biofilm formation (Figure 3). Interestingly, XadM (an Rhs family protein) homologs are present in several diverse bacteria including many Xanthomonas and animal pathogenic bacteria belonging to Burkholderia spp. The rhs genes are a family of enigmatic genes present in diverse bacteria and have been implicated to play a role of a rearrangement hot spot. Despite their ubiquity, rhs genes have not been assigned a definitive function. This is the first report of a role for XadM, an Rhs family protein, in adhesion and virulence of any pathogenic bacteria. We are doing further molecular genetics and gain of function studies to understand the contribution of rhs family of adhesins in Xanthomonas virulence.


Figure 2. XadM is required for early attachment and entry of $X$. oryzae pv. oryzae into rice leaves. (A) Inoculation of rice leaves with enhanced green fluorescent protein-tagged $X$. oryzae pv. oryzae strains were done and confocal microscopy based assay were conducted 1 h after inoculation. The panels depict confocal microscope based projection images ( 200 by 200 by $60 \mu \mathrm{~m}^{3}$ in the $\mathrm{X}, \mathrm{Y}$ and Z axis beginning from the dorsal surface) of rice leaves inoculated with wild type (BXO43), XadM1 (xadM1 mutant) and XadM1(pSC2). Scale bar: $20 \mu \mathrm{~m}$. (B) Reduction of leaf attachment and entry is associated with xadM mutation of $X$. oryzae pv. oryzae. The number of enhanced green fluorescent protein-expressing cells of different strains of $X$. oryzae pv. oryzae on the surface as well as inside rice leaves after 1 h of infection as determined using confocal microscopy. Data were collected up to approximately $600 \mu \mathrm{~m}$ in length from the tip of the leaf and a distance of approximately $80 \mu \mathrm{~m}$ in depth from the dorsal surface of each leaf. Each bar shows mean and standard deviation of values obtained from three leaves. The values obtained for the XadM1 mutant were found to be significantly different ( $p<0.05$ ) compared with either the wild type strain or the XadMI mutant harboring the complementing plasmid, XadM1(pSC1), in a student's two tailed $t$ test for independent means. Similar results were obtained in independent experiments.


Figure 3. A proposed model for the role of XadM in attachment and biofilm formation of $X$. oryzae pv. oryzae. XadM along with XadA is required for the initial attachment of Xoo on hydathodal entry points present on the leaf surface. Inside the hydathode, XadM is required for the tight attachment with the plant cell-wall and also contributes to the attachment of Xoo cells with secreted extracellular polysaccharide. XadA promotes cell-cell attachment and XadM is involved in attachment of Xoo on host cell-wall and extracellular polysaccharide, which promotes stable biofilm formation inside rice leaves.

## Publications

1. Almeida RPP, Killiny N, Newman KL, Chatterjee S, Ionescu M and Lindow SE (2012). Contribution of rpfB to cell-to-cell signal synthesis, virulence, and vector transmission of Xylella fastidiosa. Molecular Plant-Microbe Interactions 25: 453-462.
2. Pradhan BB, Ranjan M and Chatterjee S (2012). XadM, a novel adhesin of Xanthomonas oryzae pv. oryzae, exhibits similarity to Rhs family proteins and is required for optimum attachment, biofilm formation and virulence. Molecular Plant-Microbe Interactions 25 : 1157-1170.
3. Rai R, Ranjan M, Pradhan BB and Chatterjee S (2012). Atypical regulation of virulence-
associated functions by a diffusible signal factor in Xanthomonas oryzae pv. oryzae. Molecular Plant-Microbe Interactions 25: 789-801.
4. Beaulieu ED, Ionescu M, Chatterjee S, Yokota K, Trauner D and Lindow S (2013). Characterization of a Diffusible Signaling Factor (DSF) from Xylella fastidiosa. mBio 4: e00539-12.
5. Pandey A and Chaterjee S (2013). Signaling in plant-microbe interactions. Plant Stress 7: 52-59.

## Other Publications

1. Chatterjee $S$ (2013). Review of: Annual Review of Microbiology, 2011. Susan Gottesman and Caroline S Harwood (eds). Current Science 104: 653-654.

# LABORATORY OF CELL DEATH \& CELL SURVIVAL Molecular Mechanisms Controlling Cell Life and Death 

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## Objectives

1. To dissect the functional network of phosphatases regulating cell life and death;
2. To identify and characterize novel protein complexes in maintaining genomic stability; and
3. To understand the cellular functions of canonical and non-canonical ubiquitination.

Summary of work done until the beginning of this reporting year (Upto March 31, 2012)

Phosphatases are a group of ubiquitously expressing enzymes, which are responsible for the removal of a phosphate group of various substrates in a cell. Several studies have suggested an intricate involvement of phosphatases in controlling cellular life and death, but systematic studies to dissect the complex network of phosphatases and functional role of their interactions in these processes are lacking. We initiated our studies on functional phosphatase network with the identification of interacting partners of every phosphatase in the cell. In this work we have previously identified and characterized WWP2 as a novel functional E3 ubiquitin ligase for PTEN (Maddika et al., Nature Cell Biol. 2011). In addition to WWP2, we identified the protein phosphatase1 nuclear targeting subunit PNUTS (PPP1R10) as a potential PTEN-associated protein. We have shown that PNUTS directly interacted with the lipidbinding domain (C2 domain) of PTEN and sequestered it in the nucleus. Depletion of PNUTS
leads to increased apoptosis and reduced cellular proliferation in a PTEN-dependent manner. PNUTS expression was elevated in certain cancers compared with matched normal tissues. Overall, our studies revealed PNUTS as a novel PTEN regulator and a likely oncogene. (Kavela et al., Cancer Res. 2013).
Details of progress in the current reporting year (April 1, 2012 - March 31, 2013)
Project 1: Functional studies on phosphatase networks
We are continuing our studies on identification of new components in the cellular phosphatase networks. PHLPP1 (PH domain leucine rich repeat protein phosphatase 1 ) is a Serine/Thronine protein phosphatase and has recently been characterized as a potential tumor suppressor. The loss of PHLPP1 is reported in various cancers such as colon, lung, breast, ovarian and prostate cancers. Functionally, PHLPP1 is shown to directly dephosphorylate Akt and antagonizes the cellular phosphotidylinosital-3 kinase (PI3K)/Akt signalling pathway thus triggers apoptosis and suppresses tumor growth. By utilizing tandem affinity purification approach we have identified WDR48 and USP12 as novel PHLPP1 associated proteins (Figure 1A). WDR48-USP12 complex deubiquitinates PHLPP1 (Figure 1B) and thereby enhances its protein stability. Similar to PHLPP1 function, WDR48 and USP12 negatively regulate Akt activation and thus promote cellular apoptosis.

Functionally, we show that WDR48 and USP12 suppress proliferation and migration of tumor cells (Figure 1C \& 1D). Collectively, our results reveal WDR48 and USP12 as novel PHLPP1 regulators
and potential suppressors of tumor cell survival. Our further studies are focused on mapping the functional networks of other phosphatases in cells such as lipid phosphatases, dual specificity


Figure 1. WDR48-USP12 complex controls cell proliferation by regulating PHLPP1. (A) 293T cells were transfected with the indicated Flag tagged plasmids and the interaction of endogenous PHLPP with WDR48 and USP12 was evaluated by immunoprecipitation with Flag antibody followed by immunoblotting with PHLPP1 antibody. (B) HCT116 cells were transfected with either control shRNA or shRNA against WDR48 and USP12. Cell lysates prepared after 6 hour MG132 $(10 \mu \mathrm{M})$ treatment were subjected to immunoprecipitation and the ubiquitinated PHLPP1 was detected with anti-ubiquitin antibody. (C) HCT116 cells transfected with different shRNAs were seeded and their proliferation was measured by Tryphan blue exclusion for 5 days. (D) HCT116 cells expressing control shRNA or WDR48 shRNA were allowed to grow to confluence, and their ability to migrate was analyzed by using scratch assay.
phosphatases, non-receptor protein tyrosine phosphatases and receptor protein tyrosine phosphatases.

## Project 2: Roles of canonical and noncanonical ubiquitination in cells

Ubiquitination is an ATP-dependent, highly ordered multistep enzymatic process which results in covalent attachment of ubiquitin to the substrate. Ubiquitin linked to the substrates serves as a molecular tag that marks proteins for either
degradation by proteasome dependent pathway or to function in wide variety of processes in a proteasome independent manner. In this project we are interested in studying both the canonical and non-canonical functions of ubiquitination in cells. During our studies on canonical ubiquitination, we are interested in identifying proteosomal substrates of various E3 ligases. WWP2 is an E3 ubiquitin ligase that belongs to NEDD-4 like family HECTtype E3 ligases. It plays an important role in different cellular functions such as transcription, embryonic
stem cell fate, cellular transport, T-cell activation and Apoptosis. In this study we identified p73 as a novel substrate of WWP2 that might be functionally important in WWP2 pro-oncogenic function.
p73 is a p53 related transcription factor that exists in full length and N -terminal truncated $\Delta \mathrm{Np} 73$ isoforms. Due to their opposing functions in controlling cell survival it is critical to maintain the balance between these two proteins but the precise mechanism that regulate their levels are not clear. In our study, we identified WWP2, an E3 ligase as
this project are further focused on characterizing the non-canonical functions of ubiquitination.

## Publications

1. Jain MV, Paczulla AM, Klonisch T, Dimgba FN, Rao SB, Roberg K, Schweizer F, Lengerke C, Davoodpour P, Palicharla VR, Maddika S and Los M (2013). Interconnections between apoptotic, autophagic and necrotic pathways: implications for cancer therapy development. Journal of Cellular and Molecular Medicine 17: 12-29.


Figure 2. A proposed model to show the role of WWP2 and WWP2-WWP1 heterodimeric complex in regulating p73 and $\Delta \mathrm{Np} 73$ levels in normal and stress conditions.
a novel p73 associated protein that ubiquitinates and degrades p73. In contrast, WWP2 also promotes degradation of $\Delta \mathrm{Np} 73$ but independent of its catalytic function. We showed that WWP2 heterodimerizes with another HECT E3 ligase WWP1, which specifically ubiquitinates and degrades $\Delta N p 73$. During cellular stress WWP2 is inactivated that leads to upregulation of p73 whereas WWP2-WWP1 complex is intact to degrade $\Delta N p 73$ thus playing an important role in shifting the balance between p 73 and $\Delta \mathrm{Np} 73$. Collectively, our results reveal a new functional E3 ligase complex that differentially regulates cellular p73 and $\delta \mathrm{Np} 73$ (Figure 2). Our future studies in
2. Kavela S, Shinde SR, Ratheesh R, Viswakalyan K, Bashyam MD, Gowrishankar S, Vamsy M, Pattnaik S, Rao S, Sastry RA, Srinivasulu M, Chen J and Maddika S (2013). PNUTS functions as a proto-oncogene by sequestering PTEN. Cancer Research 73: 205214.
3. Dulla B, Kirla KT, Rathore V, Deora GS, Kavela S, Maddika S, Chatti K, Reiser O, Iqbal J and Pal M. Synthesis and evaluation of 3-amino/ guanidine substituted phenyl oxazoles as a novel class of LSD1 inhibitors with antiproliferative properties. Organic \& Biomolecular Chemistry (In press).

# LABORATORY OF DROSOPHILA NEURAL DEVELOPMENT <br> Understanding Patterning and Development of Central Nervous System using Drosophila melanogaster 

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## Objective

The key objective of the laboratory is to understand how neural progenitor cells attain their positional identity in developing Central Nervous System (CNS) of an organism and how does this translate into generation of a variety of cell types found in CNS (as represented in the Figure 1). Hox family of transcription factors are known to play an important role in giving the positional identity to the cells and generation of a variety of cell types along the AP axis of the CNS during development. The molecular basis of this phenomenon is not well investigated. We are interested in understanding the molecular basis of Hox function in patterning CNS using Drosophila melanogaster as our model organism, focusing mainly on early embryonic and late larval stages of development. To this end, the specific aims of our laboratory are as follows:

1. Understanding the molecular function of Hox gene Abdominal-A (Abd-A) in larval CNS patterning
Abdominal region of the Drosophila larval CNS has lesser number of neurons compared to its thoracic counterpart. Hox gene $A b d-A$ is known to cause programmed cell death (apoptosis) of neural progenitor cells (also called Neuroblasts-Nbs) and therefore limit the number of neurons in abdominal region of CNS. The precise molecular details of how Abd-A causes Nb apoptosis are unknown. Genetic evidence suggests a role for a helix-loophelix transcription factor Grainyhead (Grh) along with Abd-A in control of this apoptosis. Characterization of the molecular basis of this link is the primary goal of this project. Moreover, since Grh is involved in Nb apoptosis and is not expressed
in neuronal progeny refractory to this apoptosis, it is of interest to define grh regulation in these cells which keeps grh "on" in the Nbs and "off" in the neuronal progeny of Nbs .
2. Understanding the role of Hox gene Deformed (Dfd) in patterning of embryonic subesophageal ganglia
Hox genes express in CNS (in neural progenitor cells) in embryonic stages of development (as represented in Figure 1) but how does their expression patterns the embryonic nervous system is not well understood. Deformed (Dfd) is known to express in the cells of subesophageal ganglion of embryonic CNS, and this project focuses on understanding auto-regulation of Dfd in this region and to find out how this helps in giving cells their specific positional identity. This is being done by using a 630bp long auto-regulatory CNS specific enhancer for Dfd which recapitulates the expression of Dfd gene in developing embryonic CNS.
3. Investigating the role of Abdominal-B (AbdB) and Double-sex (Dsx) in terminal CNS patterning.
The set of Nbs in the terminal region of CNS show sex specific proliferation and survival. Although the role of the sex determining hierarchy and Hox gene Abd-B, in growth and differentiation of Drosophila genital discs, is well worked out, little is known about how sex determination hierarchy and Abd-B intersects with cell proliferation and survival behavior of terminal Nbs (tNbs) in the larval VNC. Double-sex (Dsx) is the most downstream transcription factor of the sex-specification hierarchy. I intend to test the interaction between Abd-B and Dsx in gender specific proliferation of these cells.


Figure 1. Early embryonic CNS comprises of an equivalent population of neural progenitor cells (shown in red circles) which start to express specific Hox genes and therefore acquire specific positional identity (represented by different colored circles). These cells generate a variety of different cell types in both embryonic and larval CNS. In larval stages thoracic and abdominal Nbs differ in their number and proliferation profile as shown. Thoracic Nbs stop proliferation by cell cycle exit, while abdominal Nbs (in both sexes) and terminal Nbs (tNb in females) die as a result of apoptosis, the tNbs in males continue dividing and give rise to more neurons as shown.

Summary of work done until the beginning of this reporting year (Upto March 31, 2012)

1. Understanding the molecular function of Hox gene Abd-A in larval CNS patterning

It is known that grim gene plays primary role in this apoptosis and relevant enhancer for the grim gene in Nbs lies in 23kb genomic region referred to as NBRR-Neuroblast Regulatory Region. The region has been divided into 4 overlapping genomic fragments (of $8 \mathrm{~kb}, 10 \mathrm{~kb}, 8 \mathrm{~kb}$ and 8 kb each) that are to be cloned into pCasPer-lacZ reporter construct to generate enhancer-lacZ transgenic lines for screening. These 4 fragments have been amplified by PCR from genomic DNA using region specific primers. The first 8 kb fragment has been already cloned into pCasPer-nls-lacZshuttle vector and microinjections are being done for the same. To study the Nbs specific regulation of grh, a 4 kb enhancer of grh responsible for its expression in Nb was divided into three fragments to identify the minimal enhancer which will recapitulate its expression in Nbs. The three genomic fragments have been cloned into shuttle vectors and made into transgenic lines.
2. Role of Hox gene Deformed (Dfd) in patterning of embryonic subesophageal ganglia
The CNS specific 630bp autoregulatory enhancer of Dfd was cloned into shuttle vector and transgenic lines were being made to check its expression in specific cell types (especially Nbs) in subesophageal region of the embryonic brain. Simultaneously, a protein expression was
standardized for Dfd, Exd and Hth. The 630bp Dfd autoregulatory element was scanned for Hox-Exd binding sites and two putative compound binding sites were identified for these two transcription factors. In vitro binding studies were done on these binding sites using EMSA and both of the binding sites showed binding to Dfd-Exd hetrodimer. In order to investigate the in vivo relevance of these binding sites, these sites were mutagenized in 630bp DNA element and these various mutagenized forms of the enhancers have been subcloned into the $p$ CasPer-n/s-lacZ shuttle vector and the transgenic lines are being made for the same. These transgenic lines will be tested for their capacity to activate the reporter $\beta$-galactosidase to test the relevance of the binding site and its direct role in autoregulation of Dfd gene.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

1. Understanding the molecular basis of Hox gene Abdominal-A (Abd-A) in larval CNS patterning
A systematic screening of the 23 kb NBRR is ongoing to identify Nb specific grim enhancer responsible for grim activation and Nb apoptosis. The 23kb region has been divided into 4 overlapping genomic fragments (of $8 \mathrm{~kb}, 10 \mathrm{~kb}, 8 \mathrm{~kb}$ and 8 kb each) which are being screened for their ability to drive Nb specific expression of lacZ reporter in late third instar larval (LL3) brain. These 4 fragments have been amplified by PCR using region specific primers from genomic DNA and all the four fragments have been cloned into pCasPer-lacZ shuttle vectors to make transgenic lines, and
transgenic line for two of the fragments have already been made and one of them has been analyzed which has helped us to narrow down the search for the relevant grim enhancer to an 8 kb region. The 4kb enhancer of grh has been fragmented into three parts and the relevant enhancer for the expression of grh has been further narrowed to a 1.5 kb region.
2. Role of Hox gene Deformed in patterning of embryonic subesophageal ganglia
The 630bp Dfd autoregulatory element was scanned for Hox-Exd binding sites and two putative compound binding sites were identified for these two transcription factors. In vitro binding studies were done on these binding sites using EMSA and both of the binding sites showed binding to DfdExd hetrodimer. In order to investigate the in vivo relevance of these binding sites, these sites were mutagenized in 630bp DNA element and the
mutagenized forms of the enhancers have been subcloned into the $p$ CasPer-n/s-lacZ shuttle vector and the transgenic lines have been made and are being analyzed for the same.
3. Investigating the role of Abdominal-B (Abd-B) and Double-sex (Dsx) in terminal CNS patterning
The standardizations for co-staining with BrDU and other antibodies for the larval nervous system has been done, this is important to monitor the dividing cells (Nbs and ganglion mother cells) in LL3 CNS. The co-staining procedure for BrDU and other epitopes will be used to monitor the tNbs division to test the role of Abd-B in this proliferation by making Abd-B null clones in larval CNS. This will be attempted in two genetic backgrounds, first wherein the dividing Nbs will be randomly marked by GFP using MARCM technique and in second case where GFP will be specifically driven in tNbs .

# LABORATORY OF CELL CYCLE REGULATION Elucidating the Role of Effector Proteins in G1 to S Phase Progression 

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## Objectives

1. Identification of new effector proteins involved in regulation of E2F-responsive promoters; and
2. Study of chromatin modifying proteins in cell cycle regulation.

Project 1: Identification of new effector proteins involved in regulation of E2Fresponsive promoters
One of the major roles of E2F proteins is to regulate the transition from G1 to S phase. However, how E2Fs affect passage into $S$ phase is still poorly understood. In this project we aim to identify new effector proteins involved in regulation of E2Fresponsive promoters and better understand how these effectors influence transcriptional regulation during G 1 to S phase progression.
Summary of work done until beginning of this reporting year (Upto March 31, 2012)

We decided to take a candidate based approach to look for effector proteins. In order to harvest cells in a particular cell cycle phase, we synchronized HeLa cells using double thymidine block and determined the time points that we could use to harvest cells in G1/S, S, G2/M, early G1 and late G1 phase. We also standardized Chromatin immunoprecipitation (ChIP) for immunoprecipitation against E2F1 and E2F4 antibodies followed by qPCR.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
We are aiming to identify the effector proteins which modulate the expression of E2F responsive promoters. E2F family of proteins has both
activating and repressive members. The repressive members include E2F3b, E2F4 and E2F5. We started our study with E2F4, wherein we aimed to probe for its interacting partners. E2F4 being a nuclear protein is present in very low amounts in cell. To circumvent this problem we cloned E2F4 into a bacterial expression vector $\mathrm{pGex4t1}$ which leads to expression of a fusion protein GST-E2F4. Expression and purification of GST-E2F4 was standardized and eventually we could produce GST-E2F4 to homogeneity. GST-E2F4 bound to the glutathione agarose beads will be used in future for pulldown assays to probe for the interacting partners.
One major limitation of working with endogenous proteins is the low abundance of such proteins in the cell. Since we expect to use endogenous proteins in most our experiments, their scarce availability can prove to be a limiting factor for our future experiments. Therefore, we cultured HeLa Spinner cells which can be used for producing large amounts of cells in suspension cultures. HeLa cells were grown in Joklik's media and growth conditions were standardized.

Project 2: Study of chromatin modifying proteins in cell cycle regulation
Histone 3 lysine 4 trimethylation is linked to active gene expression, but its precise role in cell cycle regulation is now being uncovered. We have shown that MLL-family of H3K4 histone methyltransferases is linked to cell cycle regulation by interacting with E2F1. In this project, we are looking at other roles of this chromatin-modifying complex that may influence cell cycle regulation.


Figure 1. HeLa spinner cultures and GST-E2F4 expression. (A) HeLa spinner cells were grown in Joklik's media in a non $\mathrm{CO}_{2}$ incubator. Up to $6 \times 10^{5}$ cells $/ \mathrm{ml}$ could be obtained. (B) GST-E2F4 was expressed and purified using GST beads. CBB, Coomassie Brilliant Blue staining; WB, Western blot using specific antibody against the E2F4 protein.

Summary of work done until beginning of this reporting year (Upto March 31, 2012)
To find out the role of MLL complex in cell cycle regulation, in our previous report we could establish the WDR5 siRNA transfection efficiency by techniques such as RT-PCR and Western-blot. Previous studies have shown that although all four core-components are essential for a functional MLL complex, the inactivation of WDR5 results in
complete loss of activity of this complex. Therefore, depletion of WDR5 by siRNA should inactivate the whole MLL complex. We are now in the process of determining the cell cycle defects that may have appeared upon WDR5 knockdown.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
To examine the cell cycle defects that may have appeared upon WDR5 knockdown, we decided to


Figure 2. Loss of WDR5 results in delay in mitosis. (A) Series of phase contrast time-lapse images of U2OS cells captured at 1 min interval after treatment with Control or WDR5 siRNA. Selected images are shown. Arrows show cells of interest. (B) Quantification of total time taken in mitosis (prophase to telophase), in Control or WDR5 siRNA-treated U2OS cells ( $n=40$ ). The start of mitosis was determined by rounding up of the cells.
monitor the siRNA treated cells using phase contrast time-lapse microscopy. Upon observation we found that WDR5 depleted cells displayed clear delay in progression through mitosis. On an average, the WDR5 depleted cell took twice as much time (mean time taken $=39.5$ minutes) to complete mitosis compared to the control samples (mean time taken $=22$ minutes). Some cells remained in mitosis for up to 6 hours after which they were no longer imaged. As the cells were imaged under phase contrast microscopy, we could roughly determine that WDR5-depleted cells spent more time in prophase than control cells. In order to establish a more direct reason as to why WDR5 knockdown cells are stalled in prophase, we will use high-magnification microscopy to image
control and WDR5 siRNA-treated samples just before the nuclear envelope breakdown (NEB). We have initiated our imaging with control cells.
Publications

1. Zargar $Z$ and Tyagi $S$ (2012). Role of host cell factor-1 in cell cycle regulation. Transcription 3: 187-192.
2.     * Michaud J, Praz V, Faresse NJ, Jnbaptiste CK, Tyagi S, Schutz F and Herr W. HCFC1 is a common component of active human CpGisland promoters and coincides with ZNF143, THAP11, YY1 and GABP transcription factor occupancy. Genome Research (In press).

* Work done elsewhere


## अन्य वैज्ञानिक सेवाएँ / सुविधाएँ Other Scientific Services / Facilities

# LABORATORY ANIMAL FACILITY 

Faculty Co-ordinators Rashna Bhandari<br>Sanjeev Khosla<br>Other Members Hole Jayant Pundalikrao<br>Suman Komjeti

Staff Scientist \& WT-DBT India<br>Alliance Senior Fellow<br>Staff Scientist<br>Officer-In-Charge<br>Technical Assistant

## Objectives

1. To breed, maintain and supply laboratory animals to institutional scientists, while ensuring animal health and well being at all times;
2. Maintain inbred transgenic strains of mice in a controlled environment, as per CPCSEA guidelines. Breeding and experimentation of all strains of mice is undertaken in individually ventilated caging systems;
3. Assist users in procuring rodents for research, and conducting IAEC approved procedures;
4. Comply with regulatory government body requirements, in facilitating ethical research with animals, and streamlining operations to improve personnel performance and reduce operational costs.

Summary of work done until the beginning of this reporting year (Upto March 31, 2012)

The CDFD Laboratory Animal Facility (LAF) started its activities on July 1, 2011, within the premises of M/s Vimta Labs Limited, located at Genome Valley, Shameerpet, Hyderabad. Infrastructure was
established to house mice in individually ventilated cages (IVCs), and conduct standard experimental procedures. All procedures conducted on animals housed in this facility are approved by the Institutional Animal Ethics Committee (IAEC) constituted by the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Environment and Forests, Govt. of India, at M/s Vimta Labs Ltd. Until March 2012, the facility housed approximately 100 mice of each transgenic strain, Ip6k1 and Nnat.


Figure 1. Athymic nude mice bred at the CDFD Laboratory Animal Facility

| Strains | Total <br> (Male+Female) | Under Breeding <br> (Male+Female) | Supplied during <br> 2012-13 |
| :---: | :---: | :---: | :---: |
| lp6k1 | $130+100$ | $0+7$ | 133 |
| Nnat | $190+167$ | $0+10$ | 45 |
| Balb/c | $18+9$ | $5+12$ | 355 |
| C57BL/6 | $74+81$ | $0+3$ | 26 |
| Foxn17u | $14+25$ | $5+21$ | 8 |

Table 1. Strain-wise break up of mouse strains housed at LAF as on March 31, 2013, and supplied to users during 2012-13.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
During this reporting year, CDFD LAF expanded substantially to house five inbred mouse strains including lp6k1, Nnat, C57BL/6, Foxn1n, and Balb/c. These strains of mice were acquired from CPCSEA registered breeders. Mice were bred to expand the colonies and meet users' requirements. Currently this facility houses approximately 900 mice in 350 IVC cages (Table 1). During the year, 567 mice were supplied to users for IEAC approved experimentation.

Procedures conducted on these animals include blood sampling for measurement of biochemical parameters, embryo collection for the preparation of embryonic fibroblasts, tail biopsies for genotyping analysis, necropsy for histopathological analysis, antibody generation, tail vein injection, harvesting


Figure 2. Nude mice are maintained in a separate room with an IVC system and air handling unit.
of peritoneal macrophages, and tumorigenesis studies. Several mice, especially those of the transgenic strains lp6k1 and Nnat are enrolled in long-term experimentation to monitor physiological

| SI. No. | Projects in progress |
| :---: | :--- |
| 1. | Functional analysis of neuronatin's second intron by knock out strategy |
| 2. | Protocol for establishment and histopathological characterization of /p6k1 knockout mice |
| 3. | Signal transduction pathway in immune cells regulating their innate and effecter functions <br> during oxidative stress |
| 4. | Studies on the role of PNUTS in tumorigenesis in nude mice |
| 5. | Protocol for comparative bio-burden study of fifteen strains of Candida glabtrata in Balb/c <br> mice |
| 6. | Immunization of Balb/c mice for generation of antibodies against few purified recombinant <br> mycobacterial proteins |
| 7. | Studying the effect of PPE 18(Rv1196) on LPS induced endotoxaemia in mice |
| 8. | Protocol for the use of nude mice in the study of tumorigenesis |
| 9. | Protocol for generation of mouse polyclonal antibodies |
| 10. | Isolation of macrophages from Balb/c mice |
| 11. | Cryopreservation of mouse embryos by vitrification |
| 12. | Understanding the role of Rab711 in phagosome maturation and immune effector signalling |
| 13. | Protocol for establishment and histopathological characterization of Ip6k2 knockout mice |
| 14. | Protocol for establishment of transgenic mouse model to study the role of $/ p 6 k 1$ in <br> tumorigenesis |

Table 2. IAEC approved projects proposed by various groups at CDFD, in progress during 2012-13.
and biochemical responses to age and diet. The IAEC approved projects in progress during this reporting year are mentioned in Table 2.

## Future direction

In the near future, apart from continuing our current activities, our goal is to set up and establish a mouse
embryo and sperm cryopreservation facility to archive and retrieve mouse strains important for our research. We are also looking into obtaining genetically modified transgenic and knockout mouse strains from various reputed international mouse laboratories to expand our colony and ensure animal supply for our researchers as and when required.

## BIOINFORMATICS

## Head

Other Members

HA Nagarajaram
R Chandra Mohan
K Prashanthi

Staff Scientist
Technical Officer
Technical Assistant

## Objectives

1. To maintain the CDFD website, to provide web based services and e-mail services;
2. To maintain various servers, workstations, PCs, printers and other peripheral devices;
3. To maintain Institute-wide LAN as well as the internet connectivity;
4. To coordinate the procurement process of workstations, PCs, laptops, printers, other peripheral devices and software required;
5. To secure the CDFD network from security threats; and
6. To integrate Institute's network into National and International grid computing networks.

Summary of work done until beginning of this reporting year (April 1, 2011 - March 31, 2012)

* Activities related to installation, administration and maintenance of servers which provide various services, databases and computational jobs were undertaken.
* Existing PC Annual Maintenance Contract was renewed. We have also renewed the agreement for remote monitoring and managed services for Sun servers in the Data Center set up.
* Migrated existing email server to Zimbra email server.
* Upgraded 4Mbps leased line from BSNL to 8Mbps.
* Proposal from DBT for the deployment of IPv6 has been initiated.
Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)
* Activities related to installation, administration and maintenance of servers which provide various services, databases and computational jobs were undertaken.
* Procured two high end servers with 4 processors, 512 GB RAM, 5TB internal storage.
* Internet, web, email-services were provided with enhanced functionalities.
* High-end PCs, workstations, laptops, scanners and printers were procured and installed.
* PC Annual Maintenance Contract was awarded to a new vendor $\mathrm{M} / \mathrm{s}$ Bharat IT Services.
* Renewed the MoU with CDAC for availing GARUDA-grid facility.
* Upgraded the Firewall, procured additional antivirus licenses.
* Initiated the process of setting up a fail-safe server for the existing email server.


## INSTRUMENTATION

## Head

Other Members

RaghavendracharJ
RN Mishra
SD Varalaxmi
M Laxman
RMK Satyanarayana
T Ramakrishna Reddy

Staff Scientist<br>Senior Technical Officer<br>Technical Officer<br>Technical Officer<br>Technical Officer<br>Technical Assistant

## Objective

To maintain repair and service all the equipments in laboratory. To provide pre-installation requirements for new instruments and to coordinate with the manufacturers/their agents in installation and warranty service of the new instruments. Also to provide the reports on the newly arrived instruments and to follow up with the suppliers for short shipped items.
Summary of work done until the beginning of this reporting year (April 1, 2011 - March 31, 2012)

During the year 2011-12, we installed 144 new equipments like Beckman High Speed Centrifuge, Hitachi Spectrofluorimeter, Robotic Protein Crystallization System, Accuri C6 Flowcytometer, Two Color IR Imaging System, Individually Ventilated Cages for Animal house, Microscopes, PCR Machines, Nanodrop Spectrophotometers, Refrigerated Centrifuges, Orbital Shakers, Electroporators, $-80^{\circ} \mathrm{C}$ Freezer, $-20^{\circ} \mathrm{C}$ Freezers, Cold cabinets etc. and had also completed 472 work orders for repair and maintenance of various laboratory equipments.
We had successfully set up the ID card printing system for issuing identity cards instantly to all our Staff, Research Scholars and Project Staff. In addition, we were involved in organizing the audio and visual requirements for presentations in various seminars, lectures and workshops, Foundation Day
lecture series, distinguished Scientist lectures etc. held in CDFD. We were actively involved in conducting the Ramalingaswamy Fellows' Conclave and the DBT Silver Jubilee function at CDFD.

Details of progress made in the current reporting year (April 1, 2012 - March 31, 2013)

During the year 2012-13, we have installed 68 new equipments like Multi Mode Reader, Inverted Microscopes, Chemiluminescence Gel Documentation System, PCR Machines, Refrigerated Centrifuges, Shaking waterbaths, Electroporators, $-80^{\circ} \mathrm{C}$ Freezer, $-20^{\circ} \mathrm{C}$ Freezers, Cooled Incubator, Refrigerators etc. and have also completed 491 work orders for repair and maintenance of various laboratory equipments. We have successfully set up the Biometric Attendance System at both Tuljaguda and Gruhakalpa complexes registering the accurate attendance of all our employees and scholars.

In addition, we were involved in organizing the audio and visual requirements for presentations in various seminars, lectures and workshops, Foundation day lectures, distinguished Scientist lectures held in CDFD. We were actively involved in conducting the Seminar Workshop on Microbial Biology during $11^{\text {th }}$ to $14^{\text {th }}$ December 2012. We have maintained most of the equipment with maximum uptime in the laboratory. Most of the instruments are maintained by our Instrumentation staff, thereby saving on the expensive AMCs and with very little downtime of the equipment.

## प्रकाशन <br> Publications

## RESEARCH PAPERS

## A. Publications during the year 2012

1 Acharya V and Nagarajaram HA (2012). Hansa: an automated method for discriminating disease and neutral human nsSNPs. Human Mutation 33: 332-337.
2. Akhter Y, Ehebauer MT, Mukhopadhyay S and Hasnain SE (2012). The PE/PPE multigene family codes for virulence factors and is a possible source of mycobacterial antigenic variation: perhaps more? Biochimie 94: 110116.
3. Ali J, Thummala SR and Ranjan A (2012). The parasite specific substitution matrices improve the annotation of apicomplexan proteins. BMC Genomics 13 (Suppl. 7): S19.
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5. Almeida RPP, Killiny N, Newman KL, Chatterjee S, Ionescu M and Lindow SE (2012). Contribution of rpfB to cell-to-cell signal synthesis, virulence, and vector transmission of Xylella fastidiosa. Molecular Plant-Microbe Interactions 25: 453-462.
6. Angalena R, Aggarwal S, Phadke SR and Dalal A (2012). Compound heterozygote condition in beta thalassemia major due to a novel single nucleotide deletion $(-T)$ at codon 69 in association with IVS 1-5 (G>C) mutation. International Journal of Laboratory Hematology 34: e7-e9.
7. Arunkumar KP, Sahu AK, Mohanty AR, Awasthi AK, Pradeep AR, Urs SR and Nagaraju J (2012). Genetic diversity and population structure of Indian golden silkmoth (Antheraea assama). PLoS One 7: e43716.
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Rv1168c of Mycobacterium tuberculosis augments transcription from HIV-1 Long Terminal Repeat promoter. Journal of Biological Chemistry 287: 16930-16946.
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18. Dutta UR, Pidugu VK and Dalal AB (2012). Molecular and cytogenetic characterization of two patients with recurrent miscarriages and X-autosome translocation. Journal of Research in Medical Sciences 17: 572-574.
19. Dutta UR, Pidugu VK, Goud V and Dalal AB (2012). Mosaic Down syndrome with a marker: molecular cytogenetic characterization of the marker chromosome. Gene 495: 199-204.
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21. Gokul $G$ and Khosla S (2012). DNA methylation and cancer. Subcellular Biochemistry 61: 597-625.
22. Hegde SR, Rajasingh H, Das C, Mande SS and Mande SC (2012). Understanding communication signals during mycobacterial latency through predicted genome-wide protein interactions and Boolean modeling. PLoS One 7: e33893.
23. Kumar P and Nagarajaram HA (2012). A study on mutational dynamics of simple sequence repeats in relation to mismatch repair system in prokaryotic genomes. Journal of Molecular Evolution 74: 127-139.
24. Kumar R, Panigrahi I, Dalal A and Agarwal S (2012). Sickle cell anemia-Molecular diagnosis
and prenatal counseling: SGPGI experience. Indian Journal of Pediatrics 79: 68-74.
25. Mahali SK and Manna SK (2012). Beta-Dglucoside protects against advanced glycation end products (AGEs)-mediated diabetic responses by suppressing ERK and inducing PPAR gamma DNA binding. Biochemical Pharmacology 84: 1681-1690.
26. Manna SK (2012). Double-edged sword effect of biochanin to inhibit nuclear factor kappaB: suppression of serine/threonine and tyrosine kinases. Biochemical Pharmacology 83: 1383-1392.
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30. Muranjan M, Agarwal S, Lahiri K and Bashyam M (2012). Novel biochemical abnormalities and genotype in Farber disease. Indian Pediatrics 49: 320-322.
31. Muteeb G, Dey D, Mishra S and Sen R (2012). A multipronged strategy of an anti-terminator protein to overcome Rho-dependent transcription termination. Nucleic Acids Research 40: 11213-11228.
32. Narra D and Srivatsava V (2012). Quantitative competitive PCR for the detection and quantification of genetically modified cotton event MON-531. International Journal of Basic and Applied Sciences 1:92-102.
33. Padma Priya T and Dalal AB (2012). Tuberous sclerosis: diagnosis and prenatal diagnosis by MLPA. Indian Journal of Pediatrics 79: 13661369.
34. Patil SJ, Bhat V, Dalal A and Santosh JS (2012). Confirmation of the Zechi-Ceide syndrome. American Journal of Medical Genetics A 158A: 1467-1471.
35. Patil SJ, Ponnala R, Shah S and Dalal A (2012). Mosaic Trisomy 9 presenting with congenital heart disease, facial dysmorphism and pigmentary skin lesions: intricate issues of genetic counseling. Indian Journal of Pediatrics 79: 806-809.
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healing and carcinogenesis. European Journal of Cell Biology 91: 739-747.
43. Ramasarma T (2012). A touch of history and a peep into the future of the lipid-quinone known as coenzyme Q and ubiquinone. Current Science 102: 1459-1471.
44. Ramasarma T (2012). Emergence of oxyl radicals as selective oxidants. Indian Journal of Biochemistry \& Biophysics 49: 295-305.
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50. Siddiq EA, Vemireddy LR and Nagaraju J (2012). Basmati rices: genetics, breeding and trade. Agricultural Research 1:25-36.
51. Singh CP, Singh J and Nagaraju J (2012). A baculovirus-encoded microRNA (miRNA) suppresses its host miRNAs biogenesis by regulating the exportin-5 co-factor Ran. Journal of Virology 86: 7867-7879.
52. Singh YT, Mazumdar-Leighton S, Saikia M, Pant P, Kashung S, Neog K, Chakravorty R, Nair S, Nagaraju J and Babu CR (2012).

Genetic variation within native populations of endemic silkmoth Antheraea assamensis (Helfer) from Northeast India indicates need for in situ conservation. PLoS One 7: e49972.
53. Sinha DK, Nagaraju J, Tomar A, Bentur JS and Nair S (2012). Pyrosequencing-based transcriptome analysis of the Asian rice gall midge reveals differential response during compatible and incompatible interaction. International Journal of Molecular Sciences 13: 13079-13103.
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B. Publications in 2013 (Till March 31, 2013)
59. Beaulieu ED, Ionescu M, Chatterjee S, Yokota K, Trauner D and Lindow S (2013). Characterization of a Diffusible Signaling Factor (DSF) from Xylella fastidiosa. mBio 4: e00539-12.
60. Jadav RS, Chanduri MVL, Sengupta S and Bhandari R (2013). Inositol pyrophosphate synthesis by inositol hexakisphosphate kinase 1 is required for homologous recombination repair. Journal of Biological Chemistry 288: 3312-3321.
61. Jain MV, Paczulla AM, Klonisch T, Dimgba FN, Rao SB, Roberg K, Schweizer F, Lengerke C,

Davoodpour P, Palicharla VR, Maddika S and Los M (2013). Interconnections between apoptotic, autophagic and necrotic pathways: implications for cancer therapy development. Journal of Cellular and Molecular Medicine 17: 12-29.
62. Kasbekar DP (2013). Neurospora duplications and genome defense by RIP and meiotic silencing. Neurospora: Genomics and Molecular Biology. Editors: DP Kasbekar and K McCluskey, Caister Academic Press, Norfolk, UK. Pages 109-127.
63. Kavela S, Shinde SR, Ratheesh R, Viswakalyan K, Bashyam MD, Gowrishankar S, Vamsy M, Pattnaik S, Rao S, Sastry RA, Srinivasulu M, Chen J and Maddika S (2013). PNUTS functions as a proto-oncogene by sequestering PTEN. Cancer Research 73: 205214.
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66. Pandey A and Chaterjee S (2013). Signaling in plant-microbe interactions. Plant Stress 7: 52-59.
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68. Sinha A and Nagarajaram HA (2013). Effect of alternative splicing on the degree centrality of nodes in protein-protein interaction networks of Homo sapiens. Journal of Proteome Research 12: 1980-1988.
69. Subbaiah EV, Royer C, Kanginakudru S, Satyavathi VV, Babu AS, Sivaprasad V, Chavancy G, DaRocha M, Jalabert A, Mauchamp B, Basha I, Couble P and Nagaraju $J$ (2013). Engineering silkworms for resistance
to baculovirus through multigene RNA interference. Genetics 193: 63-75.
C. Publications in Press (as on March 31, 2013)
70. Bairwa G, Balusu S and Kaur R. Aspartyl proteases in human pathogenic fungi: roles in physiology and virulence. The Fungal Cell Wall. Editor: Héctor M Mora-Montes. Nova Science Publishers.
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73. Dutta UR, Pidugu VK and Dalal AB. Partial proximal trisomy 14: identification and molecular characterization in a girl with global developmental delay. Genetic Counseling.
74. Dutta UR, Pidugu VK, Goud ChV, Hoefers C, Hagemann M and Dalal A. Identification and molecular cytogenetic characterization of a novel complex Y chromosome rearrangement in a boy with disorder of sexual development. Gene.
75. Dutta UR, Ponnala R, Pidugu VK and Dalal AB. Chromosomal abnormalities in amenorrhea: a retrospective study and review of 637 patients in South India. Archives of Iranian Medicine.
76. Love JM, Prosser D, Love DR, Chintakindi KP, Dalal AB and Aggarwal S. A novel glycine decarboxylase gene mutation in an Indian family with nonketotic hyperglycinemia. Journal of Child Neurology.
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78. Mohareer K, Sahdev S and Hasnain SE. Spodoptera frugiperda FKBP-46 is a consensus
p53 motif binding protein. Journal of Cellular Biochemistry.
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81. Ranganath $P$ and Dalal $A B$. Congenital metacarpal pseudoarthrosis, cleft palate, short stature, advanced bone age, and genu valgum: a new syndrome or a variant of Devriendt syndrome? Clinical Dysmorphology.
82. Surapaneni M, Vemireddy L, Begum H, Reddy P, Neetasri C, Nagaraju J, Anwar SY and Siddiq EA. Population structure and genetic analysis of different utility types of mango (Mangifera indica L.) germplasm of Andhra Pradesh state of India using microsatellite markers. Plant Systematics and Evolution.
D. Other Publications

1. Arunkumar KP (2012). Review of: Annual Review of Genetics, 2011. Bonnie L. Bassler et al. (eds). Current Science 103: 947-949.
2. Gowrishankar J (2012). Public funding for research projects: roles of experts and finance officials in decision-making. Current Science 102: 1499.
3. Kasbekar DP (2012). Green-carding the referee and Haldane's spell. Journal of Biosciences 37: 579.
4. *Kasbekar DP (2012). Lymphohematopoietic licence: sterol C-14 reductase activity of lamin B receptor (Lbr) is essential for neutrophil differentiation. Journal of Biosciences 37: 199-201.
5. Kasbekar DP (2012). The Sad paradox: mutations with dominant and recessive phenotypes. Journal of Biosciences 37: 933936.
6. Acharya V and Nagarajaram HA (2013). Response to: Statistical analysis of missense mutation classifiers. Human Mutation 34:407.
7. Chatterjee S (2013). Review of: Annual Review of Microbiology, 2011. Susan Gottesman and Caroline S Harwood (eds). Current Science 104:653-654.
8. Kasbekar DP (2013). Myth versus mutant: story of 0 . Journal of Biosciences 38: 1 .

* Work done elsewhere


## PATENTS

## Patents Granted

1. J Nagaraju et al. Single tube multiplex assay for detection of adulterants in Basmati rice samples.
Indian Patent Application No.: 662/CHE/2006 Indian Patent No.: 251825
Date of grant: April 10, 2012

## RETRACTIONS

During this reporting period, the following papers that were published from CDFD in earlier years were retracted by the respective journals at the request of the authors:

1. Journal of Clinical Immunology (2006) 26:308322.
2. Apoptosis (2007) 12: 307-318.
3. Cell Death and Differentiation (2007) 14: 158170.
4. Journal of Cellular Biochemistry (2009) 107: 203-213.
5. Journal of Medicinal Chemistry (2009) 52: 3184-3190.
6. Breast Cancer Research and Treatment(2010) 120: 671-683.
7. Journal of Biological Chemistry (2010) 285: 5888-5895.
8. Journal of Biological Chemistry (2011) 286 : 4690-4702.
9. Journal of Biological Chemistry (2011) 286 : 7339-7347.

## मानव संसाधन विकास <br> Human Resource Development

## PhD Program

For the PhD program CDFD invites applications from highly motivated candidates willing to take up challenges in modern biology, usually in the month of March. Keeping in view the interdisciplinary nature of modern biology, the Centre especially encourages persons from different scientific disciplines to take up challenges in these areas. Those admitted as Junior Research Fellows (JRFs) are encouraged to take admission in the PhD program of Manipal University or University of Hyderabad.

The eligibility for the program is MBBS or Masters degree in any branch of Science, Technology or Agriculture from a recognized University or Institute. Candidates (other than MBBS graduates) must have cleared National Eligibility Test (NET) with valid CSIR-JRF or UGC-JRF or DBT-JRF or ICMRJRF or ICAR-JRF or INSPIRE-PhD or JEST or GATE (All India top 50 ranks of all Chemistry, Life Sciences and Biotechnology streams). Those who have appeared for their final semester examination, but are awaiting results, are also eligible to apply. Those with independent Senior Research Fellowships (SRF) from CSIR can also apply. As the number of applicants outnumbers the seats available each year by a ratio of 1:40 or more, eligible candidates are invited for a written examination followed by interviews of short-listed candidates.
As of March 31, 2013 the Centre has 95 Research Scholars working for their doctorates in different
areas of research. In the reporting year 5 of the Research Scholars have completed PhD and are pursuing careers in science elsewhere in India or abroad.

## Postdoctoral Program

In addition to the JRF program, the Centre also carries out training at the post-doctoral level. The post-doctoral fellows are funded through the extramural grants that CDFD receives. Some postdoctoral fellows are also selected competitively by the DST fast track young scientist scheme, or the DBT post-doctoral fellowship program.

## Summer Training Program

CDFD provides admissions to summer training program to those students who are supported either by the Indian Academy of Science, Bangalore or Jawaharlal Nehru Centre for Advanced Scientific Research, Bangalore or the Kishore Vigyanik Protsahan Yojna, New Delhi. In the reporting year 22 students received summer training at the Centre.

## Training for students from BITS, Pilani

CDFD has an agreement with BITS, Pilani to provide project training to their M.Sc. students. Under this programme, the students spend 6 months to 1 year at CDFD and work on active projects being carried out here. The project work helps the students in gaining hands-on experience in modern biology. In the reporting year, 7 students were given the opportunity to avail training under this programme.
Research Scholars Conferred PhD Degree During the Reporting Period

| Scholar | Supervisor | Date of <br> viva voce <br> examination | Title of thesis |
| :--- | :--- | :--- | :--- |
| Khalid Hussain Bhat | Sangita Mukhopadhyay | 27.04 .2012 | Regulatory role of Mycobacterium tuberculosis PPE proteins <br> on proinflammatory signaling pathway and activation of HIV-1 <br> LTR |
| Muley Vijay Kumar | Akash Ranjan | 26.09 .2012 | Improved computational prediction and analysis of protein- <br> protein interaction networks |
| G Sreejit | Sangita Mukhopadhyay | 11.10 .2012 | Functional characterization of Mycobacterium tuberculosis <br> proteins involved in modulating macrophage functions |
| Ghazala Muteeb | Ranjan Sen | 11.12 .2012 | Studies of mechanistic aspects of antitermination of Rho <br> dependent transcription termination |
| Carmelita N Marbaniang | J Gowrishankar | 26.02 .2013 | ArgP protein of Escherichia coli: roles in osmoregulation, <br> gene regulation and inter-relationship with LysG of <br> Corynebacterium glutamicum |

## पुरस्कार एवं सम्मान <br> Awards and Honours

## AWARDS \& HONOURS

| FACULTY \& STAFF |  |
| :--- | :--- |
| Dr. M Subba Reddy | 1. Senior Innovative Young Biotechnologist Award (2013) <br> 2. Elected as Associate of the Indian Academy of Sciences, <br> Bangalore (2012) |
| Dr. Sangita Mukhopadhyay | 1. Fellowship of the Indian Academy of Sciences, Bangalore (2013) <br> 2. ICMR Kshanika Oration Award (2009) - announced in 2013 |
| Dr. J Gowrishankar | 1. Padma Shri (2013) <br> 2. Moselio Schaechter Distinguished Service Award of the <br> American Society of Microbiology (2012) |
| Ms. R Angalena | First prize for poster presentation at the 6th International <br> Conference on Genetic \& Molecular Diagnosis in Modern <br> Medicine, Hyderabad (2013) |
| Dr. Usha Dutta (with  <br> Mr. Vijay Kumar P) First prize for poster presentation \& Prof. Askell Love award for <br> best paper presentation at XI All India Conference on Cytology <br> and Genetics, Bangalore (2012) <br> Mr. Ratheesh Raman Best poster prize at 2nd Global Cancer Genome Consortium <br> Meeting at ACTREC, Mumbai (2012) <br> Mr. Vijay Gunasekaran Second prize for poster presentation at Bangalore India Bio-2012 <br> Mr. Arghya Das Best poster award at the International Immunology FIMSA <br> Conference, New Delhi (2012) <br> Mr. Khalid Hussain Bhat Best oral presentation award at the International Immunology <br> FIMSA Conference, New Delhi (2012) |  |

$$
\begin{aligned}
& \text { व्याख्यान, बैठक, कार्यशाला व } \\
& \text { अन्य महत्वपूर्ण कार्यक्रम } \\
& \text { Lectures, Meetings, Workshops } \\
& \text { and Important Events }
\end{aligned}
$$

## DISTINGUISHED VISITORS AND LECTURES

| Visitor | Title of Lecture | Date |
| :--- | :--- | :---: |
| Dr. Varsha Singh <br> Duke University, USA | Innate immune responses to bacterial pathogens: <br> control by stress response pathways and the <br> nervous system | 02.04 .2012 |
| Dr. Palani Murugan R <br> University of Cologne, <br> Germany | Sensing, shifting and degradation: regulation <br> of polyamine biosynthesis by novel mechanisms | 01.05 .2012 |
| Dr Sivakumar Vallabhapurapu <br> University of Cincinnati, USA | Novel insights into the regulation of alternative <br> NF-kB pathway: a promising step towards <br> understanding lymphoid malignancies | 02.05 .2012 |
| Dr. Vishy Aiyar <br> The University of Texas at <br> Austin, USA | Genomic views of transcriptional and post- <br> transcriptional gene regulation | 03.05 .2012 |
| Dr. Ravi S Muddashetty <br> Emory University, USA | microRNAs-dynamic modulators of neuronal <br> activity | 04.05 .2012 |
| Dr. Akanksha Chaturvedi <br> National Institute of Health, <br> USA | Integrating adaptive and innate immune receptor <br> signaling in B cells | 07.05 .2012 |
| Dr. Atanu Maiti <br> University of Maryland School <br> of Medicine, USA | Mechanism of human TDG in maintaining genetic <br> and epigenetic integrity | 21.05 .2012 |
| Dr. Sailu Yellaboina <br> C R Rao AlMSCS <br> Hyderabad, India | Integrating genomic datasets: embryonic stem <br> cells and cancer | 13.06 .2012 |
| Dr. Kiran Kulkarni <br> Institute of Cancer Research, <br> UK | Mechanistic insights into the regulation of cell <br> cycle and cell migration | 13.07 .2012 |
| Dr. Rajeshwar Rao Tekmal <br> University of Texas Health <br> Science Centre, USA | Therapeutic use of selective estrogen receptor <br> modulators in human malignancies | 17.07 .2012 |
| Dr. Deepak Kaushal <br> Tulane National Primate <br> Research Centre, USA | Genetic requirements for the survival of tubercle <br> bacilli in primate lungs | 20.07 .2012 |
| Dr. Ramanujam Srinivasan <br> University of Singapore, <br> Singapore | Fission yeast: a cellular playground for bacterial <br> cytoskeletal proteins | 31.07 .2012 |
| Dr. Sandip Kar <br> German Cancer Research <br> Center, Germany | An insight to mathematical and computational <br> modeling in biology | 31.08 .2012 |
| Dr. Nita Sachan <br> Indian School of Business, <br> Hyderabad, India | Innovation and technology commercialization: <br> what's in it for you? | 05.09 .2012 |


| Visitor |  | Date |
| :--- | :--- | :---: |
|  <br> Dr. Monika Sharma <br> German Research <br> Foundation, Hyderabad, India | Funding opportunities for Indo-German research <br> co-operation | 04.10 .2012 |
| Dr. B Ravindran <br> Institute of Life Science, <br> Bhubaneswar, India | Did malaria contribute to evolution of TLR mediated <br> inflammation in primates? | 15.10 .2012 |
| Dr. Abul Arif <br> Lerner Research Institute <br> Cleveland Clinic, USA | Noncanonical role of aminoacyl-tRNA synthetases <br> in regulation of gene expression, inflammation and <br> metabolism | 08.11 .2012 |
| Prof. Adam J Bogdanove <br> Cornell University, USA | TAL effectors of Xanthomonas: a plant pathogenic <br> bacterium delivers powerful tools to manipulate <br> the eukaryotic genome | 01.12 .2012 |
| Dr. Nick Leslie <br> University of Dundee, UK | PTEN and PI 3-kinase signaling in cancer | 03.12 .2012 |
| Dr. Syed Raza Ali <br> University of California, USA | Novel treatments against bacterial infections | 05.12 .2012 |
| Dr. Ramana Davuluri <br> The Wistar Institute, USA | Isoform-level gene regulation: implications in <br> development and disease | 10.12 .2012 |
| Dr. Devyani Haldar <br> Dr. Reddy's Institute of Life <br> Sciences, Hyderabad, India | Role of histone acetylation/deacetylation in DNA <br> metabolism | 19.12 .2012 |
| Prof. Rajendra Prasad <br> Jawaharlal Nehru University, <br> Delhi, India | A systemic study of a major multidrug ABC <br> transporter CDR1 of Candida | 11.01 .2013 |
| Prof. Ajit Varki <br> University of California, USA | Uniquely human changes in sialic acid biology: <br> implications for evolution, immunity and disease | 05.02 .2013 |
| Dr. James Chelliah <br> Scripps Research Institute, <br> USA | Pathogenic SYNGAP1 mutations impair cognitive <br> development by disrupting the maturation of <br> dendritic spines | 06.02 .2013 |
| Dr. Paras K Anand <br> St. Jude Children's Research <br> Hospital, USA | Nod-like receptors in pathogen recognition and <br> host defense | 18.02 .2013 |
| Dr. Amitabha Majumdar <br> Stowers Institute for Medical <br> Research, USA | The role of a self-sustaining amyloidogenic protein <br> in persistence of memory | 25.02 .2013 |
| Dr. Deepti Jain <br> National Centre for Biological <br> Sciences, Bangalore, India | Functional complexes of prokaryotic transcription <br> modulators: structures and mechanisms | 26.03 .2013 |

## IMPORTANT EVENTS

| Event | Partnering Institutions | Date |
| :---: | :---: | :---: |
| Institutional Bioethics Committee Meeting |  | 19.04.2012 |
| Arrival of Prof Kazuei Mita as Distinguished Visiting Professor from NIAS, Japan |  | 09.05.2012 |
| Exposure visit for in-service Biology teachers of Kendriya Vidyalaya Sangathan |  | 17.05.2012 |
| Fire drill |  | 30.05.2012-31.05.2012 |
| Summer Trainee's Colloquium |  | 22.06.2012 |
| MoU with Government of Andhra Pradesh (Crime Investigation Department and Andhra Pradesh Forensic Science Laboratory) to provide DNA fingerprinting services and training to the state | APFSL, CID and CDFD | 11.07.2012 |
| Visit of IPS officers under the Vertical Interaction Course organized by ASCI, Hyderabad |  | 01.08.2012 |
| $14^{\text {th }}$ Meeting of CDFD Research Area Panels-Scientific Advisory Committee (RAP-SAC) |  | 03.08.2012-04.08.2012 |
| Independence Day celebrations |  | 15.08.2012 |
| Official Language Implementation Committee (OLIC) Meeting |  | 22.08.2012 |
| Hindi Pakhwada Celebrations |  | 03.09.2012-14.09.2012 |
| Education tour by B.Sc. students from Avinasilingam University for Women, Coimbatore |  | 21.09.2012 |
| Dinner hosted in honour of Prof. Jules A Hoffmann, Nobel Laureate in Physiology or Medicine (2011) | IFCPAR and CDFD | 10.10.2012 |


| Event | Partnering Institutions | Date |
| :---: | :---: | :---: |
| Video-shooting for a science popularization television serial titled 'Temples of Modern India' | Pulse Media Pvt. Ltd., Vigyan Prasar and CDFD | 10.10.2012-11.10.2012 |
| Education tour by students from Dr DY Patil University, Navi Mumba |  | 12.10.2012 |
| Mini-Symposium | Wellcome Trust-DBT <br> India Alliance and CDFD | 13.10.2012 |
| Exposure visit for students and faculty from KV Pendharkar College, Pune |  | 29.10.2012 |
| $26^{\text {th }}$ Meeting of CDFD Finance Committee |  | 31.10.2012 |
| $20^{\text {th }}$ Meeting of CDFD Building Committee |  | 31.10.2012 |
| $32^{\text {nd }}$ Meeting of CDFD Governing Council |  | 31.10.2012 |
| $17^{\text {th }}$ Meeting of CDFD Society |  | 08.12.2012 |
| Seminar Workshop on Microbial Biology | CCMB and CDFD | 11.12.2012-14.12.2012 |
| Republic Day celebrations |  | 26.01.2013 |
| CDFD Foundation Day celebrations |  | 29.01.2013 |
| Educational tour by students of M.Sc.(Nursing) from Nizam's Institute of Medical Sciences, Hyderabad |  | 15.02.2013 |
| Educational tour by scholars from the Department of Biotechnology, Kathmandu University, Nepal |  | 25.02.2013 |
| Exposure visit for post graduate students of Microbiology from Aurora's Degree and P.G. College, Hyderabad |  | 25.03.2013 |

## सी डी एफ डी कर्मचारियों की विदेशों में प्रतिनियुक्ति Deputations Abroad of CDFD Personnel

DEPUTATIONS ABROAD - FACULTY \& STAFF

| Faculty/Staff | Period | Country of Visit and Purpose |
| :---: | :---: | :--- |
| JGowrishankar | 13.06 .2012 to 21.06.2012 | USA: (i) Visit to the University of Pittsburgh, <br> Pittsburg as part of NIH-funded collaboration <br> between the University and CDFD (ii) to <br> attend the Annual General Meeting of the <br> American Society for Microbiology (ASM) at <br> San Francisco where he was conferred with the <br> Moselio Schaechter Distinguished Service <br> Award of the ASM (iii) to visit the University of <br> California. <br> USA: (i) Presentation of research work at the |
|  | 16.08 .2012 to 28.08.2012 |  |
| Cold Spring Harbor Laboratory (CSHL) |  |  |
| meeting on "Bacteria, Archaea \& Phages" |  |  |
| (ii) to visit the University of Illinois College |  |  |
| of Medicine at Chicago, The Baylor College |  |  |
| of Medicine at Houston, and Columbia |  |  |
| University Medical Center at New York. |  |  |
| Taiwan: Visit to the laboratory of Prof. Sue |  |  |
| Lin-Chao at the Institute of Molecular Biology, |  |  |
| Academia Sinica, Taipei. |  |  |


| Faculty/Staff | Period | Country of Visit and Purpose |
| :--- | :--- | :--- |
| MD Bashyam | 31.03 .2012 to 12.04.2012 | USA: (i) AACR annual meeting in Chicago <br> (ii) to interact with Dr. Ramana Davuluri at the <br> Wistar Institute at Philadelphia <br> (iii) to meet Dr. YD Ramu at the University of <br> Pennsylvania. <br> USA: Visit to the laboratory of Dr. Ramana <br> Davuluri at Wistar Institute, Philadelphia to <br> learn next generation sequencing under the <br> ICMR International fellowship program. |
| From 04.01.2013 |  |  |
| (for 5 months) |  |  |\(\left|$$
\begin{array}{ll}\text { HA Nagarajaram } & 08.09 .2012 \text { to 13.09.2012 }\end{array}
$$ \begin{array}{l}Switzerland: European Conference on <br>


Computational Biology - 12 at Basel.\end{array}\right|\)| Rupinder Kaur | 27.03 .2012 to 04.04.2012 |
| :--- | :--- | | USA: 11th ASM Conference on Candida and |
| :--- |
| Candidiasis in San Francisco, California. |
| Germany: 18th Congress of the International |
| Society for Human and Animal Mycology 2012 |
| at Berlin. |


| Faculty/Staff | Period | Country of Visit and Purpose |
| :--- | :--- | :--- |
| Arun Kumar KP | 21.07 .2012 to 01.08.2012 | USA: Workshop on "Molecular evolution" at the <br> Marine Biological Laboratory, Woods Hole, <br> Massachusetts. |
| Czech Republic: Visit to the Institute of |  |  |
| Entomology, Budejovice for collaborative |  |  |
| research on the project entitled "Comparative |  |  |
| genetic analysis of sex chromosomes and |  |  |
| sex determining genes in silkmoths". |  |  |
| France: Workshop on "Lepidoptera adaptation: |  |  |
| biology and genome" followed by a meeting on |  |  |
| Spodoptera frugiperda. |  |  |
| Japan: Visit to the University of Tokyo, Tokyo |  |  |
| and National Institute of Agro-biological |  |  |
| Sciences, Tsukuba under Indo-Japan |  |  |
| Cooperative Science Programme. |  |  |$|$| MV to 20.10.2012 |
| :--- | :--- | :--- |

DEPUTATIONS ABROAD - STUDENTS

| Name of the Scholar | Period | Country of Visit and Purpose |
| :--- | :--- | :--- |
| Jamshaid Ali | 14.05 .2012 to 18.05.2012 | Germany: 8th Annual BioMalPar\|EVIMalaR <br> Conference on "Biology and Pathology of the <br> Malaria Parasite". |
| Gaurav Bairwa | 31.07 .2012 to 05.08.2012 | USA: GSA meeting on "Yeast Genetics and <br> Molecular Biology". |
| Pandilla Ramaswamy | 14.08 .2012 to 18.08.2012 | USA: Conference on "Mechanisms and Models <br> of Cancer". |
| L Shanthy | 21.08 .2012 to 25.08.2012 |  <br> Phages". |
| Carmelita N <br> Marbaniang | 03.09 .2012 to 05.09.2012 | United Kingdom: SGM Autumn Conference <br> 2012. |
| Anupam Sinha | 08.09 .2012 to 12.09.2012 | Switzerland: 11th European Conference of <br> Computational Biology. |
| Vandana | 22.09 .2012 to 24.10.2012 | Japan: Visit to the National Institute for Basic <br> Biology to carry out replications for constructing <br> 3D-pool of silkworm BAC library for PCR <br> screening of W chromosome. |
| Anujit Sarkar | 25.11 .2012 to 30.11.2012 | Thailand: 4th Asian Forensic Sciences <br> Network Annual Meeting and Symposium 2012. |
| Maruti Nandan Rai | 13.01 .2013 to 18.01.2013 | USA: Gordon Research Conference- <br> Immunology of Fungal Infections. |
| Sawanth S Kumar | 03.03 .2013 to 30.03.2013 | Japan: Visit to the University of Tokyo and <br> National Institute of Agrobiological Sciences <br> under DST sponsored Indo-Japan Cooperative <br> Science Programme. |

# सीडीएफडी के संकाय एवं अधिकारी Faculty and Officers of CDFD 

## SCIENTIFIC GROUP LEADERS (FACULTY)

Dr. J Gowrishankar
Dr. J Nagaraju (deceased 31.12.2012)
Dr. DP Kasbekar
Dr. Ranjan Sen
Dr. Sunil Kumar Manna
Dr. Sangita Mukhopadhyay
Dr. MD Bashyam
Dr. HA Nagarajaram
Dr. Akash Ranjan
Dr. Sanjeev Khosla
Dr. Gayatri Ramakrishna (till 28.09.2012)
Dr. Rupinder Kaur
Dr. Ashwin B Dalal
Dr. Rashna Bhandari
Dr. Madhusudan R Nandineni
Dr. Subhadeep Chatterjee
Dr. Abhijit A Sardesai
Dr. R Harinarayanan
Dr. Shweta Tyagi
Dr. Rohit Joshi
Dr. MV Subba Reddy
Dr. Arun Kumar KP

## ADJUNCT FACULTY

Prof. EA Siddiq
Prof. T Ramasarma
Prof. Anuradha Lohia
Dr. Renu Wadhwa
Dr. Prajnya Ranganath
Dr. Shagun Agarwal
OTHER GROUP LEADERS
Mr. Raghavendrachar J
Ms. M Kavita Rao
Dr. Ankkur Goel
SENIOR ADMINISTRATIVE STAFF
Mr. J Sanjeev Rao
Mr. B Jagannathacharyulu

## केन्द्र की समितियाँ

(31.03.2013 तक)

## Committees of the Centre

(As on 31.03.2013)

## MEMBERS OF CDFD SOCIETY

## Shri S Jaipal Reddy

Hon'ble Minister for S\&T and Earth Sciences

## Prof K VijayRaghavan

Secretary, DBT, New Delhi

Prof Samir K Brahmachari
Director General, CSIR, New Delhi

## Prof P Balaram

Director, IISc, Bangalore

## Prof VS Chauhan

Director, ICGEB, New Delhi
Prof Dipankar Chatterji
IISc, Bangalore

Shri MK Sharma
Addl. Secretary, Ministry of Law, New Delhi
(Nominee of Joint Secretary \& Legal Adviser, MoL)

## Shri Sanjay Goel

Director (Finance), DBT, New Delhi
(Nominee of Joint Secretary \& Financial Advisor, DBT)
Shri SP Sharma
Principal Scientific Officer BPR\&D, New Delhi
(Nominee of DG, BPR\&D)
Joint Secretary (PM)
Ministry of Home Affairs, New Delhi

## Dr Suman Govil

Advisor, DBT, New Delhi

Director, CDFD, Hyderabad

President

Member

Member (Ex-officio)

Member (Ex-officio)

Member

Member

Member (Ex-officio)

Member (Ex-officio)


Member (Ex-officio)

- 

Member (Ex-officio)

Member (Ex-officio)

Member Secretary

## MEMBERS OF CDFD GOVERNING COUNCIL

## Prof K VijayRaghavan

Secretary, DBT, New Delhi
Prof Samir K Brahmachari
Director General, CSIR, New Delhi
Prof P Balaram
Director, IISc, Bangalore
Prof VS Chauhan
Director, ICGEB, New Delhi
Prof Dipankar Chatterji
IISc, Bangalore

## Shri MK Sharma

Addl. Secretary, Ministry of Law, New Delhi
(Nominee of Joint Secretary \& Legal Adviser)

## Dr CN Bhattacharya

Ministry of Home Affairs, New Delhi
(Nominee of Joint Secretary (PM))

## Ms Anuradha Mitra

Joint Secretary \& Financial Advisor
DBT, New Delhi
Dr JR Gaur
Principal Scientific Officer, BPR\&D, New Delhi
(Nominee of DG, BPR\&D)

## Dr Suman Govil

Advisor, DBT, New Delhi - Member (Ex-officio)
Dr J Gowrishankar
Director, CDFD, Hyderabad

Chairperson

Member (Ex-officio)

Member (Ex-officio)

Member

Member

Member (Ex-officio)

Member (Ex-officio)

Member (Ex-officio)

Member (Ex-officio)

Member Secretary

## MEMBERS OF CDFD RESEARCH AREA PANELS SCIENTIFIC ADVISORY COMMITTEE (RAP-SAC)

## Prof P Balaram

Chairman
Director, IISc, Bangalore
Dr Ramakrishna Ramaswamy
UoH, Hyderabad

Dr Veena Parnaik
CCMB, Hyderabad

## Dr SK Apte

Member
BARC, Mumbai
Dr Ghanshyam Swarup
Member
CCMB, Hyderabad
Dr Sandhya S Visweswaraiah
Member
IISc, Bangalore
Dr Usha Vijayraghavan
Member
IISc, Bangalore
Prof Sanjeev Galande
Member
IISER, Pune
Dr Chetan E Chitnis
Member
ICGEB, New Delhi
Dr Jaya Sivaswami Tyagi
Member
AllMS, New Delhi

Dr Joyoti Basu
Member
Bose Institute, Kolkata
Dr Debasisa Mohanty
Member
NII, New Delhi
Dr MK Mathew
Member
NCBS, Bangalore
Dr Shubha R Phadke
Member
SGPGI, Lucknow

Prof Umesh Varshney
Member
IISc, Bangalore

## Dr Suman Govil

Member
DBT, New Delhi
(Nominee of DBT)
Dr KV Prabhu
Member
NBPGR, New Delhi
(Nominee of Director General, ICAR)
$\begin{array}{lll}\text { Dr Vijay Kumar } & \text { - } & \text { Member } \\ \text { ICMR, New Delhi } & & \\ \text { (Nominee of Director General, ICMR) } & \end{array}$

Dr S Sathyan
Member
CFSL, Hyderabad
(Nominee of Ministry of Home Affairs)
Dr J Gowrishankar - Member Secretary
Director, CDFD, Hyderabad

## members of cifd academic committee

## Prof AS Raghavendra

Chair
Dean, School of Life Sciences
University of Hyderabad, Hyderabad

Prof Anil K Tyagi
University of Delhi, South Campus, New Delhi

## Dr K Satyamoorthy

Director, Manipal Life Sciences Centre Manipal University, Manipal

Dr DP Kasbekar
Haldane Chair, CDFD, Hyderabad

Dr Ranjan Sen
Staff Scientist, CDFD, Hyderabad
Dr Sanjeev Khosla
Member Convenor

Staff Scientist \& Co-ordinator (Academics)
CDFD, Hyderabad

## MEMBERS OF THE INSTITUTIONAL BIO-SAFETY COMMITTEE

* Dr J Nagaraju

Staff Scientist, CDFD, Hyderabad
(Nominee of Director, CDFD)

## Dr Rupinder Kaur

Staff Scientist, CDFD, Hyderabad

## Dr N Madhusudan Reddy

Staff Scientist, CDFD, Hyderabad
Dr Ashwin Dalal
Staff Scientist, CDFD, Hyderabad
Dr Imran Siddiqi
Scientist, CCMB, Hyderabad

## Dr S Shivaji

Scientist, CCMB, Hyderabad

Chairperson

CDFD Expert

CDFD Expert

Member with medical qualifications

Outside Expert

DBT Nominee

## MEMBERS OF CDFD BUILDING COMMITTEE

\author{

Prof VS Chauhan <br> Chairman <br> Director, ICGEB, New Delhi <br> | Dr J Gowrishankar | - | Member |
| :--- | :--- | :--- |
| Director, CDFD, Hyderabad |  |  | <br> Shri S Raghavan <br> Joint Secretary, DBT, New Delhi <br> Shri VH Rao <br> Senior Consultant, NIAB, Hyderabad <br> Shri J Sanjeev Rao <br> Head-Administration, CDFD, Hyderabad <br> Shri BJ Acharyulu <br> Head-F\&A, CDFD, Hyderabad <br> Shri K Ananda Rao <br> Senior Consultant (Engg.), CDFD, Hyderabad <br> Member Convenor

}

## MEMBERS OF CDFD MANAGEMENT COMMITTEE

Dr J Gowrishankar
Director, CDFD, Hyderabad
Dr J Nagaraju
Staff Scientist, CDFD, Hyderabad

## Dr DP Kasbekar

Haldane Chair, CDFD, Hyderabad

## Dr Ranjan Sen

Staff Scientist, CDFD, Hyderabad
Dr MV Subba Reddy
Staff Scientist, CDFD, Hyderabad
Shri BJ Acharyulu
Head-F\&A, CDFD, Hyderabad
Shri J Sanjeev Rao
Head-Administration, CDFD, Hyderabad

Chairman

Member (deceased Dec. 31, 2012)

Member (since Jan. 2013)

Member

Member

Member

Member Convenor

## mEMBERS OF CDFD FINANCE COMMITTEE

Dr VS Chauhan
Director, ICGEB, New Delhi
Ms Anuradha Mitra
JS\&FA, DBT, New Delhi
Dr Suman Govil
Advisor, DBT, New Delhi
(Nominee of Senior Scientist, DBT, New Delhi)
Prof Dipankar Chatterji
IISc, Bangalore
Dr J Gowrishankar
Director, CDFD, Hyderabad

## Shri BJ Acharyulu

Head-F\&A, CDFD, Hyderabad

Chairman

Member (Ex-officio)

Member (Ex-officio)

Member

Member

Member Secretary

## MEMBERS OF SEXUAL HARASSMENT COMPLAINTS COMMITTEE

## Dr Gayatri Ramakrishna

Staff Scientist, CDFD, Hyderabad

## Shri J Sanjeev Rao

Head-Administration, CDFD, Hyderabad
Ms V Naga Sailaja
Technical Officer, CDFD, Hyderabad

## Ms MV Sukanya

Technical Officer, CDFD, Hyderabad
Shri MSA Zaman Khan
Section Officer, CDFD, Hyderabad

## Ms P Jamuna

Gramya Resource Centre for Women, Hyderabad
(Representing an NGO)

Chairperson (till Sep. 2012)

Member

Member

Member

Member

Member

## सूचना अधिकार अधिनियम, 2005 का परिपालन Implementation of RTI Act, 2005

IMPLEMENTATION OF RTI ACT, 2005

Quarter: ${ }^{\text {nd }}$ Quarter Year 2012-2013

| Progress during Quarter |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Opening <br> Balance as on beginning of $2^{\text {nd }}$ quarter | No. of applications received as transfer from other PAs u/s 6(3) | Received during the quarter (including cases transferred to other PAs) | No. of cases transferred to other PAs u/s 6(3) | Decisions where requests/appeals rejected | Decisions where requests/appeals accepted |
| Requests | 3 | 0 | 16 | 0 | 0 | 15 |
| First Appeals | 0 | N/A | 2 | N/A | 1 | 1 |
|  | Total No. of CAPIOs designated |  | Total No. of CPIOs designated |  | Total No. of AAs designated |  |
|  | 0 |  | 1 |  | 1 |  |


| Block II - Details about fee collected, penalty imposed and disciplinary action taken |  |  |  |
| :---: | :---: | :---: | :---: |
| Registration Fee Collected <br> (in Rs.) u/s 7(1) | Addl. Fee Collected (in Rs.) <br> u/s 7(3) | Penalty Amount Recovered <br> (in Rs.) as directed by CIC <br> $\mathrm{u} / \mathrm{s} \mathrm{20(1)}$ | No. of cases where <br> disciplinary action taken <br> against any officer u/s 20(2) |
| 160 | 98 | 0 | 0 |

Quarter: 3 ${ }^{\text {rd }}$ Quarter Year 2012-2013

| Progress during Quarter |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Opening Balance as on beginning of $3^{\text {rd }}$ quarter | No. of applications received as transfer from other PAs u/s 6(3) | Received during the quarter (including cases transferred to other PAs) | No. of cases transferred to other PAs u/s 6(3) | Decisions where requests/appeals rejected | Decisions where requests/appeals accepted |
| Requests | 4 | 0 | 5 | 0 | 4 | 5 |
| First Appeals | 0 | N/A | 3 | N/A | 2 | 0 |
|  | Total No. of CAPIOs designated |  | Total No. of CPIOs designated |  | Total No. of AAs designated |  |
|  | 0 |  | 1 |  | 1 |  |


| Details about fee collected, penalty imposed and disciplinary action taken |  |  |  |
| :---: | :---: | :---: | :---: |
| Registration Fee Collected <br> (in Rs.) u/s 7(1) | Addl. Fee Collected (in Rs.) <br> u/s 7(3) | Penalty Amount Recovered <br> (in Rs.) as directed by CIC <br> $\mathrm{u} / \mathrm{s} \mathrm{20(1)}$ | No. of cases where <br> disciplinary action taken <br> against any officer u/s 20(2) |
| 50 | 0 | 0 | 0 |

Quarter: 4 ${ }^{\text {th }}$ Quarter Year 2012-2013

| Progress during Quarter |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Opening Balance as on beginning of $4^{\text {th }}$ quarter | No. of applications received as transfer from other PAs u/s 6(3) | Received during the quarter (including cases transferred to other PAs) | No. of cases transferred to other PAs u/s 6(3) | Decisions where requests/appeals rejected | Decisions where requests/appeals accepted |
| Requests | 0 | 1 | 3 | 0 | 0 | 4 |
| First Appeals | 0 | N/A | 1 | N/A | 1 | 0 |
|  | Total No. of CAPIOs designated |  | Total No. of CPIOs designated |  | Total No. of AAs designated |  |
|  | 0 |  | 1 |  | 1 |  |


| Details about fee collected, penalty imposed and disciplinary action taken |  |  |  |
| :---: | :---: | :---: | :---: |
| Registration Fee Collected <br> (in Rs.) u/s 7(1) | Addl. Fee Collected (in Rs.) <br> $\mathrm{u} / \mathrm{s} 7(3)$ | Penalty Amount Recovered <br> (in Rs.) as directed by CIC <br> $\mathrm{u} / \mathrm{s} \mathrm{20(1)}$ | No. of cases where <br> disciplinary action taken <br> againstany officer u/s 20(2) |
| 30 | 0 | 0 | 0 |

## बजट एवं वित्त <br> Budget and Finance

## CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS HYDERABAD

## Budget \& Finance 2012-13

## Sources of Funds

The Financial resources of the Centre are the Core Plan Grant-in Aid provided by the Department of Biotechnology, Government of India as against Annual Budgetary projections made by the Institute. Other resources are in the form of Research Grants provided by various National and International agencies and also from Services rendered by CDFD. The components of the core grants are Plan (Recurring) essentially for meeting expenditures on salaries, Operating expenses etc., and Plan (NonRecurring) for meeting expenses on account of Equipments, Infrastructure and Furnishing etc.,

Receipts during the year 2012-13

| Particulars | Amount in Lakhs | Percentage- $\%$ |
| :--- | :---: | ---: |
| Plan Grant in Aid | 3900.00 | 85.33 |
| Sponsored Projects | 586.52 | 12.83 |
| CDFD Services | 35.71 | 0.79 |
| Misc Receipts | 48.20 | 1.05 |
| Total | $\mathbf{4 5 7 0 . 4 3}$ | $\mathbf{1 0 0 . 0 0}$ |

I. Application of Funds during 2012-13 (Plan Grant-in-Aid)

| S.No. | Particulars | Amount in Lakhs | Percentage- \% |
| :--- | :--- | :---: | ---: |
| $\mathbf{1}$ | Recurring |  |  |
|  | Salaries \& Wages | 949.37 | 23.16 |
|  | Operating Exp | 1556.16 | 37.96 |
|  | Total | $\mathbf{2 5 0 5 . 5 3}$ | $\mathbf{6 1 . 1 2}$ |
| $\mathbf{2}$ | Non-Recurring |  |  |
|  | Equipments, |  |  |
|  | Infrastructure \& Furnishing | 1593.81 | 38.88 |
|  | Total | $\mathbf{1 5 9 3 . 8 1}$ | $\mathbf{3 8 . 8 8}$ |
|  | Grand Total | $\mathbf{4 0 9 9 . 3 4}$ | $\mathbf{1 0 0 . 0 0}$ |

## II. Application of Funds during 2012-13 (Extra Mural Projects)

| S No | Particulars | Amount in Lakhs | Percentage- \% |
| :--- | :--- | :---: | ---: |
| $\mathbf{1}$ | Recurring |  |  |
|  | Salaries \& Wages | 318.15 | 30.94 |
|  | Operating Exp | 612.76 | 59.60 |
|  | Total | $\mathbf{9 3 0 . 9 1}$ | $\mathbf{9 0 . 5 4}$ |
| $\mathbf{2}$ | Non-Recurring |  |  |
|  | Equipments | 97.29 | 9.46 |
|  | Total | $\mathbf{9 7 . 2 9}$ | $\mathbf{9 . 4 6}$ |
|  | Grand Total | $\mathbf{1 0 2 8 . 2 0}$ | $\mathbf{1 0 0 . 0 0}$ |

## लेखा परिक्षक की रिपोर्ट Auditor's Report

## K R Srinivasan \& Co

Chartered Accountants

## AUDITOR'S REPORT

Date: 04-07-2013
The Director, Centre for DNA Fingerprinting and Diagnostics, Nampally, Hyderabad - 500001

We have audited the attached Balance Sheet of CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, Hyderabad, as at $31^{\text {st }}$ March 2013 and also the Income \& Expenditure Account for the year ended on that date annexed there to. These financial statements are the responsibility of the organization management. Our responsibility is to express an opinion on these financial statements based on our audit.

We report that:

1. We have obtained all the information and explanations, which are to the best of our knowledge and belief, were necessary for the purpose of our audit.
2. In our opinion, the organization has kept proper books of account as required by law so far, as appears from our examination of those books.
3. The Balance sheet and Income \& Expenditure account dealt with by this report is in agreement with the books of account.
4. (a) The centre has maintained accounts on Accrual basis.
(b) The Centre receives extra mural grants from various National \& International agencies for specific research activities. The Centre has a policy of allocating the overheads and transfer of expenditure of CDFD to different projects at the end of the financial year after taking into account the amount of maximum permissible limit of overheads and also based on the approved budget estimates of the respective projects during the financial year.
5. In our opinion and to the best of our information and according to the explanations given to us, the said Balance sheet and the Income \& Expenditure account read together with the notes thereon gives the required information in the manner so required and give a true and fair view.
a) In so far it relates to the Balance sheet as at $31^{\text {st }}$ March 2013 and
b) In so far as it relates to the Income \& Expenditure account excess of expenditure over income for the year ended on $31^{\text {st }}$ March 2013.
for K R Srinivasan \& Co
Chartered Accountants
$\stackrel{\text { Sd/- }}{[K}$ R SRINIVASAN]
Place:Hyderabad
Date: 04/07/2013

| BALANCE SHEET AS ON 31st MARCH, 2013 <br> CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD |  |  | (Amount-Rs.) |
| :---: | :---: | :---: | :---: |
| CORPUS/CAPITALFUND AND LIABILITIES | Schedule | Current Year | Previous Year |
| Corpus / Capital Fund | 1 | 1142536939.00 | 996575609.00 |
| Reserves and Surplus | 2 | 0.00 | 276778938.00 |
| Earmarked/Endowment funds | 3 | 6531021.00 | 50698171.00 |
| Secured Loans \& Borrowings | 4 | 0.00 | 0.00 |
| Unsecured Loans \& Borrowings | 5 | 0.00 | 0.00 |
| Deffered Credit Liabilities | 6 | 0.00 | 0.00 |
| Current Liabilities and Provisions | 7 | 64750516.00 | 64107025.00 |
| TOTAL |  | 1213818476.00 | 1388159744.00 |
| ASSETS |  |  |  |
| Fixed Assets | 8 | 932133417.00 | 1025989243.00 |
| Investments- From Earmarked/Endowment Funds | 9 | 62398273.00 | 62398273.00 |
| Investments - Others | 10 | 25159583.00 | 29159376.00 |
| CurrentAssets, Loans, Advances etc. | 11 | 194127203.00 | 270612851.00 |
| TOTAL | $\begin{array}{r} 24 \\ 25 \\ \hline \end{array}$ | 1213818476.00 | 1388159744.00 |
| Significant Accounting Policies |  |  |  |
| Contingent Liabilities and Notes on Accounts |  |  |  |
| DIRECTOR For KRSRINIVASAN \& CO <br> CDFD CHARTEREDACCOUNTANTS <br>  (KRSRINIVASAN) |  | HEAD - FINANCE \& ACCOUNTS |  |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD INCOME AND EXPENDITURE ACCOUNT FOR THE YEAR ENDING 31st MARCH, 2013 |  |  |  |
| :---: | :---: | :---: | :---: |
| INCOME | Schedule | Current Year | Previous Year |
| Income from Sales/Services | 12 | 3571262.00 | 3158471.00 |
| Grants/Subsides | 13 | 210000000.00 | 210200000.00 |
| Fees/Subscriptions | 14 | 0.00 | 0.00 |
| Income from Investments | 15 | 2768470.00 | 10566572.00 |
| Income from Royality, Publications etc. | 16 | 0.00 | 0.00 |
| Interest Earned | 17 | 700706.00 | 1254141.06 |
| Other Income | 18 | 1350941.00 | 2842632.60 |
| Increase/(decrease) in stock of Finished goods and works-in-progress | 19 | 0.00 | 0.00 |
| TOTAL (A) |  | 218391379.00 | 228021816.66 |
| EXPENDITURE |  |  |  |
| Establishment Expenses | 20 | 94843804.00 | 80052399.00 |
| Administrative Expenses | 21 | 143988338.00 | 143212816.86 |
| Expenditure on Grants, Subsides etc. | 22 | 0.00 | 0.00 |
| Interest | 23 | 0.00 | 0.00 |
| Depreciation (Net Total at the year-end -corresponding to Schedule 8) |  | 21763702.00 | 0.00 |
| Less:Transferred to Grants-in-Aid Provision For Salaries |  | $\begin{array}{r} 21763702.00 \\ 6932849.00 \\ \hline \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ |
| TOTAL (B) |  | 245764991.00 | 223265215.86 |
| Balance being excess of Income over Expenditure (A-B) |  | -27373612.00 | 674552248.38 |
| Transfer to Special Reserve (Specify each) <br> Transfer to/from General Reserve <br> BALANCE BEING SURPLUS/(DEFLICT) CARRIED TO CORPUS/CAPITALFUND SIGNIFICANTACOUNTING POLICIES <br> CONTINGENT LIABILITIES AND NOTES ON ACCOUNTS | $\begin{aligned} & 24 \\ & 25 \end{aligned}$ |  |  |
| DIRECTOR For K R SRINIVASAN \& CO <br> CDFD CHARTEREDACCOUNTANTS <br>  (K R SRINIVASAN) |  | HEAD - F | CE \& ACCOUNTS CDFD |


| CENTRE <br> RECE | R DNA FIN AND PAYME | GERPRINTIN TS ACCOUNT | ND DIAGNOSTICS, HYDERAB THE YEAR ENDED 31st MARCH 2013 |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| RECEIPTS | Current Year | Previous Year | PAYMENTS | Current Year | (Amount - Rs.) <br> Previous Year |
| 1.Opening Balances <br> a) Cash in hand <br> b) Bank Balances | 0.00 | 160550.00 | 1. Expenses <br> a) Establishment Expenses (corresponding to Schedule 20) <br> b) Administrative Expenses (corresponding to Schedule 21) | 94843804.00 143988338.00 | 80052399.00 143212816.86 |
| i) In current accounts | 12333378.80 | 55687650.25 | c) Schedule 22 | 0.00 | 0.00 |
| ii) In deposit accounts | 0.00 | 0.00 |  |  |  |
| iii) Savings accounts | 73301897.29 | 7645412.22 |  |  |  |
| 2.Grants Received |  |  | 2. Payments made against funds for various projects |  |  |
| a) From Government of India | 390000000.00 | 380200000.00 | (Name of the fund or project should beshown along with the |  |  |
| b) From State government | 0.00 | 0.00 | particulars of payments made for each project) |  |  |
| c) From other sources (details) |  |  | Projects (Annexure F) | 102820071.00 | 154943838.00 |
| (Grants for capital \& revenue |  |  | CSIR(Stipend) | 12496276.00 | 10949949.00 |
| exp. To be shown seperately) |  |  | DBT(Stipend) | 4595379.00 | 3239590.00 |
| Research Associates - CSIR(Stipend) | 13650331.00 | 5197614.00 | DST(Stipend) | 527012.00 | 247039.00 |
| Research Associates - DBT(Stipend) | 1292280.00 | 3477790.00 | ICMR(Stipend) | 2079781.00 | 1879593.00 |
| Research Associates - DST(Stipend) | 250400.00 | 250400.00 | IISC(Stipend) | 3083960.00 | 1869165.00 |
| Research Associates - ICMR(Stipend) | 2422008.00 | 665590.00 | UGC(Stipend) | 5164427.00 | 4282140.00 |
| Research Associates - IISC(Stipend) | 3693877.00 | 4385503.00 |  |  |  |
| Research Associates - UGC(Stipend) | 5473330.00 | 8348180.00 | 3. Investments and deposits made <br> a) Out of Earmarked/Endowement funds | 190000000.00 | 116714571.00 |
| Projects (Annexure-C) | 58652921.00 | 189431530.00 | b) Out of Own Funds (InvestmentsOthers) | 0.00 | 0.00 |
| 3. Income on Investments from <br> a) Earmarked/Endow. Funds | 2768470.13 | 10566572.00 | 4. Expenditure on Fixed Assets \& Capital Work-in-Progress <br> a) Purchases of Fixed Assets: |  |  |
| DIRECTOR | For K R SRINIVASAN \& CO CHARTEREDACCOUNTANTS (K R SRINIVASAN) |  |  | D - FINANCE | ACCOUNTS |
| CDFD |  |  |  |  | CDFD |
|  |  |  |  |  |  |



| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF BALANCE SHEET AS AT 31st MARCH 2013 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  | Current Year $\quad$ Previous Year |  |  |  |
| SCHEDULE1-CORPUS/CAPITAL FUND : <br> Balance as at the begining of the year Add : Contribution towards Corpus/Capital Fund CDFD Core - Plan (Non-Recurring) Capitalised portion of Capital Expenditure of projects | $\begin{array}{r}180000000.00 \\ 9729088.00 \\ \hline 27140382.00\end{array}$ | 996575609.00 189729088.00 | $\begin{array}{r} 170000000.00 \\ 15090039.00 \end{array}$ | 811485570.00 185090039.00 |
| Less : Lump Sum Depreciation For the Year 1996 to 2012 <br> Less : Depreciation For the Year 2012-2013 | $\begin{array}{\|r} 271409382.00 \\ 21763702.00 \end{array}$ | 293173084.00 |  | 0.00 |
| Add : Balance of net income/(Expenditure) transferred from the income and Expenditure Account |  | 249405326.30 |  | 0.00 |
| BALANCE AS AT THE YEAR - END |  | 1142536939.30 |  | 996575609.00 |



| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF BALANCE SHEET AS AT 31st MARCH 2013 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| SCHEDULE 3-EARMARKED/ENDOWMENT FUNDS | Current Year |  | Previous Year |  |
| (Refer Annexures) <br> (a) Opening balance of the Funds <br> (b) Additions to the Funds : <br> i. Donations /grants <br> ii. Income from investments made on account of funds <br> iii. Other additions | 58652921.00 0.00 0.00 | 50698171.20 $58652921.00$ | $\begin{array}{r} 189431530.00 \\ 0.00 \\ 0.00 \end{array}$ | $16210479.20$ $189431530.00$ |
| TOTAL (a+b) |  | 109351092.20 |  | 205642009.20 |
| (c) Utilisation/Expenditure towards objective of funds <br> (i) Capital Expenditure (Refer Annexures I \& II) <br> - Fixed Assets <br> - Others <br> - Total <br> (ii) Revenue Expenditure <br> - Salaries, Wages and allowances etc. <br> - Rent <br> - Other Expenses Total | 9551279.00 177809.00 31815150.00 0.00 61275833.00 | $\begin{aligned} & 9729088.00 \\ & 93090983.00 \end{aligned}$ | 15090039.00 0.00 33415240.00 0.00 106438559.00 | $\begin{array}{r} 15090039.00 \\ 139853799.00 \end{array}$ |
| TOTAL (c) |  | 102820071.00 |  | 154943838.00 |
| NET BALANCE AS AT THE YEAR-END [(a + b)-c] |  | 6531021.20 |  | 50698171.20 |





| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF BALANCE SHEET AS AT 31st MARCH 2013 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  | Current Year |  | Previous Year |  |
| SCHEDULE 7 - CURRENT LIABILITIES AND PROVISIONS |  |  |  |  |
| A. CURRENT LIABILITIES |  |  |  |  |
| 1. Acceptances | 0.00 |  | 0.00 |  |
| 2. Sundry Creditors | 0.00 |  | 0.00 |  |
| 3. Advances Received | 0.00 |  | 0.00 |  |
| 4. Interest accured but not due on: | 0.00 |  | 0.00 |  |
| 5. Statutory Liabilities: | 0.00 |  | 0.00 |  |
| 6. Other current Liabilities |  |  |  |  |
| CDFD.CP Fund A/C(Annexure-G) | 35805401.67 |  | 29159376.32 |  |
| Out Standing Liabilities | 1520556.00 |  | 11482970.44 |  |
| Collaboration -Workshop Funds | 11300000.00 |  | 15302674.50 |  |
| House Building Advance | 95087.00 |  | 95087.00 |  |
| TDS | 478747.00 |  | 386081.00 |  |
| Income Tax | 37355.00 |  | 76988.00 |  |
| Works Tax | 234688.00 |  | 234176.00 |  |
| LIC | 2550.00 |  | 2550.00 |  |
| GSLI | 44390.00 |  | 27079.00 |  |
| Others (I-Remittances) | 178985.00 |  | 0.00 |  |
| EMD | 2898534.00 |  | 3138534.00 |  |
| Security Deposit | 1708475.00 |  | 1639975.00 |  |
| Workshop \& Conference | 3161.00 |  | 0.00 |  |
| Royalty \& Consultancy | 2254740.00 |  | 1548122.00 |  |
| Professional Tax | 99187.00 |  | 3302.00 |  |
| Lab Security Deposit \& Hostel Security Deposit | 1155810.00 | 57817666.67 | 1010110.00 |  |
| TOTAL (A) |  | 57817666.67 | 64107025.26 |  |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF BALANCE SHEET AS AT 31st MARCH 2013 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  | Current Year |  | Previous Year |  |
| SCHEDULE 7 -CURRENT LIABILITIES AND PROVISIONS |  |  |  |  |
| B.PROVISIONS |  |  |  |  |
| 1. For Taxation | 0.00 |  | 0.00 |  |
| 2. Gratuity | 0.00 |  | 0.00 |  |
| 3. Superannuation/Pension | 0.00 |  | 0.00 |  |
| 4. Accumulated Leave Encashment | 0.00 |  | 0.00 |  |
| 5. Trade Warranties/Claims | 0.00 |  | 0.00 |  |
| 6. Others (Specify) | 6932849.00 | 6932849.00 | 0.00 |  |
| TOTAL (B) | 0.00 | 6932849.00 | 0.00 |  |
| TOTAL ( $\mathrm{A}+\mathrm{B}$ ) | 0.00 | 64750515.67 | 64107025.26 |  |

\begin{tabular}{|c|c|c|c|c|c|c|c|c|c|c|}
\hline \multicolumn{11}{|l|}{\multirow[t]{2}{*}{CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF BALANCE SHEET AS AT 31st MARCH 2013}} \\
\hline \& \& \& \& \& \& \& \& \& \& \\
\hline \multicolumn{11}{|l|}{(Amount - Rs.)} \\
\hline SCHEDULE 8 - FIXED ASSETS \& \multicolumn{4}{|l|}{GROSS BLOCK} \& \multicolumn{4}{|l|}{DEPRECIATION} \& \multicolumn{2}{|l|}{NET BLOCK} \\
\hline \& Cost/valuation as at beginning of the year \& Additions during the year \& Deductions during the year \& \[
\begin{aligned}
\& \text { Cost/Valuation } \\
\& \text { at the } \\
\& \text { yearend } \\
\& \hline
\end{aligned}
\] \& As at the beginning of the year \& On Additions during the year \& \[
\begin{array}{|c|}
\hline \text { On Deductions } \\
\text { during } \\
\text { the year }
\end{array}
\] \& Total up to the year end \& As at the Current yearend \& As at the Previous yearend \\
\hline \begin{tabular}{l}
A. FIXED ASSETS: \\
1. LAND: \\
a) Freehold \\
b) Leasehold \\
2. BUILDINGS \\
a) On Freehold Land \\
b) On Leasehold Land \\
c) Ownership Flats/Premises \\
d) Superstructures on Land not belongs to the entity \\
3. PLANT MACHINERY \& EQUIPMENT \\
4. VEHICLES \\
5. FURNITURE, FIXTURES \\
6. OFFICE EQUIPMENT \\
7. COMPUTER/PERIPHERALS \\
8. ELECTRIC INSTALLATIONS \\
9. LIBRARY BOOKS \\
10. TUBEWELLS \& WATER SUPPLY \\
11. OTHER FIXED ASSETS \\
Airconditioning works \\
Aluminium partition work \\
DG Set \\
Paintings \\
Typewriters \\
Miscellaneous non-consumables \\
Other Assets \\
EMB Net
\end{tabular} \& 3900000.00
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477308882.05
4126158.00
16445181.00
11413499.00
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15081576.00
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8857898.00 \& 0.00
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55128926.00
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532437808.05
4131158.00
16456881.00
11548344.00
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15966802.00
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9499432.00
8194901.00
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7769374.00
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4999.00 \\
585.00 \\
28361.00 \\
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0.00 \& $\begin{array}{r}3900000.00 \\ 0.00 \\ \\ 203548441.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ \\ 300239937.05 \\ 759820.00 \\ 6956864.00 \\ 3325082.00 \\ 0.00 \\ \hline 359508.00\end{array}$ \& 3900000.00
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477308882.05
4126158.00
16445181.00
11413499.00
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8857898.00 <br>
\hline TOTAL \& 537133194.05 \& 276218066.00 \& 0.00 \& 813351260.05 \& 271409382.00 \& 21763702.00 \& 0.00 \& 293173084.00 \& 520178176.05 \& 537133194.05 <br>
\hline B. CAPITAL WORK-IN-PROGRESS \& 488856048.70 \& (76900808) \& 0.00 \& 411955240.70 \& 0.00 \& 0.00 \& 0.00 \& 0.00 \& 411955240.70 \& 488856048.70 <br>
\hline \multicolumn{3}{|l|}{} \& 0.00 \& 1225306500.75 \& 271409382.00 \& 21763702.00 \& 0.00 \& 293173084.00 \& 932133416.75 \& <br>
\hline
\end{tabular}

| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF BALANCE SHEET AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
|  |  | (Amount-Rs.) |
| SCHEDULE 9-INVESTMENTS FROM EARMARKED/ENDOWMENT FUNDS | Current Year | Previous Year |
| 1. In Government Securities | 0.00 | 0.00 |
| 2. Other approved securities | 0.00 | 0.00 |
| 3. Shares | 0.00 | 0.00 |
| 4. Debentures and Bonds | 0.00 | 0.00 |
| 5. Subsidiaries and Joint Ventures | 0.00 | 0.00 |
| 6. Others (to be specified) - STDRs (Annexure-J) | 62398273.00 | 62398273.00 |
| TOTAL | 62398273.00 | 62398273.00 |




| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |
| :--- | ---: | ---: | | (Amount - Rs.) |
| :--- |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :--- | ---: | ---: |
| (Amount - Rs.) |  |  |
| SCHEDULE 14-FEES/SUBSCRIPTIONS | Current Year | Previous Year |
| 1) Entrance Fees | 0.00 | 0.00 |
| 2) Annual Fees/Subscriptions | 0.00 | 0.00 |
| 3) Seminar/Program Fees | 0.00 | 0.00 |
| 4) Consultancy Fees | 0.00 | 0.00 |
| 5) Others (Specify) | 0.00 | 0.00 |
| TOTAL | 0.00 | 0.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| SCHEDULE 15-INCOME FROM INVESTMENTS | Investment from Earmarked Fund |  | Investments-Others |  |
| (Income on Invest from Earmarked/Endowment Funds transferred to Funds) | Current Year | Previous Year | Current Year | Previous Year |
| 1) Interest: <br> a) On Govt. Securities <br> b) Other Bonds/Debentures | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \end{aligned}$ | 0.00 | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 0.00 |
| 2) Dividends: <br> a) On Shares | 0.00 | 0.00 | 0.00 | 0.00 |
| b) On Mutual Fund Securities | 0.00 | 0.00 | 0.00 | 0.00 |
| 3) Rents 0.00 | 0.00 | 0.00 | 0.00 |  |
| 4) Others (Specify) STDRs | 2768470.13 | 10566572.00 | 0.00 | 0.00 |
| TOTAL | 2768470.13 | 10566572.00 | 0.00 | 0.00 |
| TRANSFERRED TO EARMARKED/ENDOWMENT FUNDS |  |  |  |  |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
|  |  | (Amount-Rs.) |
| SCHEDULE 16-INCOME FROM ROYALITY, PUBLICATIONS ETC. | Current Year | Previous Year |
| 1) Income from Royalty | 0.00 | 0.00 |
| 2) Income from Publications | 0.00 | 0.00 |
| 3) Others (Specify) | 0.00 | 0.00 |
| TOTAL | 0.00 | 0.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OFINCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
| SCHEDULE 17-INTERESTEARNED | Current Year | Previous Year |
| 1) On Term Deposits |  |  |
| a) With Schedule Banks | 700706.00 | 963584.00 |
| b) With Non-Scheduled Banks | 0.00 | 0.00 |
| c) With Institutions | 0.00 | 0.00 |
| d) Others | 0.00 | 0.00 |
| 2) On Saving Accounts |  |  |
| a) With Schedule Banks | 0.00 | 290557.06 |
| b) With Non-Scheduled Banks | 0.00 | 0.00 |
| c) post Office Savings Accounts | 0.00 | 0.00 |
| d) Others | 0.00 | 0.00 |
| 3) On Loans |  |  |
| a) Employees/Staff |  |  |
| b) Others | 0.00 | 0.00 |
| 4) Interest on Debtors and Other Receivables | 0.00 | 0.00 |
| TOTAL | 700706.00 | 1254141.06 |
| Note :- Tax deducted at source to be indicated |  |  |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
|  |  | (Amount-Rs.) |
| SCHEDULE 18-OTHER INCOME | Current Year | Previous Year |
| 1) Profit on Sale/disposal of Assets: | 0.00 | 0.00 |
| a) Owned assets | 0.00 | 0.00 |
| b) Assets acquired out of grants, or received free of cost | 0.00 | 0.00 |
| 2) Export Incentives realized | 0.00 | 0.00 |
| 3) Fees for Miscellaneous Services | 0.00 | 0.00 |
| 4) Miscellaneous Receipts | 0.00 | 0.00 |
| 5) Other Receipts |  |  |
| Sundry Receipts | 1152808.30 | 2506525.60 |
| Application Fee | 69202.00 | 284300.00 |
| Sales Of Tender Forms | 63000.00 | 25000.00 |
| Licence Fee | 44400.00 | 0.00 |
| Interest On Computer Advance,Conveyance Advance And HBA | 21531.00 | 26807.00 |
| Leave Salary-Pension Contribution | 0.00 | 0.00 |
| Provident Fund Salwage | 0.00 | 0.00 |
| Free.Gifts-Donations | 0.00 | 0.00 |
| TOTAL | 1350941.30 | 2842632.60 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
|  |  | (Amount-Rs.) |
| SCHEDULE 19 - INCREASE/(DECREASE) INSTOCK OF FINISHED GOODS \& WORK IN PROGRESS | Current Year | Previous Year |
| -Finished Goods | 0.00 | 0.00 |
| -Work-in-progress | 0.00 | 0.00 |
| Total (a) | 0.00 | 0.00 |
| b) Less: Opening Stock |  |  |
| - Finished Goods | 0.00 | 0.00 |
| - Work-in-progress | 0.00 | 0.00 |
| Total (b) | 0.00 | 0.00 |
| NET INCREASE/(DECREASE) [a-b] | 0.00 | 0.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
| SCHEDULE 20-ESTABLISHMENT EXPENSES | Current Year | Previous Year |
| a) Salaries and Wages | 54234162.00 | 74470200.00 |
| b) Allowances and Bonus | 33816395.00 | 1597702.00 |
| c) Contribution to Provident Fund | 2112193.00 | 2282201.00 |
| d) Contribution to Other Fund (NPS) | 1788473.00 | 0.00 |
| e) Staff Welfare Expenses - Medical charges | 2801565.00 | 1398440.00 |
| f) Expenses on Employees Retirement and Terminal Benefits | 91016.00 | 303856.00 |
| g) Others (specify) | 0.00 | 0.00 |
| TOTAL | 94843804.00 | 80052399.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |  |
| :---: | :---: | :---: | :---: |
|  | EDULE 21 - OTHER ADMINISTRATIVE EXPENSES, ETC. | Current Year | Previous Year |
| a) | Purchases | 30974267.00 | 52186698.60 |
| b) | Electricity and power | 18257125.00 | 18343015.00 |
| c) | Water charges | 616153.00 | 0.00 |
| d) | Insurance | 80030.00 | 0.00 |
| e) | Repairs and maintenance | 18347984.00 | 21061051.00 |
| f) | Rent, Rates and Taxes | 20625866.00 | 17082514.00 |
| g) | Vehicles Running and Maintenance | 953329.00 | 873865.00 |
| h) | Postage, Telephone and Communication Charges | 3809722.00 | 3107079.00 |
| i) | Printing and Stationary | 1151153.00 | 1238899.00 |
| j) | Travelling and Conveyance Expenses | 6819565.00 | 6189789.26 |
| k) | Expenses on Seminar/Workshops | 1029747.00 | 611987.00 |
| I) | Subscription Expenses | 163532.00 | 21932.00 |
| m) | Expenses on Fees | 294361.00 | 348971.00 |
| n) | Auditors Remuneration | 28090.00 | 27395.00 |
| o) | Hospitality Expenses | 891110.00 | 1123155.00 |
| p) | Professional Charges | 3329870.00 | 2555345.00 |
| q) | Advertisement and Publicity | 4082079.00 | 3294255.00 |
| r) | Bank Charges | 35206.00 | 6864.00 |
| s) | Security \& Cleaning Contract Charges | 16177366.00 | 14901941.00 |
| t) | Training Course/Symposia | 23752.00 | 168269.00 |

CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD
SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 (Amount - Rs.)

| Current Year | Previous Year |
| ---: | ---: |
| 2178819.00 | 69792.00 |
| 1170.00 | 0.00 |
| 14099459.00 | 0.00 |
| 18583.00 | 0.00 |
| $\mathbf{1 4 3 9 8 8 3 3 8 . 0 0}$ | $\mathbf{1 4 3 2 1 2 8 1 6 . 8 6}$ | | 143988338.00 | 143212816.86 |
| :--- | :--- |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OFINCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
|  |  | (Amount-Rs.) |
| SCHEDULE 22 -EXPENDITURE ON GRANTS, SUBSIDES, ETC. | Current Year | Previous Year |
| a) Grants given to Institutions/Organisations | 0.00 | 0.00 |
| b) Subsidies given to Institutions/Organisations | 0.00 | 0.00 |
| TOTAL | 0.00 | 0.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD SCHEDULES FORMING PART OF INCOME \& EXPENDITURE AS AT 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
| SCHEDULE 23-INTEREST | Current Year | Previous Year |
| a) On Fixed Loans | 0.00 | 0.00 |
| b) On Other Loans (including Bank Charges) | 0.00 | 0.00 |
| c) Others | 0.00 | 0.00 |
| TOTAL | 0.00 | 0.00 |

## Schedule 24: Significant Accounting Policies, and Schedule 25: Contingent Liabilities \& Notes on Account for the period ended 31/03/2013

## 1. Method of Accounting:

a. The accounting system adopted by the organization is on "Accrual basis".
b. The organization has been allocating plan grant-in-aid under the "Non-recurring" \& "Recurring" heads.

## 2. Revenue recognition:

Income comprises of Grant-in-Aid, Internal Resources through services and interest from short term deposits. Income accounted on the basis of the Cash/DD/Cheques/Cr notes received.
3. Fixed Assets:
(a) Fixed assets are stated at cost. Cost includes freight, duties, and taxes etc.,
(b) Depreciation: Based on the recommendation of the Finance Committee and approval of the Governing Body of the Institute, Depreciation Account on Fixed Assets from the financial year 1996-97 to 2011-12 of the institute has since been prepared at the rate prevailing to the concerned Fixed Assets as specified in the Income Tax Act, 1961 on Written Down Value Method of Depreciation. The accumulated depreciation from the financial year 1996-97 to 2011-12 has since been set off against the Grant in Aid (Non Recurring) in the concerned account.
(c) Capital work in progress has been entered to the extent of the last running account bills paid.
(d) Realization on sale of obsolete/surplus fixed assets which is not required for the purpose of research activities are adjusted against capital cost.
4. Inventories:

All purchases of chemicals, glassware and other consumables have been charged to consumption at the time of purchase.
5. Foreign Currency transactions:

Foreign Currency transactions are recognized in the books at the exchange rates prevailing on the date of transaction.
6. Investments:

Investments in STDR's are stated at book values.
7. Advances:

It is observed from the objection book register that advances to suppliers for consumables \& Equipments are to be reconciled and adjustment entries are to be passed in the books of accounts.
8. The previous year balances have been regrouped/rearranged, wherever necessary.

Director, CDFD
Head Finance \& Accounts
for K R Srinivasan \& Co
Chartered Accountants

Sd/-
Place: Hyderabad
[K R SRINIVASAN]
Date: 04/07/2013

## CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD

## CLARIFICATION ON NOTES ON ACCOUNTS: 2012-13

* Notes on Accounts 1 to 2 \& 4 to 6: Method of Accounting/ Revenue recognition/Fixed Asset/ Inventories/ Foreign Currency transactions/Investments:
These are all only informatory items.
* Notes on Accounts 3: Fixed Assets:

Accumulated Depreciation for the period from 1996-97 to 2011-12 has been calculated on Written Down Value method and at the rates prevailing to the concerned Fixed Asset as specified on the Income Tax Act, 1961 and set off against the Grant-in-Aid (non-recurring) in the current financial year. The details of the Depreciation on Fixed Assets is at Schedule -8 is an integral part of the financial statements.

* Notes on Accounts 7: Advances:

The observation of the audit has been noted. The action has already been initiated to reconcile the objection book register.

B JACHARYULU
Head Finance \& Accounts
CDFD

Details of closing balances of various Earmarked / Endowment Funds (Refer Sch-3) for the year ended 31st March 2013

| Previous year | P No | Particulars | Current Year |
| :---: | :---: | :---: | :---: |
| -3110519.00 | COE-I | COE for Genetics and Genomics of silkmoths | -9645531.00 |
| -8969700.00 | COE-II | DBT Centre of Excellence for Microbial Biology | -12818181.00 |
| -630047.00 | P-03 | "Transgenesis and Genetic basis of Pathogen Resistance in the Silkworm, Bombyx Mori | -630047.00 |
| 244305.00 | P-09 | "NMITLI Project on - Latent M.Tuberculosis: New targets, Drug delivery systems, Bio enhancers \& Therapeutics" | 244305.00 |
| -28332.00 | P-10 | "Role of upstream sequence elements in Hyper activation of transcription from Baculovirus polyhedrin gene promoter" | -28332.00 |
| -335000.00 | P-100 | Effect of raective oxygen species on T-Cell immune response: An approach to understand the molecular mechanism of immunosuppression during tuberculosis - National Bioscience Award | -576590.00 |
| 13729401.00 | P-101 | Role of inositol pyrophosphates in cell physiology: Investigating the biochemical significance of protein pyrophosphorylation - Senior Fellowship | 4364267.00 |
| 82654.00 | P-102 | Understanding the role of Mycobacterium tuberculosis heat shockprotein 60 as Th1/Th2 immuno modular | -430020.00 |
| -300000.00 | P-103 | National Bioscience Award - Regulation of mast cell signaling, apoptosis and surface receptors | -600000.00 |
| -1394866.00 | P-104 | Virtual Centre of Excellence on Epigenetics | -2017875.00 |
| -90844.00 | P-105 | Cloning, Characterization and analysis of chromosomal rearrangements in human genetic disorders | -844946.00 |
| 190952.00 | P-106 | Clinical, Biochemical and molecular analysis of treatable lysosomal storage disorders | -189211.00 |
| 63600.00 | P-107 | IYBA Project - Mechanism and role of bacterial cell-cell signaling molecules in plant defense response | 435.00 |
| 69925.00 | P-108 | Establishment of EBV transformed cell lines from families with rare genetic disorders | -392965.00 |
| 315626.00 | P-109 | Molecular dissection of PI3-Kinase/Akt pathway by suing proteomics based approach: A study to identify novel potential oncogenes and tumor suppressors | 94426.00 |
| -168679.00 | P-110 | India-Japan research project title"Identification and analysis of sex determining genes in silkmoths" | -191391.00 |
| 431731.00 | P-111 | Ramalingaswami Fellowship - Refractoriness mechanism in Mosquito: cracking molecular codes at genomic scale | 550416.00 |
| 534630.00 | P-113 | Clinical and molecular genetic analysis of squamous cell carcinoma of the tongue | -1036754.00 |
| 51553.00 | P-114 | Evaluating the Calcineurin-NFAT Pathway and its regulators superoxide dismutase (SOD) AND RCAN1 (regular of Calcineurin) Down Syndrome | -450859.00 |
| 8039741.00 | P-115 | Setting up of the National Institute of Animal Biotechnology | -5.00 |
| -288420.00 | P-116 | DBT-India and AIST - Japan : Understanding molecular mechanisms controlling dual role of Ras, Sirtuins and CARF in relation to cellular proliferation and senescence: Novel Strategy for developing cancer therapeutics | -1251366.00 |
| -738605.00 | P-119 | Analysis of DNA copy number alterations in esophaeal cancer | -1132629.00 |
| 124600.00 | P-120 | Effect of reactive oxygen species on macrophage signalosome: impact on antigen presentation functions and T Cell priming responses | -600218.00 |
| -597186.00 | P-121 | Identification and characterization of PTEN regulators | -1130866.00 |
| 11479043.00 | P-122 | Understanding the role of Hox genes in anterior-posterior axis determination of the central nervous system | 13089682.00 |
| 2074056.00 | P-123 | Establish a Max Planck Partner Group for Genetic Diversity Studies at CDFD | 1151969.00 |
| 167284.00 | P-124 | Preparation and characterization of peroxometal compounds and studies and their biological significance in cellular signalling | -549916.00 |
| 154000.00 | P-125 | Mechanistic studies on the role of protein kinase Snfilk in cell cycle and cancer | -480981.00 |
| 1581615.00 | P-126 | Rho-dependent transcription termination machinery: mechanism of action | -685428.00 |
| 5052715.00 | P-127 | Systematic studies on the functional network of phosphatases in cell life and death | 4162538.00 |
| 2053587.00 | P-128 | Mechanism of iron acquisition and iron homeostasis in an opportunistic human pathogen Candida glabrata | 537771.00 |
| 306000.00 | P-129 | Discovery of bioactive natural products from microbes especially actinomycetes in niche biotopes in Manipur | 0.00 |
| 6737.00 | P-13 | "Programme to delineate gene functions in the post - genomics era by a systematic two gene knockout method" | 6737.00 |
| 4187000.00 | P-130 | Comparative genetic analysis of sex chromosomes and sex determining genes in silkmoths | 465973.00 |
| 1182935.00 | P-131 | Structural and functional studies of Acyl CoA Binding proteins from plasmodium falciparum | -768669.00 |
| 634323.00 | P-132 | Characterization of tumor suppressor function of ARIDIB, a component of the human SWI/SNF chromatin remodelling complex | -1228480.00 |
| 1549000.00 | P-133 | Investigating the role of Hox gene deformed in central nervous system patterning in Drosophila melanogaster | 969489.00 |
| 254000.00 | P-134 | Exploration of wild silk moth biodiversity in Manipur and their genetic characterization using molecular markers | -141437.00 |
| 7418200.00 | P-135 | Sys TB: A Network Program for Resolving the Intracellular Dynamics of Host Phthogen Interaction in TB Infection | 5376566.00 |
| 837200.00 | P-136 | Raf Kinase - a key target for modem-day theraphy against tumors | 77980.00 |
| 1500000.00 | P-137 | Signaling pathways involved in down regulation of proinflammatory responses by PPE18 protein of Mycobacterium tuberculosis: Implication of PPE18 as therapeutics | 685020.00 |
| 0.00 | P-138 | Co-evaluation of Dnmt3I and Genomic imprinting | 903944.00 |

CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS
Details of closing balances of various Earmarked / Endowment Funds (Refer Sch-3) for the year ended 31st March 2013

| Previous year | P No | Particulars | Current Year |
| :---: | :---: | :---: | :---: |
| 2467200.00 | P-139 | Evaluating the role of Sirtuins and epigenetic changes during cellular senescense in context of p53 status | 1223583.00 |
| 0.00 | P-140 | Development of baculovirus resistant silkworms strains through synthetic miRNA based knockdown of essential viral genes | 556091.00 |
| 0.00 | P-141 | Evaluating the functional role of PTEN interacting proteins in cell survival signaling and tumor suppression | 1463.00 |
| 0.00 | P-142 | Identification of H3K4 TRI Demethylase involved in erasing H3K4 trimethylation marks at E2F Responsive promoters | 360148.00 |
| 0.00 | P-143 | Microarray based characterisation of squamous cell carcinoma of the tongue occuring in non smokers | 146284.00 |
| 267184.00 | P-144 | Tri-National Training Program for Psychiatric Genetics | 0.00 |
| 0.00 | P-145 | H3K4 HMT family regulatescell cycle progression | 2208206.00 |
| 0.00 | P-146 | Role of MLL in ribosomal RNA transcription | 812209.00 |
| 0.00 | P-147 | The Effect of Parental Education, Ethics of Research Participation and Array Comparative Genomic Hybridization in Subjects with Mental Retardation (MR) and /or Autism | 315642.00 |
| 0.00 | P-148 | Transcriptional regulation of novel tumor suppressor genes in Pancreatic Cancer | 20326.00 |
| 0.00 | P-149 | Role of SUMOylation in the pathobiology of Candida Glabrata | 1770286.00 |
| 0.00 | P-150 | Genetic and genomic basis of the evolution of bombycid and sturniid silkmoths | 164706.00 |
| 0.00 | P-151 | Human Exome Sequencing to Identify Novel Genes for Medelian Disorders | 1993200.00 |
| 0.00 | P-153 | An attractive and promising stragegy for early cancer diagnosis through the assembly of the human cancer volatome" | 3000000.00 |
| 0.00 | P-155 | Studies on thecellular roles of calcium signalling proteins in Neurospora crassa | 335194.00 |
| -687887.00 | P-17 | "Studies on inosital-phosphate synthesis - a novel enzyme from Mycobacterium tuberculosis H37RV" - Transfer from IMTECH, Chandigarh | -687887.00 |
| -274286.00 | P-18 | "Mapping of receptor binding site on the Eythrocyte binding of malaria parasyte" | -274286.00 |
| -1888111.00 | P-20 | "Genomic Micro array R\&D Programmes on infectious diseases and Neurological Disorders" | -1888111.00 |
| 0.50 | P-22 | "Biotechnology for leather - towards cleaner processing" | 0.50 |
| -34495.00 | P-23 | "Development of PCR base assays for detection of GMO S" | -34495.00 |
| -529111.00 | P-25 | "Functional studies of Human Immuno - deficiency Virus Type-2 (HIV-2) Viral protien X (VPX)" | -529111.00 |
| -79533.00 | P-26 | Occurrence of Mutations in Non dividing cells of Escherichia Coli" | -79533.00 |
| -37624.00 | P-28 | Baculovirus resistance in transgenic silkworms | -37624.00 |
| -310302.00 | P-29 | "Development of Hospital Surveillance system by advanced diagnostics method \& Molecular DNA fingerprinting techniques" | -310302.00 |
| 2045696.00 | P-30 | Transcription termination and anti termination in E-coli | 2045696.00 |
| 746453.00 | P-31 | Role of K-ras in Lung type II epithelial cells | 746453.00 |
| -234000.00 | P-33 | "Molecular and Epidemiological characterisation of cryptosporidium - An enteric protozoon parasite" | -234000.00 |
| 26334.00 | P-34 | "Molecular analysis of lepidopteran - specific immune protiens from silkmoths" | 26334.00 |
| -283883.00 | P-35 | "Identification, Characterization and Physical mapping of Z-Chromosome linked genes of the silk worm, Bombyxmori" | -283883.00 |
| 2073896.00 | P-36 | "Development of Artificial retina using Bacterio rhodospin and genetically engineered analogues " | 2073896.00 |
| -226058.00 | P-40 | "Antioxidants as a potential immuno adjuvant in anti tuberculosis immunotherapy" | -4058.00 |
| 1873605.00 | P-41 | "Construction, characterization and analysis of expressed sequences from silkworm " | 1873605.00 |
| -2237285.00 | P-42 | "Structural and functional studies on Mycobacterium tuberculosis heat shock proteins". | -2237285.00 |
| 685906.70 | P-43 | "A generalized mechanism of transcription termination in prokaryotes: a quest for mechanism based transcription inhibitors for microbial pathogens". | 685906.70 |
| -457538.00 | P-44 | "Understanding of role of Ras and NO / iNOS signalling in promotion of hepatocellular carcinomas with persistent HBV infection" | -457538.00 |
| 605714.00 | P-45 | Specialized chromatin structures as epigenetic imprints to distinguish parental alleles". | 605714.00 |
| -1586965.00 | P-47 | Research cum Training for DRDO Programme | -1586965.00 |
| 151826.00 | P-48 | 'Molecular characterization of human liver stem cells for use in the treatment of hepatic diseases'. | 151826.00 |
| 440950.00 | P-49A | International Atomic Energy Agency (IAEA) | 308361.00 |
| -284065.00 | P-51 | "Understanding the mechanism of doxorubicin resistance in breast cancer celline MCF-7" | -284065.00 |
| -1231118.00 | P-52 | "Nucleo Cytoplasmic transport of HIV - 1 Vpr " | -1231118.00 |
| -37877.00 | P-54 | "Study of viability of Mycobacterium leprae in clinical samples and possibility of its presence in the environment using nucleic acid amplification techniques." | -37877.00 |
| 224.00 | P-55 | "Identification of DNA Markers for baculovirus resistance in silkworm, Bombyx mori" | 224.00 |
| -1231164.00 | P-56 | "Genetics of transcription-replication interplay and of stress adaptation in bacteria" | -1231164.00 |
| -2215024.00 | P-59 | "An integrated Approach towards understanding the biology of Mycobacterium tuberculosis: Genetic, biochemical, immunological and structural analyses." | -2215024.00 |

CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS
Details of closing balances of various Earmarked / Endowment Funds (Refer Sch-3) for the year ended 31st March 2013

| Amount in Rs. |  |  |  |
| :---: | :---: | :---: | :---: |
| Previous year | P No | Particulars | Current Year |
| 482124.00 | P-60 | "National Database of Prevalent Genetic Disorders in India: Development, Curation and Services" | 482124.00 |
| -280000.00 | P-61 | "Dissection of a novel phenotype of lethal accumulation of potassium in Escherichia coli mutants defective in thioredoxin/thioredoxin reductase and nucleoied protein H-NS" | -280000.00 |
| -278928.00 | P-62 | "HIV - 1 Pathogenesis: Role of Integrase in Reverse Transciption and Nuclear Transport of Viral Genome" | -278928.00 |
| -837574.00 | P-63 | "Upgradation of the existing computing infrastructure at the Bioinformatics facility at CDFD" | -837574.00 |
| -158.00 | P-64 | Biotechnology for Leather: Towards cleaner processing phase-II | -158.00 |
| -582647.00 | P-65 | "Molecular, genetic and functional analysis of the chromosomal plasticity region of the gastric pathogen Helicobater pylori" | -582647.00 |
| 18938021.00 | P-65A | APEDA-CDFD Centre for Basmati DNA Analysis | 19734821.00 |
| -681246.00 | P-66 | Human Epigenome Variation: Analysis of CpG island methylation in chromosomes 18 and Y , and in some Hox, insulin signaling and chromatin reprogramming genes | -681246.00 |
| -113545.00 | P-67 | Identification of novel Esophageal Squamous cell carcinoma (ESCC) genes by using a combination of array-based CGH and gene expression micro arrays | -113545.00 |
| -59874.00 | P-68 | Identification of High risk individual with pre-cancerous states of esophageal cancer. | -59874.00 |
| -21336.00 | P-70 | Identification of disease causing mutations in familial hypertrophic cardiomyopathy (FHC) patients from Andhra Pradesh | -21336.00 |
| 15829.00 | P-71 | Referral Centre for Genetic fidelity testing of tissue culture raised plants | 0.00 |
| -1421653.00 | P-72 | Nuances of non coding DNA near insulin-responsive genes. | -1421653.00 |
| -857136.00 | P-73 | Identification and characterization of pancreatic cancer genes located within novel localized cpy number alterations | -857136.00 |
| -10840.00 | P-75 | Preparing blueprint for the macromolecular crystallography beamline at Indus-II synchrotron source | -10840.00 |
| -50234.00 | P-76 | A study of molecular markers in childhood autism with special references to nuclear factors - $\pm$ APPA B | -50234.00 |
| 124277.00 | P-77 | Functional characterization of Mycobacterium tuberculosis PE/PPE proteins having SH3 binding domain Understanding their role in modulating macrophage functions | 124277.00 |
| 1304.00 | P-78 | Task force- IMD Newborn screening for Congenital Hypothyroidism \& Congenital Adrenal Hyperplasia: A multicentric study | 1304.00 |
| -105086.00 | P-79 | Understanding the role of AGE proteins in inducing inflammatory responses and its regulation | -105086.00 |
| -608222.00 | P-80 | Referral centre for detection of genetically modified foods employing DNA-based markets | -608222.00 |
| 143470.00 | P-81 | Reconstructing Cellular Networks: Two-component regulatory systems | 143470.00 |
| 62620.00 | P-81A | Financial assistance for award of J C Bose Fellowship to Dr J Gowrishankar | 562620.00 |
| 155859.00 | P-82 | Functional genomic analysis of Candida Glabrata-macrophage | -367721.00 |
| -1155594.00 | P-83 | Prokaryotic Transcription termination factor, Rho: Mechanism of Action and Biology | -1155594.00 |
| -126140.00 | P-83A | Understanding the mechanism of Azadirachtin-mediated cell signaling: role in anti-inflammation and anti-tumorigenesis | 0.00 |
| -1150.00 | P-84 | Preparing for vaccine efficacy trials: Baseline epidemiology, improved diagnosis, markers of protection and phase I/II trials | -1150.00 |
| -106479.00 | P-84A | Human epigenetic to the rescue of human identification process: Enriching human DNA from DNA mixture employing antibodies directed against 5-methylcytosine followed by whole genome amplification | -106479.00 |
| -1118755.00 | P-85 | IdeR associated gene regulatory network in mycobacteria | -1118755.00 |
| -65698.00 | P-87 | Comparative genomics of wild silkmoths | -65698.00 |
| 0.00 | P-88 | Introduction of anti-baculoviral property in commercial silkworm strains by expression of multiple RNAi viral targets | 218818.00 |
| -636286.00 | P-90 | Role of Yapsins in the Pathobiology of Candida Glabrata | -636286.00 |
| -1098900.00 | P-91 | DMMT3L: epigenetic correlation with cancer | -1098900.00 |
| -1260461.00 | P-92 | Swarnajayanti fellowship proj on "Designing transcription anti-terminators: a novel approach for making new inhibitors of gene expression" | -3090255.00 |
| -675810.00 | P-93/A1 | Virtual Centre of Excellence on multidisciplinary approaches aimed at interventions against tuberculosis | -661454.00 |
| -831076.00 | P-93/A2 | Virtual Centre of Excellence on multidisciplinary approaches aimed at interventions against Mycobacterium tuberculosis | -2446997.00 |
| -98464.00 | P-97 | Proteome-wide Analysis of Serine pyrophosphorylation by inositol pyrophosphates | -146870.00 |
| -63019.00 | P-98 | Role of cell - cell signaling mediated by Diffusible signaling factor (DSF) in Xanthomonas virulence | -255844.00 |
| -1261900.00 | P-99 | Role of inositol Pyrophosphates in eukaryotic cell growth, proliferation and ribosomae biogenesis | -315780.00 |
| 50698171.20 |  |  | 6531021.20 |

CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS Details of Fixed Assets Fund (Capitalised portion of Project Grants) for the year ended 31st March 2013

Amount in Rs.

| Previous year | P No | Particulars | Current Year |
| :---: | :---: | :---: | :---: |
| 11713327 | COE-I | COE for Genetics and Genomics of silkmoths | 11713327 |
| 10000000 | COE-II | DBT Centre of Excellence for Microbial Biology | 1000000 |
| 600000 | P-03 | "Transgenesis and Genetic basis of Pathogen Resistance in the Silkworm, Bombyx Mori | 60000 |
| 329289 | P-07 | "Collection of well characterised clinical samples and strains of Mycobacterium tuberculosis and development of molecular techniques for detection of drug resistant strains - Multi Centric Project" | 329289 |
| 588400 | P-09 | "NMITLI Project on - Latent M.Tuberculosis: New targets, Drug delivery systems, Bio enhancers \& Therapeutics" | 588400 |
| 47400 | P-10 | "Role of upstream sequence elements in Hyper activation of transcription from Baculovirus polyhedrin gene promoter" | 47400 |
| 17784 | P-100 | Effect of raective oxygen species on T-Cell immune response: An approach to understand the molecular mechanism of immunosuppression during tuberculosis - National Bioscience Award | 17784 |
| 6276263 | P-101 | Role of inositol pyrophosphates in cell physiology: Investigating the biochemical significance of protein pyrophosphorylation - Senior Fellowship | 10645294 |
| 681121 | P-102 | Understanding the role of Mycobacterium tuberculosis heat shockprotein 60 as Th1/Th2 immuno modular | 681121 |
| 1000000 | P-107 | IYBA Project - Mechanism and role of bacterial cell-cell signaling molecules in plant defense response | 1000000 |
| 915278 | P-109 | Molecular dissection of PI3-Kinase/Akt pathway by suing proteomics based approach: A study to identify novel potential oncogenes and tumor suppressors | 915278 |
| 268914 | P-113 | Clinical and molecular genetic analysis of squamous cell carcinoma of the tongue | 670095 |
| 294008 | P-114 | Evaluating the Calcineurin-NFAT Pathway and its regulators superoxide dismutase (SOD) AND RCAN1 (regular of Calcineurin) Down Syndrome | 475900 |
| 4580214 | P-115 | Setting up of the National Institute of Animal Biotechnology | 4580214 |
| 800000 | P-116 | DBT-India and AIST - Japan : Understanding molecular mechanisms controlling dual role of Ras, Sirtuins and CARF in relation to cellular proliferation and senescence: Novel Strategy for developing cancer therapeutics | 800000 |
| 183443 | P-118 | Construction of regulatory networks in Mycobacterium tuberculosis through analysis of gene expression data and transcription regulation predictions. (MOU with Russian Foundation) | 183443 |
| 529750 | P-12 | Molecular genetics and Functional genomics of M.Tuberculosis patient isolates in India | 529750 |
| 438084 | P-122 | Understanding the role of Hox genes in anterior-posterior axis determination of the central nervous system | 459324 |
| 101800 | P-123 | Establish a Max Planck Partner Group for Genetic Diversity Studies at CDFD | 453095 |
| 0 | P-126 | Rho-dependent transcription termination machinery: mechanism of action | 385404 |
| 2225907 | P-127 | Systematic studies on the functional network of phosphatases in cell life and death | 2897196 |
| 1334600 | P-128 | Mechanism of iron acquisition and iron homeostasis in an opportunistic human pathogen Candida glabrata | 1594393 |
| 1334600 | P-13 | "Programme to delineate gene functions in the post - genomics era by a systematic two gene knockout method" | 1334600 |
| 0 | P-133 | Investigating the role of Hox gene deformed in central nervous system patterning in Drosophila melanogaster | 474792 |
| 5163243 | P-14 | "Comparitive and functional genomics approaches for the identification and characterization of genes responsible for multi drug resistance of mycobacterium tuberculosis" | 5163243 |
| 0 | P-142 | Identification of H3K4 TRI Demethylase involved in erasing H3K4 trimethylation marks at E2F Responsive promoters | 424914 |
| 0 | P-146 | Role of MLL in ribosomal RNA transcription | 359711 |
| 6000000 | P-15 | "The Helicobacter Pylori genome programme - Genome sequencing, functional analysis and comparitive genomics of the strains obtained from Indian patients" | 6000000 |
| 1814901 | P-16 | NMITLI Project on - Latent M.Tuberculosis: New targets, Drug delivery systems, Bio enhancers \& Therapeutics | 1814901 |
| 244400 | P-17 | "Studies on inosital-phosphate synthesis - a novel enzyme from Mycobacterium tuberculosis H37RV" Transfer from IMTECH, Chandigarh | 244400 |
| 344020 | P-18 | "Mapping of receptor binding site on the Eythrocyte binding of malaria parasyte" | 344020 |
| 7246511 | P-19 | "Construction of Integrated RAPD, RFLP and Microsatellite linkage map of the Silkworm, Bombyx mori and its corelation with the Phenotypic linkage Map" | 7246511 |
| 27331134 | P-20 | "Genomic Micro array R\&D Programmes on infectious diseases and Neurological Disorders" | 27331134 |
| 5300000 | P-21 | Development of Versatile, portable software for Bio-informatics | 5300000 |
| 603747 | P-22 | "Biotechnology for leather - towards cleaner processing" | 603747 |
| 375999 | P-23 | "Development of PCR base assays for detection of GMO S" | 375999 |
| 600000 | P-25 | "Functional studies of Human Immuno - deficiency Virus Type-2 (HIV-2) Viral protien X (VPX)" | 600000 |
| 500000 | P-26 | Occurrence of Mutations in Non dividing cells of Escherichia Coli" | 500000 |
| 260367 | P-29 | "Development of Hospital Surveillance system by advanced diagnostics method \& Molecular DNA fingerprinting techniques" | 260367 |
| 3746538 | P-30 | Transcription termination and anti termination in E-coli | 3746538 |
| 3131006 | P-31 | Role of K-ras in Lung type II epithelial cells | 3131006 |
| 4857938 | P-36 | "Development of Artificial retina using Bacterio rhodospin and genetically engineered analogues " | 4857938 |
| 358470 | P-39 | "Computational analysis and functional characterization of mycobacterial protien(s) interacting with macrophase effector - APC functions - an approach to understand the molecular basis of pathogenesis of M. tuberculosis" | 358470 |
| 49738 | P-40 | "Antioxidants as a potential immuno adjuvant in anti tuberculosis immunotherapy" | 49738 |
| 3894086 | P-41 | "Construction, characterization and analysis of expressed sequences from silkworm" | 3894086 |
| 9500000 | P-42 | "Structural and functional studies on Mycobacterium tuberculosis heat shock proteins". | 9500000 |

CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS
Details of Fixed Assets Fund (Capitalised portion of Project Grants)
for the year ended 31st March 2013
Amount in Rs.

| Previous year | P No | Particulars | Current Year |
| :---: | :---: | :---: | :---: |
| 11970000 | P-43 | "A generalized mechanism of transcription termination in prokaryotes: a quest for mechanism based transcription inhibitors for microbial pathogens". | 11970000 |
| 3331377 | P-45 | Specialized chromatin structures as epigenetic imprints to distinguish parental alleles". | 3331377 |
| 416137 | P-46 | "Effect of reactive oxygen species (ROS) on immune response : Relevance in immunosuppression and Pathogenesis" | 416137 |
| 377567 | P-47 | Research cum Training for DRDO Programme | 377567 |
| 1413292 | P-48 | 'Molecular characterization of human liver stem cells for use in the treatment of hepatic diseases'. | 1413292 |
| 198095 | P-50 | "Cervical cancer prevention by multiple strategies in rural community in Andhra pradesh" | 198095 |
| 401738 | P-51 | "Understanding the mechanism of doxorubicin resistance in breast cancer celline MCF-7" | 401738 |
| 1359129 | P-52 | "Nucleo Cytoplasmic transport of HIV - 1 Vpr " | 1359129 |
| 1114495 | P-53 | Collaborative research project on molecular ecology and systematics | 1114495 |
| 1163764 | P-56 | "Genetics of transcription-replication interplay and of stress adaptation in bacteria" | 1163764 |
| 2131403 | P-57 | Improved genome annotation through a combination of machine learning and experimental methods: Plasmodium falciparum as a case study. | 2131403 |
| 63000 | P-58 | "Indo-Malaysian collaboration in Bioinformatics: Development and maintenance of a web-portal hosting databases and tools of common interest" | 63000 |
| 32974662 | P-59 | "An integrated Approach towards understanding the biology of Mycobacterium tuberculosis: Genetic, biochemical, immunological and structural analyses." | 32974662 |
| 5720800 | P-60 | "National Database of Prevalent Genetic Disorders in India: Development, Curation and Services" | 5720800 |
| 4308314 | P-62 | "HIV - 1 Pathogenesis: Role of Integrase in Reverse Transciption and Nuclear Transport of Viral Genome" | 4308314 |
| 9637574 | P-63 | "Upgradation of the existing computing infrastructure at the Bioinformatics facility at CDFD" | 9637574 |
| 600585 | P-64 | Biotechnology for Leather: Towards cleaner processing phase-II | 600585 |
| 260000 | P-65 | "Molecular, genetic and functional analysis of the chromosomal plasticity region of the gastric pathogen Helicobater pylori" | 260000 |
| 16924622 | P-65A | APEDA-CDFD Centre for Basmati DNA Analysis | 16924622 |
| 264430 | P-66 | Human Epigenome Variation: Analysis of CpG island methylation in chromosomes 18 and Y , and in some Hox, insulin signaling and chromatin reprogramming genes | 264430 |
| 622747 | P-67 | Identification of novel Esophageal Squamous cell carcinoma (ESCC) genes by using a combination of array-based CGH and gene expression micro arrays | 622747 |
| 235593 | P-69 | ICMR adhoc New Scheme Understanding the role of PE/PPE family of M tuberculosis in the activation of HIV virus type I long terminal repeat (HIV-ILTP) | 235593 |
| 1012807 | P-70 | Identification of disease causing mutations in familial hypertrophic cardiomyopathy (FHC) patients from Andhra Pradesh | 1012807 |
| 1573795 | P-71 | Referral Centre for Genetic fidelity testing of tissue culture raised plants | 1573795 |
| 45653 | P-72 | Nuances of non coding DNA near insulin-responsive genes. | 45653 |
| 1000000 | P-74 | Molecular basic of insect plant interactions in rice under the national fund for basic and strategic research in agriculture | 1000000 |
| 33672 | P-75 | Preparing blueprint for the macromolecular crystallography beamline at Indus-II synchrotron source | 33672 |
| 245266 | P-76 | A study of molecular markers in childhood autism with special references to nuclear factors - $\pm$ APPA $B$ | 245266 |
| 1543605 | P-77 | Functional characterization of Mycobacterium tuberculosis PE/PPE proteins having SH3 binding domain : Understanding their role in modulating macrophage functions | 1543605 |
| 496826 | P-79 | Understanding the role of AGE proteins in inducing inflammatory responses and its regulation | 496826 |
| 4192480 | P-80 | Referral centre for detection of genetically modified foods employing DNA-based markets development of molecular | 4192480 |
| 195728 | P-81A | Financial assistance for award of J C Bose Fellowship to Dr J Gowrishankar | 195728 |
| 1441427 | P-82 | Functional genomic analysis of Candida Glabrata-macrophage | 1480220 |
| 912255 | P-83 | Prokaryotic Transcription termination factor, Rho: Mechanism of Action and Biology | 912255 |
| 388583 | P-83A | Understanding the mechanism of Azadirachtin-mediated cell signaling: role in anti-inflammation and anti-tumorigenesis | 388583 |
| 44854 | P-84 | Preparing for vaccine efficacy trials: Baseline epidemiology, improved diagnosis, markers of protection and phase I/II trials | 44854 |
| 1430573 | P-84A | Human epigenetic to the rescue of human identification process: Enriching human DNA from DNA mixture employing antibodies directed against 5-methylcytosine followed by whole genome amplification | 1430573 |
| 374630 | P-89 | Characterization of Mycobacterium tuberculosis transcription machinery and Bacteriophage metagenomics | 374630 |
| 1376869 | P-90 | Role of Yapsins in the Pathobiology of Candida Glabrata | 1376869 |
| 932151 | P-91 | DMMT3L: epigenetic correlation with cancer | 932151 |
| 8500000 | P-92 | Swarnajayanti fellowship proj on "Designing transcription anti-terminators: a novel approach for making new inhibitors of gene expression" | 8500000 |
| 2212534 | P-93/A1 | Virtual Centre of Excellence on multidisciplinary approaches aimed at interventions against tuberculosis | 2212534 |
| 296034 | P-93/A2 | Virtual Centre of Excellence on multidisciplinary approaches aimed at interventions against Mycobacterium tuberculosis | 655403 |
| 246320 | P-95 | Construction of regulatory networks in prokaryotes through protein: Protein interaction predictions and transcription regulation predictions. (MOU with Russian Foundation) | 246320 |
| 918196 | P-97 | Proteome-wide Analysis of Serine pyrophosphorylation by inositol pyrophosphates | 966602 |
| 2783795 | P-98 | Role of cell - cell signaling mediated by Diffusible signaling factor (DSF) in Xanthomonas virulence | 2789420 |
| 2921729 | P-99 | Role of inositol Pyrophosphates in eukaryotic cell growth, proliferation and ribosomae biogenesis | 2963482 |
| 254856786 |  |  | 264585874 |


| Annexure: A F | CENTRE FOR DNA FINGERPRINTI FOR THE YEAR ENDED 31 <br> ming part of Receipts \& Paymen |  |
| :---: | :---: | :---: |
| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
|  | I-Remittances |  |
| 4740809.00 | TDS | 4976630.00 |
| 5678791.00 | Income Tax | 6114373.00 |
| 14920.00 | Works Tax | 4586.00 |
| 1102504.00 | LIC | 1335912.00 |
| 225224.00 | GSLI | 219721.00 |
| 1574630.00 | PPF | 1904410.00 |
| 546897.00 | Professional Tax | 574296.00 |
| 166864.00 | Service Tax | 1979139.00 |
| 0.00 | Others (I-Remittances) | 678505.00 |
| 14050639.00 |  | 17787572.00 |

## CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS <br> FOR THE YEAR ENDED 31st MARCH 2013

Annexure: B Forming part of Receipts \& Payment a/c

| Previous Year <br> Amount <br> Rs. |  | Current Year <br> Amount |
| ---: | :--- | ---: |
|  | Rs. |  |



| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS FOR THE YEAR ENDED 31st MARCH 2013 <br> Annexure: C Forming part of Receipts \& Payment a/c |  |  |
| :---: | :---: | :---: |
| Previous Year Amount Rs. |  | Current Year Amount Rs. |
| 929200.00 | P-132 |  |
| 1849000.00 | P-133 | 763000.00 |
| 400000.00 | P-134 | 0.00 |
| 7943200.00 | P-135 | 0.00 |
| 837200.00 | P-136 | 0.00 |
| 1500000.00 | P-137 | 0.00 |
| 0.00 | P-138 | 1799600.00 |
| 2467200.00 | P-139 | 500000.00 |
| 0.00 | P-140 | 3700000.00 |
| 0.00 | P-141 | 500000.00 |
| 0.00 | P-142 | 1514000.00 |
| 0.00 | P-143 | 714000.00 |
| 267184.00 | P-144 | 0.00 |
| 0.00 | P-145 | 3885200.00 |
| 0.00 | P-147 | 805900.00 |
| 0.00 | P-148 | 700000.00 |
| 0.00 | P-149 | 1979600.00 |
| 0.00 | P-150 | 210000.00 |
| 0.00 | P-151 | 1993200.00 |
| 0.00 | P-153 | 3000000.00 |
| 0.00 | P-155 | 335194.00 |
| 22433000.00 | COE-I | 4000000.00 |
| 8570000.00 | COE-II | 7881000.00 |
| 189431530.00 |  | 58652921.00 |

## CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS FOR THE YEAR ENDED 31st MARCH 2013

Annexure: D Forming part of Receipts \& Payment a/c

| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
| :---: | :---: | :---: |
|  | Advances |  |
| 35000.00 | Computer Advance [Research Fellows] | 100000.00 |
| 131250.00 | Festival Advance | 86250.00 |
| 376415.00 | Others [Advances] | 113479.00 |
| 9667900.00 | General Deposits And Advances | 1038800.00 |
| 2178922.00 | EMD | 241800.00 |
| 9750.00 | Security Deposit | 2500.00 |
| 223500.00 | Revolving Advance | 326500.00 |
| 264000.00 | Advance for purchases by Staff | 229702.00 |
| 538860.00 | Workshop \& Conference | 1698172.00 |
| 129000.00 | LSD \& HSD | 93000.00 |
| 11000.00 | Trainee Security Deposit | 12500.00 |
| 0.00 | GDA [Others] | 135277.00 |
| 1239852.00 | Royalty \& Consultancy | 493382.00 |
| 28472236.00 | Equipment | 12673898.00 |
| 0.00 | Office Equipment | 22700.00 |
| 1118680.00 | LTC [Advance] | 1229250.00 |
| 0.00 | Medical [Advance] | 300000.00 |
| 3785265.00 | TA-India \& Abroad [Advance] | 2876325.00 |
| 0.00 | Honorarium [Advance] | 5000.00 |
| 0.00 | Chemicals [Advance] | 9660410.00 |
| 13039405.00 | Consumables, glassware and Spares [Advance] | 714700.00 |
| 0.00 | AMC for Equipment [Advance] | 38250.00 |
| 0.00 | Other Research Expenses [Advance] | 28090.00 |
| 30000.00 | Computer Advance [Staff] | 210000.00 |
| 60000.00 | Conveyance [Advance] | 148200.00 |
| 64000.00 | Rent [Advance] | 0.00 |
| 916289.00 | NIMS Advance | 0.00 |
| 62291324.00 |  | 32478185.00 |

Annexure: E Forming part of Receipts \& Payment a/c

| Previous Year |  | Particulars |
| ---: | :--- | ---: |
| Amount Rs. | Current Year <br> Amount Rs. |  |
|  | I-Remittances |  |
| 4709062.00 | TDS | 4883964.00 |
| 5622209.00 | Income Tax | 6154006.00 |
| 26093.00 | Works Tax | 4074.00 |
| 1099954.00 | LIC | 1335912.00 |
| 210600.00 | GSLI | 202410.00 |
| 1570900.00 | PPF | 2012875.00 |
| 544966.00 | Professional Tax | 478411.00 |
| 166864.00 | Service Tax | 2155621.00 |
| 0.00 | Others (I-Remittances) | 499520.00 |
| $\mathbf{1 3 9 5 0 6 4 8 . 0 0}$ |  | $\mathbf{1 7 7 2 6 7 9 3 . 0 0}$ |

## CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS

FOR THE YEAR ENDED 31st MARCH 2013
Annexure: F Forming part of Receipts \& Payment a/c

| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
| :---: | :---: | :---: |
|  | Projects - Expenditure |  |
| 29363.00 | P - 49A | 132589.00 |
| 2547094.00 | P-65A | 271200.00 |
| 412681.00 | P-71 | 15829.00 |
| 554228.00 | P-80 | 0.00 |
| 70745.00 | P-81 | 0.00 |
| 1345180.00 | P-81A | 860000.00 |
| 1106732.00 | P-82 | 523580.00 |
| 783120.00 | P-84A | 0.00 |
| 740000.00 | P - 88 | 461182.00 |
| 802987.00 | P-90 | 0.00 |
| 311836.00 | P-91 | 0.00 |
| 3521916.00 | P-92 | 1829794.00 |
| 2832707.00 | P-93 | 2246565.00 |
| 424041.00 | P-95 | 0.00 |
| 796770.00 | P-96 | 0.00 |
| 1118988.00 | P-97 | 48406.00 |
| 1237665.00 | P-98 | 192825.00 |
| 1874442.00 | P-99 | 270880.00 |
| 35000.00 | P-100 | 241590.00 |
| 6430446.00 | P-101 | 9365134.00 |
| 558377.00 | P-102 | 1016456.00 |
| 300000.00 | P-103 | 300000.00 |
| 1692479.00 | P-104 | 2060009.00 |
| 917815.00 | P-105 | 754102.00 |
| 965294.00 | P-106 | 885316.00 |

## CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS FOR THE YEAR ENDED 31st MARCH 2013

Annexure: F Forming part of Receipts \& Payment a/c

| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
| :---: | :---: | :---: |
| 1311406.00 | P-107 | 880165.00 |
| 584498.00 | P-108 | 462890.00 |
| 1902478.00 | P-109 | 787200.00 |
| 193068.00 | P-110 | 22712.00 |
| 1568400.00 | P-111 | 1368315.00 |
| 1803726.00 | P-112 | 0.00 |
| 864774.00 | P-113 | 1571384.00 |
| 1481208.00 | P-114 | 1262412.00 |
| 62019564.00 | P-115 | 8182978.00 |
| 1981237.00 | P-116 | 962946.00 |
| 5251500.00 | P-117 | 0.00 |
| 1115770.00 | P-118 | 0.00 |
| 1298947.00 | P-119 | 1646824.00 |
| 492400.00 | P-120 | 724818.00 |
| 634282.00 | P-121 | 533680.00 |
| 2127215.00 | P-122 | 3269871.00 |
| 810754.00 | P-123 | 1969087.00 |
| 651716.00 | P-124 | 717200.00 |
| 610000.00 | P-125 | 634981.00 |
| 957685.00 | P-126 | 2267043.00 |
| 6044881.00 | P-127 | 5527587.00 |
| 753613.00 | P-128 | 2533016.00 |
| 0.00 | P-129 | 306000.00 |
| 1800000.00 | P-130 | 3721027.00 |
| 716265.00 | P-131 | 1951604.00 |
| 294877.00 | P-132 | 1862803.00 |
| 300000.00 | P-133 | 1342511.00 |
| 146000.00 | P-134 | 395437.00 |
| 525000.00 | P-135 | 2041634.00 |
| 0.00 | P-136 | 759220.00 |
| 0.00 | P-137 | 814980.00 |
| 0.00 | P-138 | 895656.00 |
| 0.00 | P-139 | 1743617.00 |
| 0.00 | P-140 | 1293909.00 |
| 0.00 | P-141 | 498537.00 |
| 0.00 | P-142 | 1153852.00 |
| 0.00 | P-143 | 567716.00 |
| 0.00 | P-144 | 267184.00 |
| 0.00 | P-145 | 1676994.00 |
| 0.00 | P-146 | 1037791.00 |
| 0.00 | P-147 | 490258.00 |
| 0.00 | P-148 | 679674.00 |
| 0.00 | P-149 | 209314.00 |
| 0.00 | P-150 | 45294.00 |
| 12344655.00 | COE-I | 10535012.00 |
| 12948013.00 | COE - II | 11729481.00 |
| 154943838.00 |  | 102820071.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS FOR THE YEAR ENDED 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
| Annexure: G Forming part of Receipts \& Payment a/c |  |  |
| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
|  | CDFD C.P.FACCOUNT |  |
| 33724337.32 | Opening Balance | 29159376.32 |
|  | Add: |  |
| 7891447.00 | Employee subscription/ refunds | 5355840.00 |
| 62280.00 | Transfer from other departments | 0.00 |
| 3323326.00 | Institute contribution (inc. Projects staff) | 2112193.00 |
| 85173.00 | Interest received | 3277120.35 |
| 45086563.32 | Less: | 39904529.67 |
| 15927187.00 | Advances/withdrawals/Transfer/Adjst | 4099128.00 |
| 29159376.32 |  | 35805401.67 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS FOR THE YEAR ENDED 31st MARCH 2013 |  |  |
| :---: | :---: | :---: |
| Annexure: H Forming part of Receipts \& Payment a/c |  |  |
| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
|  | LOANS AND ADVANCES |  |
| 0.00 | TA-India \& Abroad [Advance] | 1069515.56 |
| 0.00 | Honorarium [Advance] | 5000.00 |
| 304569.00 | Rent [Advance] | 304569.00 |
| 69666260.00 | Chemicals [Advance] | 79326670.00 |
| 982164.00 | LTC [Advance] | 1972353.00 |
| 0.00 | Medical [Advance] | 300000.00 |
| 0.00 | Consumables, glassware and Spares [Advance] | 714700.00 |
| 0.00 | AMC for Equipment [Advance] | 38250.00 |
| 0.00 | Other Research Expenses [Advance] | 28090.00 |
| 0.00 | Trainee Security Deposit | 27000.00 |
| 13414314.00 | Research Fellows-Associates | 8468959.00 |
| 68078113.45 | Equipment [Advance] | 37848368.45 |
| 0.00 | Office Equipment [Advance] | 22700.00 |
| 4310.00 | GSLI Recovery | 4310.00 |
| 355710.00 | CDFD STAFF Reserve Fund | 0.00 |
| 37913.00 | Computer Advance [Research Fellows] | 67909.00 |
| 77925.00 | Festival Advance | 53550.00 |
| 5258884.00 | Others [Advances] | 5333288.00 |
| 0.00 | PPF | 85575.00 |
| 72.00 | Service Tax | 176554.00 |
| 83237.00 | Revolving Advance | 102343.00 |
| 248219.50 | Advance for purchases by Staff | 238838.50 |
| 10334109.00 | NIMS Advance | 0.00 |
| 2400288.00 | Workshop \& Conference | 0.00 |
| 0.00 | Conveyance Advance | 44620.00 |
| 0.00 | Computer Advance [Staff] | 88500.00 |
| 171246087.95 |  | 136321662.51 |

## CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS <br> FOR THE YEAR ENDED 31st MARCH 2013

Annexure: I Forming part of Balance Sheet

| Previous Year <br> Amount Rs. |  | Particulars |
| ---: | :--- | ---: |
|  | DEPOSITS | Current Year |
| Amount Rs. |  |  |$|$| 14583375.00 | General Deposits And Advances |
| ---: | ---: |
| 0.00 | GDA[Others] |

CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS FOR THE YEAR ENDED 31st MARCH 2013

Annexure: J Forming part of Balance sheet

| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
| :---: | :---: | :---: |
| $\begin{aligned} & 51098273.00 \\ & 11300000.00 \end{aligned}$ | INVESTMENT A/C <br> Investments Other Investments | $\begin{aligned} & 51098273.00 \\ & 11300000.00 \end{aligned}$ |
| 62398273.00 |  | 62398273.00 |

CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS
FOR THE YEAR ENDED 31st MARCH 2013
Annexure: K Forming part of Balance sheet

| Previous Year Amount Rs. | Particulars | Current Year Amount Rs. |
| :---: | :---: | :---: |
|  | CDFD C.P.F INVESTMENT A/C |  |
| 29159376.32 | Deposit with Banks | 23202519.00 |
| 0.00 | Employee subscription | 7140112.00 |
| 0.00 | Less: Transfer to Bank A/c | $\begin{array}{r} 30342631.00 \\ 5183048.00 \end{array}$ |
| 29159376.32 |  | 25159583.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-03: "Transgenesis and Genetic basis of Pathogen Resistance in the Silkworm, Bombyx Mori P.I: <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| 0.00 | Opening Balance | 0.00 | 630047.00 | Opening Balance | 630047.00 |
| 0.00 | Grant In Aid | 0.00 | 0.00 | Salaries - Manpower | 0.00 |
|  |  |  | 0.00 | Consumables | 0.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 0.00 |  | 0.00 | 630047.00 |  | 630047.00 |
| 630047.00 | Excess of Expenditure over Income | 630047.00 | 0.00 | Closing Balance | 0.00 |
| 630047.00 |  | 630047.00 | 630047.00 |  | 630047.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-09: "NMITLI Project on - Latent M.Tuberculosis: New targets, Drug delivery systems, Bio enhancers \& Therapeutics" <br> P.I: Dr Seyed E Hasnain <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount | Previous Year. <br> Amount <br> Rs | Payments | Current Year Amount |
| 244305.00 | Opening Balance | 244305.00 |  |  |  |
| 0.00 | Grant In Aid | 0.00 | 0.00 | Salaries - Manpower | 0.00 |
|  |  |  | 0.00 | Consumables | 0.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 244305.00 |  | 244305.00 | 0.00 |  | 0.00 |
| 0.00 | Excess of Expenditure over Income | 0.00 | 244305.00 | Closing Balance | 244305.00 |
| 2443050.00 |  | 244305.00 | 244305.00 |  | 244305.00 |


$225$









| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-44: "Understanding of role of Ras and NO / iNOS signalling in promotion of hepatocellular carcinomas with persistent HBV infection" <br> P.I: Dr Gayatri Ramakrishna <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year <br> Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| 0.00 | Opening Balance | 0.00 | 457538.00 | Opening Balance | 457538.00 |
| 0.00 | Grant In Aid | 0.00 | 0.00 | Salaries - Manpower | 0.00 |
|  |  |  | 0.00 | Consumables | 0.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 0.00 |  | 0.00 | 457538.00 |  | 457538.00 |
| 457538.00 | Excess of Expenditure over Income | 457538.00 | 0.00 | Closing Balance | 0.00 |
| 457538.00 |  | 457538.00 | 457538.00 |  | 457538.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-45: Specialized chromatin structures as epigenetic imprints to distinguish parental alleles". <br> P.I: Dr Sanjeev Khosla <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount $\quad$ Rs. | Previous Year. <br> Amount <br> Rs | Payments | Current Year Amount Rs |
| 605714.00 | Opening Balance | 605714.00 |  |  |  |
| 0.00 | Grant In Aid | 0.00 | 0.00 | Salaries - Manpower | 0.00 |
|  |  |  | 0.00 | Consumables | 0.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 605714.00 |  | 605714.00 | 0.00 |  | 0.00 |
| 0.00 | Excess of Expenditure over Income | 0.00 | 605714.00 | Closing Balance | 605714.00 |
| 605714.00 |  | 605714.00 | 605714.00 |  | 605714.00 |




| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-52: "Nucleo Cytoplasmic transport of HIV - 1 Vpr" <br> P.I: Dr Mahalingam \& Dr Manna <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 1231118.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 1231118.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 0.00 \\ 1231118.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 0.00 \\ 1231118.00 \\ \hline \end{array}$ | $\begin{array}{r} 1231118.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 1231118.00 \\ 0.00 \\ \hline \end{array}$ |
| 1231118.00 |  | 1231118.00 | 1231118.00 |  | 1231118.00 |
| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-54: "Study of viability of Mycobacterium leprae in clinical samples and possibility of its presence in the environment using nucleic acid amp techniques." <br> P.I: Dr Niyaz Ahmed <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 37877.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | $\begin{array}{r} \hline 37877.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ \hline \end{array}$ |
| $\begin{array}{r} 0.00 \\ 37877.00 \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 0.00 \\ 37877.00 \end{array}$ | $\begin{array}{r} 37877.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 37877.00 \\ 0.00 \\ \hline \end{array}$ |
| 37877.00 |  | 37877.00 | 37877.00 |  | 37877.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-55: "Identification of DNA Markers for baculovirus resistance in silkworm, Bombyx mori" <br> P.I: Dr J Nagaraju <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} \hline 224.00 \\ 0.00 \end{array}$ | Opening Balance Grant In Aid | $\begin{array}{r} 224.00 \\ 0.00 \end{array}$ | 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ |
| $\begin{array}{r} 224.00 \\ 0.00 \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 224.00 \\ 0.00 \end{array}$ | $\begin{array}{r} 0.00 \\ 224.00 \end{array}$ | Closing Balance | $\begin{array}{r} 0.00 \\ 224.00 \end{array}$ |
| 224.00 |  | 224.00 | 224.00 |  | 224.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-56: "Genetics of transcription-replication interplay and of stress adaptation in bacteria" <br> P.I: Dr Gowrishankar \& Dr K Anupama <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year <br> Amount <br> Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | $\begin{array}{r} 1231164.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ \hline \end{array}$ | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 1231164.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 0.00 \\ 1231164.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 0.00 \\ 1231164.00 \\ \hline \end{array}$ | $\begin{array}{r} 1231164.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 1231164.00 \\ 0.00 \\ \hline \end{array}$ |
| 1231164.00 |  | 1231164.00 | 1231164.00 |  | 1231164.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-59: "An integrated Approach towards understanding the biology of Mycobacterium tuberculosis: Genetic, biochemical, immunological and struc analyses." <br> P.I: Dr Hasnain, Dr Gowrishankar, Dr Mande, Dr Ranjan Sen <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| 0.00 | Opening Balance | 0.00 | 2215024.00 | Opening Balance | 2215024.00 |
| 0.00 | Grant In Aid | 0.00 | 0.00 | Salaries - Manpower | 0.00 |
|  |  |  | 0.00 | Consumables | 0.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 0.00 |  | 0.00 | 2215024.00 |  | 2215024.00 |
| 2215024.00 | Excess of Expenditure over Income | 2215024.00 | 0.00 | Closing Balance | 0.00 |
| 2215024.00 |  | 2215024.00 | 2215024.00 |  | 2215024.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-60: "National Database of Prevalent Genetic Disorders in India: Development, Curation and Services" <br> P.I: Dr H A Nagarajaram <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount $\qquad$ | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| 482124.00 | Opening Balance | 482124.00 |  |  |  |
| 0.00 | Grant In Aid | 0.00 | 0.00 | Salaries - Manpower | 0.00 |
|  |  |  | 0.00 | Consumables | 0.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 482124.00 |  | 482124.00 | 0.00 |  | 0.00 |
| 0.00 | Excess of Expenditure over Income | 0.00 | 482124.00 | Closing Balance | 482124.00 |
| 482124.00 |  | 482124.00 | 482124.00 |  | 482124.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-61: "Dissection of a novel phenotype of lethal accumulation of potassium in Escherichia coli mutants defective in thioredoxin/thioredoxin reductase <br> nucleoied protein H-NS" <br> P.I: Dr Abhijit A Sardesai <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year | Receipts | Current Year | Previous Year. | Payments | Current Year |
| Amount Rs |  | Amount Rs. | Amount Rs |  | Amount Rs |
| 0.00 | Opening Balance Grant In Aid | 0.00 | 280000.00 | Opening Balance | 280000.00 |
| 0.00 |  | 0.00 | 0.00 | Salaries - Manpower | 0.00 |
|  |  |  | 0.00 | Consumables | 0.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 0.00 | Excess of Expenditure over Income | 0.00 | 280000.00 |  | 280000.00 |
| 280000.00 |  | 280000.00 | 0.00 | Closing Balance | 0.00 |
| 280000.00 |  | 280000.00 | 280000.00 |  | 280000.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-62: "HIV - 1 Pathogenesis: Role of Integrase in Reverse Transciption and Nuclear Transport of Viral Genome" <br> P.I: Dr S Mahalingam <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 278928.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 278928.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 0.00 \\ 278928.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} \hline 0.00 \\ 278928.00 \end{array}$ | $\begin{array}{r} \hline 278928.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 278928.00 \\ 0.00 \end{array}$ |
| 278928.00 |  | 278928.00 | 278928.00 |  | 278928.00 |



| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-65: "Molecular, genetic and functional analysis of the chromosomal plasticity region of the gastric pathogen Helicobater pylori" <br> P.I: Dr Ayesha Alvi <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 582647.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 582647.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 0.00 \\ 582647.00 \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 0.00 \\ 582647.00 \end{array}$ | $\begin{array}{r} 582647.00 \\ 0.00 \end{array}$ | Closing Balance | $\begin{array}{r} 582647.00 \\ 0.00 \end{array}$ |
| 582647.00 |  | 582647.00 | 582647.00 |  | 582647.00 |
| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-65A: APEDA-CDFD Centre for Basmati DNA Analysis <br> P.I: Dr J Nagaraju <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} 16381715.00 \\ 0.00 \\ 1103400.00 \\ 4000000.00 \end{array}$ | Opening Balance <br> Grant In Aid <br> Basmati Analysis Charges <br> AMC Amount Received | $\begin{array}{r} 18938021.00 \\ 0.00 \\ 1068000.00 \\ 0.00 \end{array}$ | 0.00 543948.00 2000000.00 0.00 0.00 0.00 0.00 0.00 3146.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Consultancy \& Knowledge Fee <br> Vehicle <br> Equipment | 0.00 271200.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| 21485115.00 | Excess of expenditure over income | 20006021.00 | $\begin{array}{r} 2547094.00 \\ 18938021.00 \end{array}$ | Closing Balance | $\begin{array}{r} 271200.00 \\ 19734821.00 \end{array}$ |
| 21485115.00 |  | 20006021.00 | 21485115.00 |  | 20006021.00 |

$243$





| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-78: Task force- IMD Newborn screening for Congenital Hypothyroidism \& Congenital Adrenal Hyperplasia: A multicentric study <br> P.I: Dr A Radha Rama Devi <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} 1304.00 \\ 0.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 1304.00 \\ 0.00 \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 1304.00 \\ 0.00 \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 1304.00 \\ 0.00 \end{array}$ | $\begin{array}{r} 0.00 \\ 1304.00 \end{array}$ | Closing Balance | $\begin{array}{r} 0.00 \\ 1304.00 \end{array}$ |
| 1304.00 |  | 1304.00 | 1304.00 |  | 1304.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-79: Understanding the role of AGE proteins in inducing inflammatory responses and its regulation <br> P.I: Dr S K Manna <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} 0.00 \\ 1795900.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 1900986.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 105086.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 1795900.00 \\ 105086.00 \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} \mathbf{0 . 0 0} \\ 105086.00 \end{array}$ | $\begin{array}{r} 1900986.00 \\ 0.00 \end{array}$ | Closing Balance | $\begin{array}{r} 105086.00 \\ 0.00 \end{array}$ |
| 1900986.00 |  | 105086.00 | 1900986.00 |  | 105086.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-80: Referral centre for detection of genetically modified foods employing DNA-based markets <br> P.I: Dr Madhusudan Reddy <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 53994.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 554228.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 608222.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 0.00 \\ 608222.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 0.00 \\ 608222.00 \\ \hline \end{array}$ | $\begin{array}{r} 608222.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 608222.00 \\ 0.00 \end{array}$ |
| 608222.00 |  | 608222.00 | 608222.00 |  | 608222.00 |
|  |  |  |  |  |  |
| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-81: Reconstructing Cellular Networks: Two-component regulatory systems <br> P.I: Dr Shekhar Mande <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} 214215.00 \\ 0.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 143470.00 \\ 0.00 \end{array}$ | $\begin{array}{r} 70745.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ \hline \end{array}$ | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ |
| $\begin{array}{r} 214215.00 \\ 0.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 143470.00 \\ 0.00 \\ \hline \end{array}$ | $\begin{array}{r} 70745.00 \\ 143470.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 0.00 \\ 143470.00 \\ \hline \end{array}$ |
| 214215.00 |  | 143470.00 | 214215.00 |  | 143470.00 |








| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-95: Construction of regulatory networks in prokaryotes through protein: Protein interaction predictions and transcription regulation predictions. with Russian Foundation) <br> P.I: Dr Shekar C Mande |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Curren Amount | $\begin{aligned} & \text { ear } \\ & \text { Rs } \end{aligned}$ |
| $\begin{array}{r} 424041.00 \\ 0.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 424041.00 | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds |  | 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 424041.00 \\ 0.00 \end{array}$ | Excess of Expenditure over Income | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | $\begin{array}{r} 424041.00 \\ 0.00 \end{array}$ | Closing Balance |  | 0.00 0.00 |
| 424041.00 |  | 0.00 | 424041.00 |  |  | 0.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-96: Molecular Characterization of sporadic colorectal cancer in the young from India <br> P.I: Dr M D Bashyam <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Curren Amount | $\begin{aligned} & \text { ear } \\ & \text { Rs } \end{aligned}$ |
| $\begin{array}{r} 0.00 \\ 1452712.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 655942.00 26700.00 713546.00 0.00 56524.00 0.00 0.00 0.00 0.00 0.00 0.00 | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds |  | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ |
| $\begin{array}{r} 1452712.00 \\ 0.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | $\begin{array}{r} 1452712.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance |  | $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ |
| 1452712.00 |  | 0.00 | 1452712.00 |  |  | 0.00 |




| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-101: Role of inositol pyrophosphates in cell physiology: Investigating the biochemical significance of protein pyrophosphorylation - Senior Fello <br> P.I: Dr Rashna Bhandari <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount | Receipts | $\begin{gathered} \text { Current Year } \\ \text { Amount } \quad \text { Rs. } \end{gathered}$ | Previous Year. Amount Rs | Payments | $\begin{gathered} \text { Current Year } \\ \text { Amount } \quad \text { Rs } \end{gathered}$ |
| 15688931.00 | Opening Balance Grant In Aid | 13729401.00 |  |  |  |
| 4470916.00 |  | 0.00 | 1787120.00 | Salaries - Manpower | 1818930.00 |
|  |  |  | 1100000.00 | Consumables | 2300000.00 |
|  |  |  | 0.00 | Contingencies | 849721.00 |
|  |  |  | 73814.00 | Travel | 27452.00 |
|  |  |  | 584586.00 | Overheads | 0.00 |
|  |  |  | 2884926.00 | Equipment | 4191222.00 |
|  |  |  | 0.00 | Books | 177809.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 20159847.00 | Excess of Expenditure over Income | 13729401.00 | 6430446.00 |  | 9365134.00 |
| 0.00 |  | 0.00 | 13729401.00 | Closing Balance | 4364267.00 |
| 20159847.00 |  | 13729401.00 | 20159847.00 |  | 13729401.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-102: Understanding the role of Mycobacterium tuberculosis heat shockprotein 60 as Th1/Th2 immuno modular <br> P.I: Dr Sangita Mukhopadhyay <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| $\begin{aligned} & \hline \text { Previous } \text { Year } \\ & \text { Amount } \quad \text { Rs } \end{aligned}$ | Receipts | $\begin{gathered} \text { Current Year } \\ \text { Amount } \quad \text { Rs. } \end{gathered}$ | Previous Year. Amount Rs | Payments | Current Year Amount |
| 0.00 | Opening Balance | 82654.00 | 445133.00 |  |  |
| 1086164.00 | Grant In Aid | 503782.00 | 289800.00 | Salaries - Manpower | 386400.00 |
|  |  |  | 200000.00 | Consumables | 600000.00 |
|  |  |  | 0.00 | Contingencies | 29605.00 |
|  |  |  | 20627.00 | Travel | 451.00 |
|  |  |  | 25000.00 | Overheads | 0.00 |
|  |  |  | 22950.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 1086164.00 |  | 586436.00 | 1003510.00 |  | 1016456.00 |
| 0.00 | Excess of Expenditure over Income | 430020.00 | 82654.00 | Closing Balance | 0.00 |
| 1086164.00 |  | 1016456.00 | 1086164.00 |  | 1016456.00 |



| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-105: Cloning, Characterization and analysis of chromosomal rearrangements in human genetic disorders <br> P.I: Dr Ashwin B Dalal <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount $\quad$ Rs | Receipts | Current Year Amount | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| 145971.00 | Opening Balance | 0.00 |  | Opening Balance | 90844.00 |
| 681000.00 | Grant In Aid | 0.00 | 187200.00 | Salaries - Manpower | 126202.00 |
|  |  |  | 700000.00 | Consumables | 600000.00 |
|  |  |  | 20000.00 | Contingencies | 20000.00 |
|  |  |  | 10615.00 | Travel | 7900.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 826971.00 |  | 0.00 | 917815.00 |  | 844946.00 |
| 90844.00 | Excess of Expenditure over Income | 844946.00 | 0.00 | Closing Balance | 0.00 |
| 917815.00 |  | 844946.00 | 917815.00 |  | 844946.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-106: Clinical, Biochemical and molecular analysis of treatable lysosomal storage disorders <br> P.I: Dr Ashwin B Dalal <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year <br> Amount Rs | Receipts | Current Year <br> Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| 0.00 | Opening Balance | 190952.00 | 446056.00 |  |  |
| 1602302.00 | Grant In Aid | 505153.00 | 484977.00 | Salaries - Manpower | 409530.00 |
|  |  |  | 450000.00 | Consumables | 450000.00 |
|  |  |  | 30317.00 | Contingencies | 25786.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 1602302.00 |  | 696105.00 | 1411350.00 |  | 885316.00 |
| 0.00 | Excess of Expenditure over Income | 189211.00 | 190952.00 | Closing Balance | 0.00 |
| 1602302.00 |  | 885316.00 | 1602302.00 |  | 885316.00 |


CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD
P-110: India-Japan research project title"Identification and analysis of sex determining

| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-110: India-Japan research project title"Identification and analysis of sex determining genes in silkmoths" <br> P.I: Dr J Nagaraju <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} 24389.00 \\ 0.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 0.00 0.00 0.00 193068.00 0.00 0.00 0.00 0.00 0.00 0.00 | Opening Balance <br> Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 168679.00 0.00 0.00 0.00 22712.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 24389.00 \\ 168679.00 \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 0.00 \\ 191391.00 \\ \hline \end{array}$ | $\begin{array}{r} 193068.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 191391.00 \\ 0.00 \\ \hline \end{array}$ |
| 193068.00 |  | 191391.00 | 193068.00 |  | 191391.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-111: Ramalingaswami Fellowship - Refractoriness mechanism in Mosquito: cracking molecular codes at genomic scale P.I: Dr Shweta Tyagi <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} 488631.00 \\ 1511500.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 431731.00 \\ 1487000.00 \end{array}$ | $\begin{array}{r} 1168400.00 \\ 400000.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ \hline \end{array}$ | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | $\begin{array}{r} 1106000.00 \\ 262315.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ 0.00 \\ \hline \end{array}$ |
| $\begin{array}{r} 2000131.00 \\ 0.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 1918731.00 \\ 0.00 \\ \hline \end{array}$ | $\begin{array}{r} 1568400.00 \\ 431731.00 \\ \hline \end{array}$ | Closing Balance | $\begin{array}{r} 1368315.00 \\ 550416.00 \end{array}$ |
| 2000131.00 |  | 1918731.00 | 2000131.00 |  | 1918731.00 |
| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-112: Ramanujan Fellowship <br> P.I: Dr Rohit Joshi <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} 803726.00 \\ 1000000.00 \end{array}$ | Opening Balance Grant In Aid | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | 626400.00 157326.00 0.00 0.00 20000.00 0.00 0.00 0.00 0.00 1000000.00 | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ |
| $\begin{array}{r} 1803726.00 \\ 0.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | $\begin{array}{r} 1803726.00 \\ 0.00 \\ \hline \end{array}$ | Closing Balance | $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ |
| 1803726.00 |  | 0.00 | 1803726.00 |  | 0.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-113: Clinical and molecular genetic analysis of squamous cell carcinoma of the tongue P.I: Dr M D Bashyam <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 550715.00 \\ & 848689.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 534630.00 \\ 0.00 \end{array}$ | 519587.00 <br> 320000.00 <br> 0.00 <br> 0.00 <br> 25187.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 <br> 0.00 | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 544619.00 590000.00 34084.00 1500.00 0.00 401181.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 1399404.00 \\ 0.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 534630.00 \\ 1036754.00 \end{array}$ | $\begin{aligned} & 864774.00 \\ & 534630.00 \\ & \hline \end{aligned}$ | Closing Balance | $\begin{array}{r} 1571384.00 \\ 0.00 \\ \hline \end{array}$ |
| 1399404.00 |  | 1571384.00 | 1399404.00 |  | 1571384.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-117: Joint New Indigo Era-Net project titled "Mycobacterium Tuberculosis:bioinformatic and structural strategies towards treatment <br> P.I: Dr Shekhar C Mande <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year |  |
| 5251500.00 | Opening Balance | 0.00 |  |  |  |  |
| 0.00 | Grant In Aid | 0.00 | 138667.00 | Salaries - Manpower |  | 0.00 |
|  |  |  | 300000.00 | Consumables |  | 0.00 |
|  |  |  | 25000.00 | Contingencies |  | 0.00 |
|  |  |  | 140754.00 | Travel |  | 0.00 |
|  |  |  | 50000.00 | Overheads |  | 0.00 |
|  |  |  | 0.00 | Equipment |  | 0.00 |
|  |  |  | 0.00 | Books |  | 0.00 |
|  |  |  | 0.00 | AMC |  | 0.00 |
|  |  |  | 0.00 | Others |  | 0.00 |
|  |  |  | 4597079.00 | Transfer of Funds |  | 0.00 |
| 5251500.00 |  | 0.00 | 5251500.00 |  |  | 0.00 |
| 0.00 | Excess of Expenditure over Income | 0.00 | 0.00 | Closing Balance |  | 0.00 |
| 5251500.00 |  | 0.00 | 5251500.00 |  |  | 0.00 |





| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-125: Mechanistic studies on the role of protein kinase Snfilk in cell cycle and cancer <br> P.I: $\operatorname{Dr}$ M Subba Reddy <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year | Previous Year. Amount Rs | Payments | Current Year |
| 0.00 | Opening Balance | 154000.00 |  |  |  |
| 764000.00 | Grant In Aid | 0.00 | 110000.00 | Salaries - Manpower | 134981.00 |
|  |  |  | 500000.00 | Consumables | 500000.00 |
|  |  |  | 0.00 | Contingencies | 0.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transter of Funds | 0.00 |
| 0.00 | Excess of Expenditure over Income | 480981.00 | 154000.00 | Closing Balance | 0300 |
| 764000.00 |  | 634981.00 | 764000.00 |  | 634981.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-126: Rho-dependent transcription termination machinery: mechanism of action P.I: Dr Ranjan Sen Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| $\begin{aligned} & \hline \text { Previous Year } \\ & \text { Amount } \quad \text { Rs } \\ & \hline \end{aligned}$ | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year <br> Amount Rs |
| 0.00 | Opening Balance | 1581615.00 |  |  |  |
| 2539300.00 | Grant In Aid | 0.00 | 111548.00 | Salaries - Manpower | 488532.00 |
|  |  |  | 800000.00 | Consumables | 1381445.00 |
|  |  |  | 30000.00 | Contingencies | 3081.00 |
|  |  |  | 16137.00 | Travel | 8581.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 385404.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 2539300.00 0.00 | Excess of Expenditure over Income | 1581615.00 <br> 685428.00 | 957685.00 <br> 1581615.00 | Closing Balance | 2267043.00 0.00 |
| 2539300.00 |  | 2267043.00 | 2539300.00 |  | 2267043.00 |







| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-137: Signaling pathways involved in down regulation of proinflammatory responses by PPE18 protein of Mycobacterium tuberculosis: Implicatio <br> PPE18 as therapeutics <br> P.I: Dr Sangita Mukhopadhyay <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year <br> Amount Rs | Receipts | $\begin{gathered} \text { Current Year } \\ \text { Amount } \quad \text { Rs. } \end{gathered}$ | Previous Year. Amount | Payments | Current Year <br> Amount Rs |
| $\begin{array}{r} 0.00 \\ 1500000.00 \end{array}$ | Opening Balance | 1500000.00 |  |  |  |
|  | Grant In Aid | 0.00 | 0.00 | Salaries - Manpower | 99840.00 |
|  |  |  | 0.00 | Consumables | 603140.00 |
|  |  |  | 0.00 | Contingencies | 112000.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| $\begin{array}{r} 1500000.00 \\ 0.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | 1500000.00 | $0.00$ |  | 814980.00 |
|  | Excess of Expenditure over Income |  |  | Closing Balance |  |
| 1500000.00 |  | 1500000.00 | 1500000.00 |  | 1500000.00 |
|  |  |  |  |  |  |
| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-138: Co-evaluation of Dnmt3I and Genomic imprinting <br> P.I: Dr Sanjeev Khosla <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year <br> Amount Rs |
| 0.00 | Opening Balance | 0.00 |  |  |  |
| 0.00 | Grant In Aid | 1799600.00 | 0.00 | Salaries - Manpower | 70656.00 |
|  |  |  | 0.00 | Consumables | 800000.00 |
|  |  |  | 0.00 | Contingencies | 25000.00 |
|  |  |  | 0.00 | Travel | 0.00 |
|  |  |  | 0.00 | Overheads | 0.00 |
|  |  |  | 0.00 | Equipment | 0.00 |
|  |  |  | 0.00 | Books | 0.00 |
|  |  |  | 0.00 | AMC | 0.00 |
|  |  |  | 0.00 | Others | 0.00 |
|  |  |  | 0.00 | Transfer of Funds | 0.00 |
| 0.00 |  | 1799600.00 | 0.00 |  | 895656.00 |
| 0.00 | Excess of Expenditure over Income | 0.00 | 0.00 | Closing Balance | 903944.00 |
| 0.00 |  | 1799600.00 | 0.00 |  | 1799600.00 |


| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-139: Evaluating the role of Sirtuins and epigenetic changes during cellular senescense in context of p53 status <br> P.I: Dr Gayatri Ramakrishna, Dr Sanjeev Khosla <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{array}{r} \hline 0.00 \\ 2467200.00 \end{array}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 2467200.00 \\ 500000.00 \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 80617.00 1623000.00 40000.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{array}{r} 2467200.00 \\ 0.00 \\ \hline \end{array}$ | Excess of Expenditure over Income | $\begin{array}{r} 2967200.00 \\ 0.00 \\ \hline \end{array}$ | $\begin{array}{r} 0.00 \\ 2467200.00 \\ \hline \end{array}$ | Closing Balance | $\begin{aligned} & 1743617.00 \\ & 1223583.00 \\ & \hline \end{aligned}$ |
| 2467200.00 |  | 2967200.00 | 2467200.00 |  | 2967200.00 |
| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-140: Development of baculovirus resistant silkworms strains through synthetic miRNA based knockdown of essential viral genes <br> P.I: Dr K P Arun Kumar <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
|  |  |  |  |  |  |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 0.00 \\ 1850000.00 \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 193909.00 1084288.00 0.00 15712.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Excess of Expenditure over Income | $\begin{array}{r} 1850000.00 \\ 0.00 \\ \hline \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | Closing Balance | $\begin{array}{r} 1293909.00 \\ 556091.00 \\ \hline \end{array}$ |
| 0.00 |  | 1850000.00 | 0.00 |  | 1850000.00 |




| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-145: H3K4 HMT family regulatescell cycle progression P.I: Dr Shweta Tyagi <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 0.00 \\ 3885200.00 \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 76994.00 1600000.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Excess of Expenditure over Income | $\begin{array}{r} 3885200.00 \\ 0.00 \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Closing Balance | $\begin{aligned} & 1676994.00 \\ & 2208206.00 \end{aligned}$ |
| 0.00 |  | 3885200.00 | 0.00 |  | 3885200.00 |
|  |  |  |  |  |  |
| CENTRE FOR DNA FINGERPRINTING AND DIAGNOSTICS, HYDERABAD <br> P-146: Role of MLL in ribosomal RNA transcription <br> P.I: Dr Shweta Tyagi <br> Receipts and Payments Account from 01/04/2012 to 31/03/2013 |  |  |  |  |  |
| Previous Year Amount Rs | Receipts | Current Year Amount Rs. | Previous Year. Amount Rs | Payments | Current Year Amount $\quad$ Rs |
| $\begin{aligned} & 0.00 \\ & 0.00 \end{aligned}$ | Opening Balance <br> Grant In Aid | $\begin{array}{r} 0.00 \\ 1850000.00 \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \\ & 0.00 \end{aligned}$ | Salaries - Manpower <br> Consumables <br> Contingencies <br> Travel <br> Overheads <br> Equipment <br> Books <br> AMC <br> Others <br> Transfer of Funds | 78080.00 600000.00 0.00 0.00 0.00 359711.00 0.00 0.00 0.00 0.00 |
| $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | Excess of Expenditure over Income | $\begin{array}{r} 1850000.00 \\ 0.00 \\ \hline \end{array}$ | $\begin{aligned} & 0.00 \\ & 0.00 \\ & \hline \end{aligned}$ | Closing Balance | $\begin{array}{r} 1037791.00 \\ 812209.00 \\ \hline \end{array}$ |
| 0.00 |  | 1850000.00 | 0.00 |  | 1850000.00 |






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फोटो गैलरी
Photo Gallery


Exposure visit for in-service biology teachers of Kendriya Vidyalaya Sangathan on 17 May 2012


Fire drill in the CDFD Administration Block on 30 May 2012


Signing of the MoU with the Government of Andhra Pradesh on 11 July 2012 to promote partnership in the area of DNA fingerprinting examination and training.


Flag hoisting on the occasion of Independence Day 2012


Celebration of Hindi Day on 14 September 2012


Lecture by Prof. Aravinda Chakravarti from the Johns Hopkins University School of Medicine, Baltimore, USA during the Mini-Symposium on 13 October 2012


Participants of the Seminar Workshop on Microbial Biology organized from
11-14 December 2012


Lecture by Prof. Ajit Varki, University of California, San Diego, USA
on 5 February 2013


Glimpses of CDFD Foundation Day-2013


Dr. J. Gowrishankar, Director, CDFD being conferred with the Padma Shri Award-2013 by Hon'ble President of India Shri Pranab Mukherjee.

